

2'-BRANCHED NUCLEOSIDES AND *FLAVIVIRIDAE* MUTATION

CROSS-REFERENCE

This application claims priority to U.S. Provisional Application No. 60/426,675, filed on November 15, 2002, the disclosure of which is herein incorporated by reference.

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FIELD OF THE INVENTION

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This invention is a method for the treatment of *Flaviviridae* infection in a host, such as a human, in need of such therapy, that includes the administration of a 2'-branched nucleoside, or a pharmaceutically acceptable salt, ester, or prodrug thereof, in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that are associated with such a mutation. The invention also includes a method for the treatment of *Flaviviridae* infection in a host, such as a human, in need of such therapy, that includes the administration of a 2'-branched nucleoside, or a pharmaceutically acceptable salt, ester, or prodrug thereof, in combination and/or alternation with interferon. The invention also includes a method to detect a mutant strain of *Flaviviridae* and a method for its treatment, and kits and materials for such detection.

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BACKGROUND OF THE INVENTION

The *Flaviviridae* family of viruses comprises at least three distinct genera: *pestiviruses*, which cause disease in cattle and pigs; *flaviviruses*, which are the primary cause of diseases such as dengue fever and yellow fever; and *hepaciviruses*, whose sole member is HCV. The flavivirus genus includes more than 68 members separated into

groups on the basis of serological relatedness (Calisher et al., *J. Gen. Virol*, **1993**, 70, 37-43). Clinical symptoms vary and include fever, encephalitis and hemorrhagic fever (*Fields Virology*, Editors: Fields, B. N., Knipe, D. M., and Howley, P. M., Lippincott-Raven Publishers, Philadelphia, PA, **1996**, Chapter 31, 931-959). Flaviviruses of global concern that are associated with human disease include the dengue hemorrhagic fever viruses (DHF), yellow fever virus, West Nile virus, shock syndrome and Japanese encephalitis virus (Halstead, S. B., *Rev. Infect. Dis.*, **1984**, 6, 251-264; Halstead, S. B., *Science*, 239:476-481, 1988; Monath, T. P., *New Eng. J. Med.*, **1988**, 319, 641-643).

The pestivirus genus includes bovine viral diarrhea virus (BVDV), classical swine fever virus (CSFV, also called hog cholera virus) and border disease virus (BDV) of sheep (Moennig, V. et al. *Adv. Vir. Res.* **1992**, 41, 53-98). Pestivirus infections of domesticated livestock (cattle, pigs and sheep) cause significant economic losses worldwide. BVDV causes mucosal disease in cattle and is of significant economic importance to the livestock industry (Meyers, G. and Thiel, H.-J., *Advances in Virus Research*, **1996**, 47, 53-118; Moennig V., et al, *Adv. Vir. Res.* **1992**, 41, 53-98). Human pestiviruses have not been as extensively characterized as the animal pestiviruses. However, serological surveys indicate considerable pestivirus exposure in humans.

Pestiviruses and *hepaciviruses* are closely related virus groups within the *Flaviviridae* family. Other closely related viruses in this family include the GB virus A, GB virus A-like agents, GB virus-B and GB virus-C (also called hepatitis G virus, HGV). The hepacivirus group (hepatitis C virus; HCV) consists of a number of closely related but genotypically distinguishable viruses that infect humans. There are approximately 6 HCV genotypes and more than 50 subtypes. HCV is a major cause of hepatitis globally. Most HCV infections become persistent and about 75% of cases develop chronic liver disease. Chronic HCV infection can lead to development of cirrhosis, hepatocellular carcinoma and liver failure. Due to the similarities between pestiviruses and hepaciviruses, combined with the poor ability of hepaciviruses to grow efficiently in cell culture, bovine viral diarrhea virus (BVDV) is often used as a surrogate to study the HCV virus.

The genetic organization of pestiviruses and hepaciviruses is very similar. These positive stranded RNA viruses possess a single large open reading frame (ORF)

encoding all the viral proteins necessary for virus replication. These proteins are expressed as a polyprotein that is co- and post-translationally processed by both cellular and virus-encoded proteinases to yield the mature viral proteins. The viral proteins responsible for the replication of the viral genome RNA are located within approximately the carboxy-terminal two-thirds of the ORF and are termed nonstructural (NS) proteins. The genetic organization and polyprotein processing of the nonstructural protein portion of the ORF for pestiviruses and hepaciviruses is very similar. For both the pestiviruses and hepaciviruses, the mature nonstructural (NS) proteins, in sequential order from the amino-terminus of the nonstructural protein coding region to the carboxy-terminus of the ORF, consist of p7, NS2, NS3, NS4A, NS4B, NS5A, and NS5B.

The NS proteins of pestiviruses and hepaciviruses share sequence domains that are characteristic of specific protein functions. For example, the NS3 proteins of viruses in both groups possess amino acid sequence motifs characteristic of serine proteinases and of helicases (Gorbalenya et al. (1988) *Nature* 333:22; Bazan and Fletterick (1989) *Virology* 171:637-639; Gorbalenya et al. (1989) *Nucleic Acid Res.* 17:3889-3897). Similarly, the NS5B proteins of pestiviruses and hepaciviruses have the motifs characteristic of RNA-directed RNA polymerases (Koonin, E.V. and Dolja, V.V. (1993) *Crit. Rev. Biochem. Molec. Biol.* 28:375-430).

Furthermore, the actual roles and functions of the NS proteins of pestiviruses and hepaciviruses in the lifecycle of the viruses are directly analogous. In both cases, the NS3 serine proteinase is responsible for all proteolytic processing of polyprotein precursors downstream of its position in the ORF (Wiskerchen and Collett (1991) *Virology* 184:341-350; Bartenschlager et al. (1993) *J. Virol.* 67:3835-3844; Eckart et al. (1993) *Biochem. Biophys. Res. Comm.* 192:399-406; Grakoui et al. (1993) *J. Virol.* 67:2832-2843; Grakoui et al. (1993) *Proc. Natl. Acad. Sci. USA* 90:10583-10587; Hijikata et al. (1993) *J. Virol.* 67:4665-4675; Tome et al. (1993) *J. Virol.* 67:4017-4026). The NS4A protein, in both cases, acts as a cofactor with the NS3 serine protease (Bartenschlager et al. (1994) *J. Virol.* 68:5045-5055; Failla et al. (1994) *J. Virol.* 68:3753-3760; Lin et al. (1994) 68:8147-8157; Xu et al. (1997) *J. Virol.* 71:5312-5322). The NS3 protein of both viruses also functions as a helicase (Kim et al. (1995) *Biochem. Biophys. Res. Comm.* 215: 160-166; Jin and Peterson (1995) *Arch. Biochem. Biophys.*,

323:47-53; Warrener and Collett (1995) J. Virol. 69:1720-1726). Finally, the NS5B proteins of pestiviruses and hepaciviruses have the predicted RNA-directed RNA polymerases activity (Behrens et al.(1996) EMBO J. 15:12-22; Lchmannet al.(1997) J. Virol. 71:8416-8428; Yuan et al.(1997) Biochem. Biophys. Res. Comm. 232:231-235; Hagedorn, PCT WO 97/12033; Zhong et al.(1998) J. Virol. 72:9365-9369).

Examples of antiviral agents that have been identified as active against the *Flaviviridae* family of viruses include:

(1) Interferon

Interferons (IFNs) are compounds that have been commercially available for the treatment of chronic hepatitis for nearly a decade. IFNs are glycoproteins produced by immune cells in response to viral infection. IFNs inhibit viral replication of many viruses, including HCV, and when used as the sole treatment for hepatitis C infection, IFN suppresses serum HCV-RNA to undetectable levels. Additionally, IFN normalizes serum amino transferase levels. Unfortunately, the effects of IFN are temporary and a sustained response occurs in only 8%-9% of patients chronically infected with HCV (Gary L. Davis. *Gastroenterology* 118:S104-S114, 2000).

A number of patents disclose HCV treatments using interferon-based therapies. For example, U.S. Patent No. 5,980,884 to Blatt *et al.* discloses methods for retreatment of patients afflicted with HCV using consensus interferon. U.S. Patent No. 5,942,223 to Bazer *et al.* discloses an anti-HCV therapy using ovine or bovine interferon-tau. U.S. Patent No. 5,928,636 to Alber *et al.* discloses the combination therapy of interleukin-12 and interferon alpha for the treatment of infectious diseases including HCV. U.S. Patent No. 5,908,621 to Glue *et al.* discloses the use of polyethylene glycol modified interferon for the treatment of HCV. U.S. Patent No. 5,849,696 to Chretien *et al.* discloses the use of thymosins, alone or in combination with interferon, for treating HCV. U.S. Patent No. 5,830,455 to Valtuena *et al.* discloses a combination HCV therapy employing interferon and a free radical scavenger. U.S. Patent No. 5,738,845 to Imakawa discloses the use of human interferon tau proteins for treating HCV. Other interferon-based treatments for HCV are disclosed in U.S. Patent No. 5,676,942 to Testa *et al.*, U.S. Patent No. 5,372,808 to Blatt *et al.*, and U.S. Patent No. 5,849,696.

(2) Ribavirin (Battaglia, A.M. *et al.*, *Ann. Pharmacother*, 2000, 34, 487-494); Berenguer, M. *et al. Antivir. Ther.*, 1998, 3 (Suppl. 3), 125-136).

Ribavirin (1- β -D-ribofuranosyl-1-1,2,4-triazole-3-carboxamide) is a synthetic, non-interferon-inducing, broad spectrum antiviral nucleoside analog. It is sold under the trade names VirazoleTM (The Merck Index, 11th edition, Editor: Budavari, S., Merck & Co., Inc., Rahway, NJ, p1304, 1989); Rebetol (Schering Plough) and Co-Pegasus (Roche). United States Patent No. 3,798,209 and RE29,835 (ICN Pharmaceuticals) disclose and claim ribavirin. Ribavirin is structurally similar to guanosine, and has in vitro activity against several DNA and RNA viruses including *Flaviviridae* (Gary L. Davis. *Gastroenterology* 118:S104-S114, 2000). U.S. Patent No 4,211,771 (to ICN Pharmaceuticals) discloses the use of ribavirin as an antiviral agent. Ribavirin reduces serum amino transferase levels to normal in 40% of patients, but it does not lower serum levels of HCV-RNA (Gary L. Davis. *Gastroenterology* 118:S104-S114, 2000). Thus, ribavirin alone is not effective in reducing viral RNA levels. Additionally, ribavirin has significant toxicity and is known to induce anemia.

Combination of Interferon and Ribavirin

Schering-Plough sells ribavirin as Rebetol® capsules (200 mg) for administration to patients with HCV. The U.S. FDA has approved Rebetol capsules to treat chronic HCV infection in combination with Schering's alpha interferon-2b products Intron® A and PEG-IntronTM. Rebetol capsules are *not* approved for monotherapy (*i.e.*, administration independent of Intron®A or PEG-Intron), although Intron A and PEG-Intron are approved for monotherapy (*i.e.*, administration without ribavirin). Hoffman La Roche is selling ribavirin under the name Co-Pegasus in Europe and the United States, also for use in combination with interferon for the treatment of HCV. Other alpha interferon products include Roferon-A (Hoffmann-La Roche), Infergen® (Intermune, formerly Amgen's product), and Wellferon® (Wellcome Foundation) are currently FDA-approved for HCV monotherapy. Interferon products currently in development for HCV include: Roferon-A (interferon alfa-2a) by Roche, PEGASYS (pegylated interferon alfa-2a) by Roche, INFERGEN (interferon alfacon-1) by InterMune, OMNIFERON (natural interferon) by Viragen, ALBUFERON by Human Genome Sciences, REBIF (interferon beta-1a) by Ares-Serono, Omega Interferon by

BioMedicine, Oral Interferon Alpha by Amarillo Biosciences, and Interferon gamma-1b by InterMune.

The combination of IFN and ribavirin for the treatment of HCV infection has been reported to be effective in the treatment of IFN naïve patients (Battaglia, A.M. *et al.*, *Ann. Pharmacother.* 34:487-494, 2000). Combination treatment is effective both before hepatitis develops and when histological disease is present (Berenguer, M. *et al.* *Antivir. Ther.* 3(Suppl. 3):125-136, 1998). Currently, the most effective therapy for HCV is combination therapy of pegylated interferon with ribavirin (2002 NIH Consensus Development Conference on the Management of Hepatitis C). However, the side effects of combination therapy can be significant and include hemolysis, flu-like symptoms, anemia, and fatigue (Gary L. Davis. *Gastroenterology* 118:S104-S114, 2000).

(3) Substrate-based NS3 protease inhibitors (for example, Attwood *et al.*, *Antiviral peptide derivatives*, PCT WO 98/22496, 1998; Attwood *et al.*, *Antiviral Chemistry and Chemotherapy* 1999, 10, 259-273; Attwood *et al.*, *Preparation and use of amino acid derivatives as anti-viral agents*, German Patent Pub. DE 19914474; Tung *et al.* *Inhibitors of serine proteases, particularly hepatitis C virus NS3 protease*, PCT WO 98/17679), including alphaketoamides and hydrazinoureas, and inhibitors that terminate in an electrophile such as a boronic acid or phosphonate (Llinas-Brunet *et al.*, *Hepatitis C inhibitor peptide analogues*, PCT WO 99/07734).

(4) Non-substrate-based inhibitors, for example, 2,4,6-trihydroxy-3-nitro-benzamide derivatives (Sudo K. *et al.*, *Biochemical and Biophysical Research Communications*, 1997, 238, 643-647; Sudo K. *et al.* *Antiviral Chemistry and Chemotherapy*, 1998, 9, 186), including RD3-4082 and RD3-4078, the former substituted on the amide with a 14 carbon chain and the latter processing a *para*-phenoxyphenyl group;

(5) Thiazolidine derivatives which show relevant inhibition in a reverse-phase HPLC assay with an NS3/4A fusion protein and NS5A/5B substrate (for example Sudo K. *et al.*, *Antiviral Research*, 1996, 32, 9-18), especially compound RD-1-6250,

possessing a fused cinnamoyl moiety substituted with a long alkyl chain, RD4 6205 and RD4 6193;

5 (6) Thiazolidines and benzanilides (for example Kakiuchi N. *et al. J. EBS Letters* 421, 217-220; and Takeshita N. *et al. Analytical Biochemistry*, 1997, 247, 242-246);

10 (7) A phenanthrenequinone possessing activity against protease in a SDS-PAGE and autoradiography assay isolated from the fermentation culture broth of *Streptomyces* sp., for example, Sch 68631 (Chu M. *et al., Tetrahedron Letters*, 1996, 37, 7229-7232), and Sch 351633, isolated from the fungus *Penicillium griseofulvum*, which demonstrates activity in a scintillation proximity assay (Chu M. *et al., Bioorganic and Medicinal Chemistry Letters* 9, 1949-1952);

15 (8) Selective NS3 inhibitors, for example, those based on the macromolecule elgin c, isolated from leech (Qasim M.A. *et al., Biochemistry*, 1997, 36, 1598-1607);

20 (9) Helicase inhibitors (for example Diana G.D. *et al., Compounds, compositions and methods for treatment of hepatitis C*, U.S. Pat. No. 5,633,358; Diana G.D. *et al., Piperidine derivatives, pharmaceutical compositions thereof and their use in the treatment of hepatitis C*, PCT WO 97/36554);

25 (10) Polymerase inhibitors for example nucleotide analogues, gliotoxin (Ferrari R. *et al. Journal of Virology*, 1999, 73, 1649-1654), and the natural product cerulenin (Lohmann V. *et al., Virology*, 1998, 249, 108-118);

 (11) Antisense phosphorothioate oligodeoxynucleotides (S-ODN) complementary to sequence stretches in the 5' non-coding region (NCR) of the virus (Alt M. *et al., Hepatology*, 1995, 22, 707-717), or nucleotides 326-348 comprising the 3' end

of the NCR and nucleotides 371-388 located in the core coding region of the HCV RNA (Alt M. *et al.*, *Archives of Virology*, **1997**, *142*, 589-599; Galderisi U. *et al.*, *Journal of Cellular Physiology*, **1999**, *181*, 251-257).

5 (12) Inhibitors of IRES-dependent translation (Ikeda N *et al.*, *Agent for the prevention and treatment of hepatitis C*, Japanese Patent Pub. JP-08268890; Kai Y. *et al.* *Prevention and treatment of viral diseases*, Japanese Patent Pub. JP-10101591).

10 (13) Nuclease-resistant ribozymes (for example Maccjak, D. J. *et al.*, *Hepatology* **1999**, *30*, abstract 995).

 (14) Nucleoside analogs have also been developed for the treatment of *Flaviviridae* infections.

15 Idenix Pharmaceuticals, Ltd. discloses branched nucleosides, and their use in the treatment of HCV and flaviviruses and pestiviruses in US Patent Publication No. 2003/0050229 A1 and US Patent Publication No. 2003/0060400 A1, which correspond to International Publication Nos. WO 01/90121 and WO 01/92282. A method for the treatment of hepatitis C infection (and flaviviruses and pestiviruses) in humans and other host animals is disclosed in the Idenix publications that includes administering an effective amount of a biologically active 1', 2', 3' or 4'-branched β -D or β -L nucleosides or a pharmaceutically acceptable salt or prodrug thereof, administered either alone or in combination, optionally in a pharmaceutically acceptable carrier.

25 Other patent applications disclosing the use of certain nucleoside analogs to treat hepatitis C virus include: International Patent Publication Nos. WO 01/32153 (PCT/CA00/01316; filed November 3, 2000) and WO 01/60315 (PCT/CA01/00197; filed February 19, 2001) filed by BioChem Pharma, Inc. (now Shire Biochem, Inc.); US Patent Publication No. 2002/0147160 and the corresponding International Patent Publication Nos. WO 02/057425 (PCT/US02/01531; filed January 18, 2002) and WO 02/057287 (PCT/US02/03086; filed January 18, 2002) filed by Merck & Co., Inc.; US Patent Publication Nos. 2003/083307 A1 and US 2003/008841 A1, and the

corresponding International Patent Publication Nos. WO 02/18404 (PCT/EP01/09633; published August 21, 2001); WO 02/100415 and WO 02/094289, filed by Hoffman-LaRoche; US Patent Publication No. 2003/028013 A1 and the corresponding International Patent Publication Nos. WO 03/062255 and WO 03/061385 filed by Ribapharm; and WO 01/79246 and WO 02/32920 filed by Pharmasset.

(15) Miscellaneous compounds including 1-amino-alkylcyclohexanes (U.S. Patent No. 6,034,134 to Gold *et al.*), alkyl lipids (U.S. Pat. No. 5,922,757 to Chojkier *et al.*), vitamin E and other antioxidants (U.S. Pat. No. 5,922,757 to Chojkier *et al.*), squalene, amantadine, bile acids (U.S. Pat. No. 5,846,964 to Ozeki *et al.*), N-(phosphonoacetyl)-L-aspartic acid, (U.S. Pat. No. 5,830,905 to Diana *et al.*), benzenedicarboxamides (U.S. Pat. No. 5,633,388 to Diana *et al.*), polyadenylic acid derivatives (U.S. Pat. No. 5,496,546 to Wang *et al.*), 2',3'-dideoxyinosine (U.S. Pat. No. 5,026,687 to Yarchoan *et al.*), and benzimidazoles (U.S. Pat. No. 5,891,874 to Colacino *et al.*).

(16) Other compounds currently in clinical development for treatment of hepatitis c virus include: Interleukin-10 by Schering-Plough, IP-501 by Interneuron, Merimebodib VX-497 by Vertex, AMANTADINE (Symmetrel) by Endo Labs Solvay, HEPTAZYME by RPI, IDN-6556 by Idun Pharma., XTL-002 by XTL., HCV/MF59 by Chiron, CIVACIR by NABI, LEVOVIRIN by ICN, VIRAMIDINE by ICN, ZADAXIN (thymosin alfa-1) by Sci Clone, CEPLANE (histamine dihydrochloride) by Maxim, VX 950 / LY 570310 by Vertex/Eli Lilly, ISIS 14803 by Isis Pharmaceutical/Elan, IDN-6556 by Idun Pharmaceuticals, Inc. and JTK 003 by AKROS Pharma.

Drug-resistant variants of viruses can emerge after prolonged treatment with an antiviral agent. Drug resistance most typically occurs by mutation of a gene that encodes for an enzyme used in viral replication, and, for example, in the case of HIV, reverse transcriptase, protease, or DNA polymerase. It has been demonstrated that the efficacy of a drug against viral infection can be prolonged, augmented, or restored by administering the compound in combination or alternation with a second, and perhaps

third, antiviral compound that is effective in combating the virus. The pharmacokinetics, biodistribution, or other parameter of the drug can be altered by such combination or alternation therapy. In general, combination therapy is typically preferred over alternation therapy because it induces multiple simultaneous pressures on the virus. One cannot predict, however, what mutations will be induced in the viral genome by a given drug, whether the mutation is permanent or transient, or how an infected cell with or without a mutated viral sequence will respond to therapy with other agents in combination or alternation. This is exacerbated by the fact that there is a paucity of data on the kinetics of drug resistance in long-term cell cultures treated with modern antiviral agents.

It is an object of the present invention to optimize the treatment of HCV infection.

It is a further object to provide the optimal administration of 2'-branched nucleosides, and in particular, 2'-branched pyrimidine nucleosides, for the treatment of *Flaviviridae* infections.

It is another object of the present invention to provide a method and composition that includes 2'-branched nucleosides for the treatment of patients infected with *pestiviruses*, *flaviviruses*, or *hepaciviruses* that exhibit advantageous or improved pharmacokinetic, biodistribution, metabolic, resistance or other parameters over administration of 2'-branched pyrimidine nucleosides alone.

It is yet another object of the present invention to provide a method and composition for the treatment of patients infected with *Flaviviridae* in which 2'-branched nucleosides, and in particular, 2'-branched pyrimidine nucleosides are administered in combination and/or alternation with one or more compounds that act synergistically or advantageously with 2'-branched pyrimidine nucleosides against the virus.

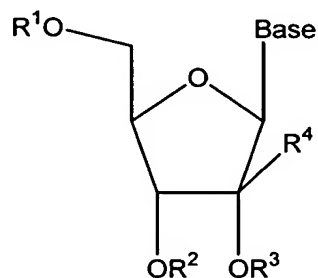
It is still another object of the present invention to provide a method and composition for the treatment of patients infected with a drug resistant form of *pestiviruses*, *flaviviruses*, or *hepaciviruses*.

It is also an object of the invention to provide a method and kit to identify a mutant strain of *Flaviviridae*.

SUMMARY OF THE INVENTION

It has been discovered that prolonged use of a 2'-branched nucleoside, for example a 2'-branched nucleoside depicted below, and in particular, a 2'-branched pyrimidine nucleoside, such as the compound β -D-2'-CH₃-riboC, or a 2'-branched purine nucleoside, including the compound β -D-2'-CH₃-riboAdenosine or β -D-2'-CH₃-ribo-6-N-methyl amino adenosine, is associated with a mutation at a nucleotide that encodes for serine in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region (**Figure 11**) of *Flaviviridae*, which results in a change in the amino acid residue serine to a different amino acid, for example, threonine. This domain is found in the NS5B region of the HCV genome, as well as in genomes of other flaviviruses. It is highly conserved among all hepaci-, pesti- and flavivirus genomes (**Figure 11**, Lai et al. *J Virol.* **1999**, 73, 10129-36).

In one embodiment of the invention, the 2'-branched nucleoside is of the general formula:



or its pharmaceutically acceptable prodrug and/or salt, wherein

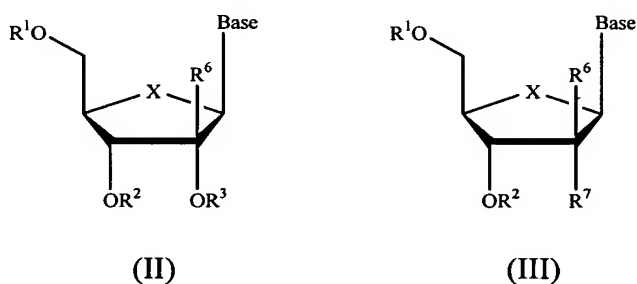
R¹, R², and R³ are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; lipid

(including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R^1 , R^2 or R^3 is independently H or phosphate when administered *in vivo*; and

5 R^4 is hydrogen, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, $-C(O)O(alkyl)$, $-C(O)O(lower\ alkyl)$, $-O(acyl)$, $-O(lower\ acyl)$, $-O(alkyl)$, $-O(lower\ alkyl)$, $-O(alkenyl)$, chloro, bromo, fluoro, iodo, NO_2 , NH_2 , $-NH(lower\ alkyl)$, $-NH(acyl)$, $-N(lower\ alkyl)_2$, or $-N(acyl)_2$;

and Base is a purine or pyrimidine as further described herein.

10 In a second embodiment of the invention, the 2'-branched nucleoside is of the general formula:



or its pharmaceutically acceptable prodrug and/or salt, wherein

Base is a purine or pyrimidine base as defined herein;

15 R^1 , R^2 and R^3 are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate); straight chained, branched or cyclic alkyl (including lower alkyl); acyl (including lower acyl); CO-alkyl, CO-aryl, CO-alkoxyalkyl, CO-aryloxyalkyl, CO-substituted aryl, sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally
20 substituted with one or more substituents as described in the definition of aryl given herein; alkylsulfonyl, arylsulfonyl, aralkylsulfonyl, lipid (including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R^1 , R^2 or R^3 is independently H or phosphate when administered *in vivo*;

25 R^6 is alkyl (including lower alkyl and halogenated alkyl), CH_3 , CF_3 , azido, cyano, alkenyl, alkynyl, Br-vinyl, 2-Br-ethyl, $-C(O)O(alkyl)$, $-C(O)O(lower\ alkyl)$, $-O(acyl)$,

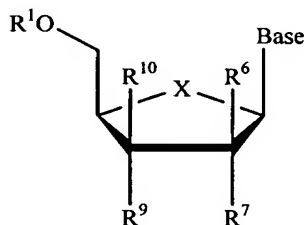
-O(lower acyl), -O(alkyl), -O(lower alkyl), -O(alkenyl), CF₃, chloro, bromo, fluoro, iodo, NO₂, NH₂, -NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂; and

R⁷ is hydrogen, OR³, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, -C(O)O(alkyl), -C(O)O(lower alkyl), -O(acyl), -O(lower acyl),
5 -O(alkyl), -O(lower alkyl), -O(alkenyl), fluorine, chlorine, bromine, iodine, NO₂, NH₂, -NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂; and

X is O, S, SO₂ or CH₂.

and Base is a purine or pyrimidine as further described herein.

In a third embodiment the invention, the 2'-branched nucleoside is of the general
10 formula:



(IV)

wherein:

Base is a purine or pyrimidine base as defined herein;

15 R⁶ is alkyl (including lower alkyl and halogenated alkyl), CH₃, CF₃, azido, cyano, alkenyl, alkynyl, Br-vinyl, 2-Br-ethyl, -C(O)O(alkyl), -C(O)O(lower alkyl), -O(acyl), -O(lower acyl), -O(alkyl), -O(lower alkyl), -O(alkenyl), CF₃, fluoro, chloro, bromo, iodo, NO₂, NH₂, -NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂;

20 R⁷ is OR², hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, halo-vinyl, -C(O)O(alkyl), -C(O)O(lower alkyl), -O(acyl), -O(lower acyl), -O(alkyl), -O(lower alkyl), -O(alkenyl), fluorine, chlorine, bromine, iodine, NO₂, NH₂, -NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂;

R⁹ is hydrogen, OR³, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, -C(O)O(alkyl), -C(O)O(lower alkyl), -O(acyl), -O(lower acyl),

-O(alkyl), -O(lower alkyl), -O(alkenyl), chlorine, bromine, iodine, NO₂, NH₂,
-NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂;

R¹⁰ is H, alkyl (including lower alkyl), fluorine, chlorine, bromine or iodine;

5 R¹, R² and R³ are independently H, phosphate (including mono-, di- or triphosphate and
a stabilized phosphate); straight chained, branched or cyclic alkyl (including lower
alkyl); acyl (including lower acyl); CO-alkyl, CO-aryl, CO-alkoxyalkyl, CO-
aryloxyalkyl, CO-substituted aryl, sulfonate ester (including alkyl or arylalkyl
sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally
10 substituted with one or more substituents as described in the definition of aryl given
herein; alkylsulfonyl, arylsulfonyl, aralkylsulfonyl, lipid (including a phospholipid);
amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable
leaving group that provides a compound wherein R¹, R² or R³ is independently H or
phosphate when administered *in vivo*; and

X is O, S, SO₂ or CH₂.

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In the case of BVDV infection, 2'-branched nucleosides, and, in particular, 2'-
branched pyrimidine nucleosides such as the compound β-D-2'-CH₃-riboC induce a
mutation from a guanine (G) to cytidine (C) at residue 1214 of the RNA polymerase of
BVDV, which results in a change in the amino acid residue serine to threonine at
20 position 405 of the enzyme. This serine residue is located in the conserved consensus
sequence (XRXS_GXXXXT) of the RNA polymerase domain B (**Figures 5 and 11**),
identified by mutational analysis (Lai V. C., Kao C.C., Ferrari E., Park J., Uss A.S.,
Wright-Minogue J., Hong Z., and J.Y. Lau. "Mutational analysis of bovine viral diarrhea
virus RNA-dependent RNA polymerase" J Virol. **1999**, 73, 10129-36).

25 In the case of HCV infection, 2'-branched nucleosides, and, in particular, 2'-
branched pyrimidine nucleosides such as the compound β-D-2'-CH₃-riboC induce a
mutation at a nucleotide that encodes for Serine₂₈₂ in the highly conserved consensus
sequence, XRXS_GXXXXT, of domain B of the RNA polymerase region (**Figure 11**),
which results in a change from serine to a different amino acid, such as threonine.

Furthermore, it has been discovered that 2'-branched nucleosides and interferon act synergistically to inhibit *Flaviviridae*. In particular 2'-branched pyrimidine nucleosides such as the compound β -D-2'-CH₃-riboC or a 2'-branched purine nucleosides such as the compound β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methyl amino adenosine, and interferon alpha-2b administered in combination and/or alternation act synergistically to inhibit *Flaviviridae*. Moreover, it has been discovered that the resistant viral populations, which emerge after 2'-branched nucleoside treatment, for example, β -D-2'-CH₃-riboC treatment, show increased sensitivity to subsequent treatment with interferon. Thus, sequential and/or combination therapy of a 2'-branched nucleoside and interferon can substantially reduce *Flaviviridae* infections.

One aspect of the present invention provides a method to treat a *Flaviviridae* infection by administering a therapeutically effective amount of a 2'-branched nucleoside, for example, a 2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC, or its pharmaceutically acceptable prodrug and/or salt, to a host, such as a human, in need of such therapy, in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that are associated with such a mutation. This highly conserved serine residue corresponds to amino acid position 405 of the RNA polymerase region of the BVDV genome. It also corresponds to amino acid position 282 of the RNA polymerase region of the HCV genome (**Figure 11**; Lai et al. *J Virol.*, 1999, 73, 10129-36).

Another aspect of the present invention provides a method to treat and/or to substantially cure a *Flaviviridae* infection in a host infected with a *Flaviviridae* that contains a serine to threonine mutation at the conserved serine residue of a *Flaviviridae* (**Figure 11**), for example, amino acid 405 of the RNA polymerase region of BVDV or amino acid 282 of the RNA polymerase of HCV, by administering a therapeutically effective amount of interferon. In a specific embodiment, interferon alpha-2b is administered to treat and/or to substantially cure the infection caused by a mutated *Flaviviridae* virus.

The invention disclosed herein also minimally includes at least the following embodiments:

- (i) A pharmaceutical composition effective for the treatment of a *Flaviviridae* infection in a host, such as a human, comprising an effective amount of a 2'-branched nucleoside, for example, a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or its pharmaceutically acceptable prodrug and/or salt, optionally in a pharmaceutically acceptable carrier or diluent, in combination with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, for example, other than nucleotide 1214 (G to C) or 405 Ser to Thr of the RNA polymerase region of BVDV or nucleotide 8443 (G to C) of the HCV genome or 282 Ser to Thr of the RNA polymerase region of HCV (**Figure 11**; Lai et al. *J Virol.*, **1999**, 73, 10129-36), and/or one or more drugs that are associated with such a mutation.
- (ii) A pharmaceutical composition effective for the treatment of a *Flaviviridae* infection in a host, such as a human, comprising an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable prodrug and/or salt thereof, optionally in a pharmaceutically acceptable carrier or diluent, in combination with interferon. Interferons include: Intron-A (interferon alpha-2b) by Schering, PEG-INTRON (pegylated interferon alpha-2b) by Schering, Roferon-A (interferon alfa-2a) by Roche, PEGASYS (pegylated interferon alfa-2a) by Roche, INFERGEN (interferon alfacon-1) by InterMune, OMNIFERON (natural interferon) by Viragen, ALBUFERON by Human Genome Sciences, REBIF (interferon beta-1a)

by Ares-Serono, Omega Interferon by BioMedicine, Oral Interferon Alpha by Amarillo Biosciences, and Interferon gamma-1b by InterMune.

- (iii) A pharmaceutical composition effective for the treatment of a *Flaviviridae* infection in a host, such as a human, comprising:

an effective amount of a 2', 3' and/or 5'-prodrug of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, including the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, including the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, or a pharmaceutically acceptable salt thereof, optionally in a pharmaceutically acceptable carrier or diluent;

in combination with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that are associated with such a mutation.

- (iv) A method for treating a *Flaviviridae* infection in a host, such as a human, comprising administering an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or its pharmaceutically acceptable prodrug and/or salt to the human, optionally in a pharmaceutically acceptable carrier or diluent, in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, for example, other than nucleotide 1214 (G to C) or 405 Ser to Thr of the RNA polymerase region of BVDV or nucleotide 8443 (G to C) of the HCV genome or 282 Ser to Thr of the RNA polymerase region of HCV (**Figure 11**; Lai et al. J

Virol., 1999, 73, 10129-36), and/or one or more drugs that are associated with such a mutation.

- (v) A method for treating a *Flaviviridae* infection in a host, such as a human, comprising administering an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable prodrug and/or salt thereof to the host, optionally in a pharmaceutically acceptable carrier or diluent, in combination and/or alternation with interferon. Interferons include: Intron-A (interferon alpha-2b) by Schering, PEG-INTRON (pegylated interferon alpha-2b) by Schering, Roferon-A (interferon alfa-2a) by Roche, PEGASYS (pegylated interferon alfa-2a) by Roche, INFERGEN (interferon alfacon-1) by InterMune, OMNIFERON (natural interferon) by Viragen, ALBUFERON by Human Genome Sciences, REBIF (interferon beta-1a) by Ares-Serono, Omega Interferon by BioMedicine, Oral Interferon Alpha by Amarillo Biosciences, and Interferon gamma-1b by InterMune.
- (vi) A method for treating a patient infected with a *Flaviviridae* virus that is resistant to a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, comprising administering an effective amount of interferon, optionally in a pharmaceutically acceptable carrier or diluent, optionally in a manner that substantially eliminates the viral load.
- (iv) A method for treating a patient infected with *Flaviviridae* comprising:
administering an effective amount of a 2', 3' and/or 5'-prodrug of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-

riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, optionally in a pharmaceutically acceptable carrier or diluent;

in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that are associated with such a mutation.

(v) A method for treating a patient infected with *Flaviviridae* comprising:

(a) administering to the patient an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; optionally in a pharmaceutically acceptable carrier or diluent;

(b) assaying the blood of the patient to test for seroconversion from wildtype to mutant virus; and

(c) administering an effective amount of interferon; optionally in a pharmaceutically acceptable carrier or diluent.

(vi) A method for assaying a sample suspected of containing a *Flaviviridae* resistant to a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; comprising:

(a) contacting a sample containing a *Flaviviridae* nucleic acid sequence with a detectable oligonucleotide probe having a sequence complementary a codon

that encodes a serine in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region of *Flaviviridae* (Figure 11);

(b) allowing the probe to hybridize to the sequence; and

5 (c) detecting the hybridization of the probe the sequence.

(vii) A method for assaying a sample suspected of containing a *Flaviviridae* resistant to a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; comprising:

10

(a) contacting a sample containing a *Flaviviridae* nucleic acid sequence with a detectable oligonucleotide probe having a sequence complementary to the cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or the cytidine at nucleotide 8443 of HCV;

15

(b) allowing the probe to hybridize to the sequence; and

(c) detecting the hybridization of the probe to cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or at nucleotide 8443 of HCV.

(viii) A method for treating a patient infected with *Flaviviridae* comprising:

20 (a) administering an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; optionally in a pharmaceutically acceptable carrier or diluent;

25

(b) obtaining a viral sample from the patient;

(c) determining the replication fitness of the virus;

(d) determining whether the replication fitness of the virus in the sample is less than the replication fitness of the wild-type virus, which indicates resistance to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; and

(e) administering an effective amount of interferon to those patients that are resistant to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof.

(ix) A method for treating a patient infected with *Flaviviridae* comprising:

(a) administering an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; optionally in a pharmaceutically acceptable carrier or diluent;

(b) obtaining a viral culture sample from the patient;

(c) culturing the sample and comparing the plaque growth between the sample and wild type virus;

(d) determining whether the plaque growth of the sample is smaller than the plaque growth of the wildtype, which indicates resistance to the 2'-branched nucleoside; and

(e) administering an effective amount of interferon to those patients that are resistant to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof.

(x) A method for diagnosing the presence of a *Flaviviridae* resistant to a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, in a patient comprising:

(a) obtaining a sample suspected of containing a *Flaviviridae* nucleic acid sequence;

(b) contacting the sample with a detectable oligonucleotide probe having a sequence complementary a codon that encodes a serine in the highly conserved consensus sequence, XXSGXXXT, of domain B of the RNA polymerase region of *Flaviviridae* (Figure 11);

(b) allowing the probe to hybridize to the sequence; and

(c) detecting the hybridization of the probe the sequence to determine the presence of a *Flaviviridae* resistant to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof.

(xi) A method for diagnosing of the presence of a *Flaviviridae* resistant to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-

CH₃-riboC, or a β-D-2'-branched purine nucleoside, for example β-D-2'-CH₃-riboA or β-D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, in a patient comprising:

- 5 (a) obtaining a sample suspected of containing a *Flaviviridae* nucleic acid sequence;
- (b) contacting the sample with a detectable oligonucleotide probe having a sequence complementary to the cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or the cytidine at nucleotide 8443 of HCV;
- 10 (b) allowing the probe to hybridize to the sequence; and
- (c) detecting the hybridization of the probe to cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or at nucleotide 8443 of HCV to determine the presence of a *Flaviviridae* resistant to the 2'-branched nucleoside, such as β-D-2'-branched pyrimidine nucleoside, for example β-D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β-D-2'-CH₃-riboC, or a β-D-2'-branched purine nucleoside, for example β-D-2'-CH₃-riboA or β-D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof.
- 15

20 The disclosed combination and/or alternation regimens are useful in the prevention and treatment of *Flaviviridae* infections, including BVDV, BDV, CSFV, DHF, yellow fever virus, shock syndrome, Japanese encephalitis virus, and HCV.

In addition, the corresponding amino acid sequences of the *Flaviviridae* viral markers diagnostic for long term response of *Flaviviridae* carriers to 2'-branched nucleoside therapy can be determined from the illustrative *Flaviviridae* nucleotide sequences.

25

In addition to identifying viral markers for the purposes of identifying *Flaviviridae* strains that are associated with 2'-branched nucleoside failure, the present invention can be utilized to also identify *Flaviviridae* strains that respond to 2'-branched

nucleoside therapy. In this respect, the absence of viral markers correlated with 2'-branched nucleoside therapy can be used to prescribe a course of treatment that includes 2'-branched nucleoside as a modality for those individuals that carrier *Flaviviridae* lacking viral markers correlated with 2'-branched nucleoside therapy failure.

5 In another embodiment, the invention provides an oligonucleotide primer for amplifying an *Flaviviridae* nucleic acid sequence. In one embodiment, the oligonucleotide is at least 14 nucleotides in length and hybridizes under sequence-specific, stringent hybridization conditions to a nucleotide sequence that contains the marker correlated with therapy failure.

10 Oligonucleotide sequences used as the hybridizing region of a primer can also be used as the hybridizing region of a probe. Suitability of a primer sequence for use as a probe depends on the hybridization characteristics of the primer. Similarly, an oligonucleotide used as a probe can be used as a primer.

15 Additionally, the invention provides a method, materials and a kit to detect proteins, peptides or peptide fragments that contain amino acids (as described extensively herein) that are predictive of the long term response of an *Flaviviridae* carrier to 2'-branched nucleoside therapy, or antibodies to those proteins, peptides or peptide fragments. Host sera or tissue can be tested for either the protein or peptide or the antibody to the protein or peptide, depending on convenience and perhaps
20 concentration of the diagnostic material.

The protein, peptide or peptide fragment can be confirmed by reaction with an antibody, preferably a monoclonal antibody, for example using a Western blot method. Alternatively, the protein or peptide can be isolated and sequenced or otherwise identified by any means known in the art, including by 2D PAGE. In one embodiment, a
25 reactive antibody binds to an *Flaviviridae* protein or peptide sequence that includes a threonine rather than serine in the highly conserved consensus sequence, XXRXSGXXXT, of domain B of the RNA polymerase region, for example at position 405 of the RNA polymerase region of BVDV genome or at position 282 of the RNA polymerase region of the HCV genome.

30 In another embodiment, the reactive antibody binds specifically to a peptide sequence that includes a threonine rather than serine in the highly conserved consensus

sequence, XRXSGXXXT, of domain B of the RNA polymerase region, for example at position 405 of the RNA polymerase region of BVDV genome or at position 282 of the RNA polymerase region of the HCV genome, which represent a specific point mutations in the RNA polymerase region of *Flaviviridae* that is correlated with therapy failure.

5 In specific embodiments, an antibody is used that binds to at least one peptide or peptide fragment encoded for by the nucleic acid sequences in sequence ID Nos. 1-31.

 In specific embodiments, an antibody is used that binds to at least one peptide or peptide fragment encoded for by the nucleic acid sequences in sequence ID Nos.32-62.

BRIEF DESCRIPTION OF THE FIGURES

10 **Figure 1** shows the emergence of β -D-2'-CH₃-riboC-resistant BVDV from untreated BVDV.

Figure 2 illustrates the phenotype of focus formation by (A) the wild type (wt) BVDV (strain I-N-dIns), and by (B) the β -D-2'-CH₃-riboC -resistant BVDV (I-N-dIns β -D-2'-CH₃-riboC-R).

15 **Figure 3** demonstrates the growth kinetics of wild-type BVDV I-N-dIns and its β -D-2'-CH₃-riboC -resistant mutant, I-N-dIns β -D-2'-CH₃-riboC-R (Resistant Mutant).

Figure 4 shows the effect of β -D-2'-CH₃-riboC on the virus yield in *de novo*-infected MDBK cells (wherein I-N-dIns is wt BVDV, and the resistant mutant is I-N-dIns β -D-2'-CH₃-riboC-R (β -D-2'-CH₃-riboC-resistant BVDV)).

20 **Figure 5** is a schematic representation of the BVDV NS5B region showing the proposed functional domains, based on the mutational analysis (Vassilev, V. B. and R. O. Donis. (2000) Bovine viral diarrhea virus induced apoptosis correlates with increased intracellular viral RNA accumulation. Virus Res. 69 (2): 95-107). The large arrow indicates the position of the only amino acid change found in the NS5B region of the β -D-2'-CH₃-riboC – resistant BVDV (Ser 405 to Thr 405).

25

Figure 6 illustrates the effect of interferon alpha-2b on the virus yields in *de novo*-infected MDBK cells (I-N-dIns: wt BVDV; I-N-dIns β -D-2'-CH₃-riboC-R: β -D-2'-CH₃-riboC -resistant BVDV (resistant mutant)).

Figure 7 illustrates the effect of β -D-2'-CH₃-riboC and interferon alpha-2b on BVDV (strain I-N-dIns) titers in persistently infected MDBK cells

Figure 8 demonstrates the effect of β -D-2'-CH₃-riboC in combination with interferon alpha-2b on wild-type BVDV (strain I-N-dIns) titers in persistently infected MDBK cells.

Figure 9 shows the effect of β -D-2'-CH₃-riboC and interferon alpha-2b (Intron A) on BVDV (strain NY-1) titers in persistently infected MDBK cells.

Figure 10 illustrates the effect of β -D-2'-CH₃-riboC and interferon alpha-2b (Intron A) on wild-type BVDV (strain I-N-dIns) titers in persistently infected MDBK cells.

Figure 11 illustrates the alignment of the RNA Polymerase in domain B of various *Flaviviridae*; bold type with larger font show amino acids that are 100% conserved; underlined serine amino acid residues are ones that may be mutated to threonine after treatment with a 2'-branched nucleoside. [The 100% conserved serine residue is also underlined and represents the amino acid that is mutated to Threonine after treatment with a 2'-branched nucleoside (Ser₄₀₅ of the RNA polymerase region of BVDV; Ser₂₈₂ of the RNA polymerase region of HCV). See Lai et al. *J Virol.* **1999**, 73, 10129-36.]

DETAILED DESCRIPTION OF THE INVENTION

It has been discovered that prolonged use of a 2'-branched nucleoside, for example a 2'-branched nucleoside depicted below, and in particular, a 2'-branched pyrimidine nucleoside such as the compound β -D-2'-CH₃-riboC, is associated with a mutation at a nucleotide that encodes for serine in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region (**Figure 11**) of *Flaviviridae* resulting in a change in the amino acid residue serine to a different amino

acid, for example, threonine. This domain is found in the NS5B region of the HCV genome, as well as in genomes of other flaviviruses. It is highly conserved among all hepaci-, pesti- and flavivirus genomes (**Figure 11**, Lai et al. *J Virol.* **1999**, 73, 10129-36).

5 In the case of BVDV infection, 2'-branched nucleosides, and, in particular, 2'-branched pyrimidine nucleosides such as the compound β -D-2'-CH₃-riboC induce a mutation from a guanine (G) to cytidine (C) at residue 1214 of the RNA polymerase of BVDV causing a change in the amino acid residue serine to threonine at position 405 of the enzyme. This serine residue is located in the conserved consensus sequence (XRXSGXXXXT) of the RNA polymerase domain B (**Figures 5 and 11**), identified by
10 mutational analysis (Lai V. C., Kao C.C., Ferrari E., Park J., Uss A.S., Wright-Minogue J., Hong Z., and J.Y. Lau. "Mutational analysis of bovine viral diarrhea virus RNA-dependent RNA polymerase" *J Virol.* **1999**, 73, 10129-36).

15 In the case of HCV infection, 2'-branched nucleosides, and, in particular, 2'-branched pyrimidine nucleosides such as the compound β -D-2'-CH₃-riboC induce a mutation at a nucleotide that encodes for Serine₂₈₂ in the highly conserved consensus sequence, XRXSGXXXXT, of domain B of the RNA polymerase region (**Figure 11**) resulting in a change from serine to a different amino acid, such as threonine.

20 Furthermore, it has been discovered that 2'-branched nucleosides and interferon act synergistically to inhibit *Flaviviridae*. In particular 2'-branched pyrimidine nucleosides such as the compound β -D-2'-CH₃-riboC and interferon alpha-2b administered in combination and/or alternation act synergistically to inhibit *Flaviviridae*. Moreover, it has been discovered that the resistant viral populations, which emerge after 2'-branched nucleoside treatment, for example, β -D-2'-CH₃-riboC treatment, show
25 increased sensitivity to subsequent treatment with interferon. Thus, sequential and/or combination therapy of a 2'-branched nucleoside and interferon can substantially reduce *Flaviviridae* infections.

30 One aspect of the present invention provides a method to treat a *Flaviviridae* infection by administering a therapeutically effective amount of a 2'-branched nucleoside, for example, a 2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC, or its pharmaceutically acceptable prodrug and/or salt, to a host, such as a human,

in need of such therapy, in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that are associated with such a mutation. This highly conserved serine residue corresponds to amino acid position 405 of the RNA polymerase region of the BVDV genome. It also corresponds to amino acid position 282 of the RNA polymerase region of the HCV genome (**Figure 11**; Lai et al. *J Virol.*, 1999, 73, 10129-36).

Another aspect of the present invention provides a method to treat and/or to substantially cure a *Flaviviridae* infection in a host infected with a *Flaviviridae* that contains a serine to threonine mutation at the conserved serine residue of a *Flaviviridae* (**Figure 11**), for example, amino acid 405 of the RNA polymerase region of BVDV or amino acid 282 of the RNA polymerase of HCV, by administering a therapeutically effective amount of interferon. In a specific embodiment, interferon alpha-2b is administered to treat and/or to substantially cure the infection caused by a mutated *Flaviviridae* virus.

The invention disclosed herein also minimally includes at least the following embodiments:

- (i) A pharmaceutical composition effective for the treatment of a *Flaviviridae* infection in a host, such as a human, comprising an effective amount of a 2'-branched nucleoside, for example, a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or its pharmaceutically acceptable prodrug and/or salt, optionally in a pharmaceutically acceptable carrier or diluent, in combination with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA

polymerase region, for example, other than nucleotide 1214 (G to C) or 405 Ser to Thr of the RNA polymerase region of BVDV or nucleotide 8443 (G to C) of the HCV genome or 282 Ser to Thr of the RNA polymerase region of HCV (**Figure 11**; Lai et al. J Virol., **1999**, 73, 10129-36), and/or one or more drugs that are associated with such a mutation.

- (ii) A pharmaceutical composition effective for the treatment of a *Flaviviridae* infection in a host, such as a human, comprising an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable prodrug and/or salt thereof, optionally in a pharmaceutically acceptable carrier or diluent, in combination with interferon. Interferons include: Intron-A (interferon alpha-2b) by Schering, PEG-INTRON (pegylated interferon alpha-2b) by Schering, Roferon-A (interferon alfa-2a) by Roche, PEGASYS (pegylated interferon alfa-2a) by Roche, INFERGEN (interferon alfacon-1) by InterMune, OMNIFERON (natural interferon) by Viragen, ALBUFERON by Human Genome Sciences, REBIF (interferon beta-1a) by Ares-Serono, Omega Interferon by BioMedicine, Oral Interferon Alpha by Amarillo Biosciences, and Interferon gamma-1b by InterMune.

- (iii) A pharmaceutical composition effective for the treatment of a *Flaviviridae* infection in a host, such as a human, comprising:

an effective amount of a 2', 3' and/or 5'-prodrug of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, including the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, including the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, or a pharmaceutically acceptable salt thereof, optionally in a pharmaceutically acceptable carrier or diluent;

in combination with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that

results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT , of domain B of the RNA polymerase region, and/or one or more drugs that are associated with such a mutation.

(iv) A method for treating a *Flaviviridae* infection in a host, such as a human, comprising administering an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'- CH_3 -riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'- CH_3 -riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'- CH_3 -riboA or β -D-2'- CH_3 -ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or its pharmaceutically acceptable prodrug and/or salt to the human, optionally in a pharmaceutically acceptable carrier or diluent, in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT , of domain B of the RNA polymerase region, for example, other than nucleotide 1214 (G to C) or 405 Ser to Thr of the RNA polymerase region of BVDV or nucleotide 8443 (G to C) of the HCV genome or 282 Ser to Thr of the RNA polymerase region of HCV (**Figure 11**; Lai et al. *J Virol.*, **1999**, 73, 10129-36), and/or one or more drugs that are associated with such a mutation.

(v) A method for treating a *Flaviviridae* infection in a host, such as a human, comprising administering an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'- CH_3 -riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'- CH_3 -riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'- CH_3 -riboA or β -D-2'- CH_3 -ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable prodrug and/or salt thereof to the host, optionally in a pharmaceutically acceptable carrier or diluent, in combination and/or alternation with interferon. Interferons include: Intron-A (interferon alpha-2b) by Schering, PEG-INTRON (pegylated interferon alpha-2b) by Schering, Roferon-A (interferon alfa-2a) by Roche, PEGASYS (pegylated interferon alfa-2a) by Roche, INFERGEN (interferon alfacon-1) by InterMune, OMNIFERON (natural

interferon) by Viragen, ALBUFERON by Human Genome Sciences, REBIF (interferon beta-1a) by Ares-Serono, Omega Interferon by BioMedicine, Oral Interferon Alpha by Amarillo Biosciences, and Interferon gamma-1b by InterMune.

(vi) A method for treating a patient infected with a *Flaviviridae* virus that is resistant to a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, comprising administering an effective amount of interferon, optionally in a pharmaceutically acceptable carrier or diluent, optionally in a manner that substantially eliminates the viral load.

(iv) A method for treating a patient infected with *Flaviviridae* comprising:

administering an effective amount of a 2', 3' and/or 5'-prodrug of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, optionally in a pharmaceutically acceptable carrier or diluent;

in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that are associated with such a mutation.

(v) A method for treating a patient infected with *Flaviviridae* comprising:

(a) administering to the patient an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-

- 5 riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; optionally in a pharmaceutically acceptable carrier or diluent;
- (b) assaying the blood of the patient to test for seroconversion from wildtype to mutant virus; and
- (c) administering an effective amount of interferon; optionally in a pharmaceutically acceptable carrier or diluent.
- 10 (vi) A method for assaying a sample suspected of containing a *Flaviviridae* resistant to a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the
- 15 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; comprising:
- (a) contacting a sample containing a *Flaviviridae* nucleic acid sequence with a detectable oligonucleotide probe having a sequence complementary a codon that encodes a serine in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region of *Flaviviridae* (Figure 11);
- 20 (b) allowing the probe to hybridize to the sequence; and
- (c) detecting the hybridization of the probe the sequence.

(vii) A method for assaying a sample suspected of containing a *Flaviviridae* resistant to a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; comprising:

(a) contacting a sample containing a *Flaviviridae* nucleic acid sequence with a detectable oligonucleotide probe having a sequence complementary to the cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or the cytidine at nucleotide 8443 of HCV;

(b) allowing the probe to hybridize to the sequence; and

(c) detecting the hybridization of the probe to cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or at nucleotide 8443 of HCV.

(viii) A method for treating a patient infected with *Flaviviridae* comprising:

(a) administering an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; optionally in a pharmaceutically acceptable carrier or diluent;

(b) obtaining a viral sample from the patient;

(c) determining the replication fitness of the virus;

(d) determining whether the replication fitness of the virus in the sample is less than the replication fitness of the wild-type virus, which indicates resistance to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine

or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; and

- (e) administering an effective amount of interferon to those patients that are resistant to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof.

(ix) A method for treating a patient infected with *Flaviviridae* comprising:

- (a) administering an effective amount of a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof; optionally in a pharmaceutically acceptable carrier or diluent;

(b) obtaining a viral culture sample from the patient;

(c) culturing the sample and comparing the plaque growth between the sample and wild type virus;

(d) determining whether the plaque growth of the sample is smaller than the plaque growth of the wildtype, which indicates resistance to the 2'-branched nucleoside; and

(e) administering an effective amount of interferon to those patients that are resistant to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof.

(x) A method for diagnosing the presence of a *Flaviviridae* resistant to a 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, in a patient comprising:

(a) obtaining a sample suspected of containing a *Flaviviridae* nucleic acid sequence;

(b) contacting the sample with a detectable oligonucleotide probe having a sequence complementary a codon that encodes a serine in the highly conserved consensus sequence, XXRSGXXXT, of domain B of the RNA polymerase region of *Flaviviridae* (**Figure 11**);

(b) allowing the probe to hybridize to the sequence; and

(c) detecting the hybridization of the probe the sequence to determine the presence of a *Flaviviridae* resistant to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof.

(xi) A method for diagnosing of the presence of a *Flaviviridae* resistant to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof, in a patient comprising:

(a) obtaining a sample suspected of containing a *Flaviviridae* nucleic acid sequence;

(b) contacting the sample with a detectable oligonucleotide probe having a sequence complementary to the cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or the cytidine at nucleotide 8443 of HCV;

(b) allowing the probe to hybridize to the sequence; and

5 (c) detecting the hybridization of the probe to cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or at nucleotide 8443 of HCV to determine the presence of a *Flaviviridae* resistant to the 2'-branched nucleoside, such as β -D-2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC or a prodrug, such as the 3'-valine ester prodrug of β -D-2'-
10 CH₃-riboC, or a β -D-2'-branched purine nucleoside, for example β -D-2'-CH₃-riboA or β -D-2'-CH₃-ribo-6-N-methylaminopurine or a prodrug, such as the 3'-valine ester prodrug, or a pharmaceutically acceptable salt thereof.

15 The disclosed combination and/or alternation regimens are useful in the prevention and treatment of *Flaviviridae* infections, including BVDV, BDV, CSFV, DHF, yellow fever virus, shock syndrome, Japanese encephalitis virus, and HCV.

In addition, the corresponding amino acid sequences of the *Flaviviridae* viral markers diagnostic for long term response of *Flaviviridae* carriers to 2'-branched nucleoside therapy can be determined from the illustrative *Flaviviridae* nucleotide
20 sequences.

In addition to identifying viral markers for the purposes of identifying *Flaviviridae* strains that are associated with 2'-branched nucleoside failure, the present invention can be utilized to also identify *Flaviviridae* strains that respond to 2'-branched nucleoside therapy. In this respect, the absence of viral markers correlated with 2'-
25 branched nucleoside therapy can be used to prescribe a course of treatment that includes 2'-branched nucleoside as a modality for those individuals that carrier *Flaviviridae* lacking viral markers correlated with 2'-branched nucleoside therapy failure.

In another embodiment, the invention provides an oligonucleotide primer for amplifying an *Flaviviridae* nucleic acid sequence. In one embodiment, the
30 oligonucleotide is at least 14 nucleotides in length and hybridizes under sequence-

specific, stringent hybridization conditions to a nucleotide sequence that contains the marker correlated with therapy failure.

Oligonucleotide sequences used as the hybridizing region of a primer can also be used as the hybridizing region of a probe. Suitability of a primer sequence for use as a probe depends on the hybridization characteristics of the primer. Similarly, an oligonucleotide used as a probe can be used as a primer.

Additionally, the invention provides a method, materials and a kit to detect proteins, peptides or peptide fragments that contain amino acids (as described extensively herein) that are predictive of the long term response of an *Flaviviridae* carrier to 2'-branched nucleoside therapy, or antibodies to those proteins, peptides or peptide fragments. Host sera or tissue can be tested for either the protein or peptide or the antibody to the protein or peptide, depending on convenience and perhaps concentration of the diagnostic material.

The protein, peptide or peptide fragment can be confirmed by reaction with an antibody, preferably a monoclonal antibody, for example using a Western blot method. Alternatively, the protein or peptide can be isolated and sequenced or otherwise identified by any means known in the art, including by 2D PAGE. In one embodiment, a reactive antibody binds to an *Flaviviridae* protein or peptide sequence that includes a threonine rather than serine in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, for example at position 405 of the RNA polymerase region of BVDV genome or at position 282 of the RNA polymerase region of the HCV genome.

In another embodiment, the reactive antibody binds specifically to a peptide sequence that includes a threonine rather than serine in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, for example at position 405 of the RNA polymerase region of BVDV genome or at position 282 of the RNA polymerase region of the HCV genome, which represent a specific point mutations in the RNA polymerase region of *Flaviviridae* that is correlated with therapy failure.

In specific embodiments, an antibody is used that binds to at least one peptide or peptide fragment encoded for by the nucleic acid sequences in sequence ID Nos. 1-31.

In specific embodiments, an antibody is used that binds to at least one peptide or peptide fragment encoded for by the nucleic acid sequences in sequence ID Nos.32-62.

I. Definitions

As used herein, the term “resistant virus” refers to a virus that exhibits a three, and more typically, five or greater fold increase in EC₅₀ compared to naive virus.

The term “amino acid” includes naturally occurring and synthetic α , β γ or δ amino acids, and includes but is not limited to, amino acids found in proteins, *i.e.* glycine, alanine, valine, leucine, isoleucine, methionine, phenylalanine, tryptophan, proline, serine, threonine, cysteine, tyrosine, asparagine, glutamine, aspartate, glutamate, lysine, arginine and histidine. In a preferred embodiment, the amino acid is in the L-configuration, but can also be in the D-configuration. Alternatively, the amino acid can be a derivative of alanyl, valinyl, leucinyl, isoleuccinyl, prolinyl, phenylalaninyl, tryptophanyl, methioninyl, glycyl, serinyl, threoninyl, cysteinyl, tyrosinyl, asparaginyl, glutaminyl, aspartoyl, glutaroyl, lysinyl, argininyl, histidinyl, β -alanyl, β -valinyl, β -leucinyl, β -isoleuccinyl, β -prolinyl, β -phenylalaninyl, β -tryptophanyl, β -methioninyl, β -glycyl, β -serinyl, β -threoninyl, β -cysteinyl, β -tyrosinyl, β -asparaginyl, β -glutaminyl, β -aspartoyl, β -glutaroyl, β -lysinyl, β -argininyl or β -histidinyl.

The abbreviations of amino acids used herein are described in **Table 1**.

Table 1

Amino Acids			Codons			
Alanine	Ala	A	GCA	GCC	GCG	GCU
Cysteine	Cys	C	UGC	UGU		
Aspartic Acid	Asp	D	GAC	GAU	GAC	GAU
Glutamic Acid	Glu	E	GAA	GAG		
Phenylalanine	Phe	F	UUC	UUU		

Amino Acids			Codons						
Glycine	Gly	G	GGA	GCG	GGG	GGU			
Histidine	His	H	CAC	CAU					
Isoleucine	Ile	I	AUA	AUC	AUU				
Lysine	Lys	K	AAA	AAG					
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	GUU	
Methionine	Met	M	AUG						
Asparagine	Asn	N	AAC	AAU					
Proline	Pro	P	CCA	CCC	CCG	CCU			
Glutamine	Gln	Q	CAA	CAG					
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU	
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU	
Threonine	Thr	T	ACA	ACC	ACG	ACU			
Valine	Val	V	GUA	GUC	GUG	GUU			
Tryptophan	Trp	W	UGG						
Tyrosine	Tyr	Y	UAC	UAU					

"Amplification reagents" refer to the various buffers, enzymes, primers, deoxynucleoside triphosphates (both conventional and unconventional), and primers used to perform the selected amplification procedure.

5 "Amplifying" or "Amplification", which typically refers to an "exponential" increase in target nucleic acid, is being used herein to describe both linear and exponential increases in the numbers of a select target sequence of nucleic acid.

"Bind(s) substantially" refers to complementary hybridization between an oligonucleotide and a target sequence and embraces minor mismatches that can be

accommodated by reducing the stringency of the hybridization media to achieve the desired priming for the PCR polymerases or detection of hybridization signal.

"Hybridizing" refers the binding of two single stranded nucleic acids via complementary base pairing.

5 "Nucleic acid" refers to a deoxyribonucleotide or ribonucleotide polymer in either single- or double-stranded form, and unless otherwise limited, would encompass known analogs of natural nucleotides that can function in a similar manner as naturally occurring nucleotides.

10 "Nucleotide polymerases" refers to enzymes able to catalyze the synthesis of DNA or RNA from nucleoside triphosphate precursors. In amplification reactions, the polymerases are template-dependent and typically add nucleotides to the 3'-end of the polymer being formed. The polymerase can be thermostable as described in U.S. Pat. Nos. 4,889,818 and 5,079,352.

15 The term "oligonucleotide" refers to a molecule comprised of two or more deoxyribonucleotides or ribonucleotides, such as primers, probes, nucleic acid fragments to be detected, and nucleic acid controls. The exact size of an oligonucleotide depends on many factors and the ultimate function or use of the oligonucleotide. Oligonucleotides can be prepared by any suitable method, including, for example, cloning and restriction of appropriate sequences and direct chemical synthesis by a method such as the
20 phosphotriester method of Narang et al., *Meth. Enzymol.* **1979**, 68:90-99; the phosphodiester method of Brown et al, *Meth. Enzymol.*, **1979**, 68:109-151; the diethylphosphoramidite method of Beaucage et al., *Tetrahedron Lett.*, **1981**, 22:1859-1862; and the solid support method of U.S. Pat. No. 4,458,066.

25 The term "primer" refers to an oligonucleotide, whether natural or synthetic, capable of acting as a point of initiation of DNA synthesis under conditions in which synthesis of a primer extension product complementary to a nucleic acid strand is induced, i.e., in the presence of four different nucleoside triphosphates and an agent for polymerization (i.e., DNA polymerase or reverse transcriptase) in an appropriate buffer and at a suitable temperature. A primer is preferably a single-stranded
30 oligodeoxyribonucleotide. The appropriate length of a primer depends on the intended use of the primer but typically ranges from about 14 or about 15 to 25 or 30 nucleotides.

Short primer molecules generally require cooler temperatures to form sufficiently stable hybrid complexes with the template. A primer need not reflect the exact sequence of the template but must be sufficiently complementary to hybridize with a template.

The term "primer" can refer to more than one primer, particularly in the case where there is some ambiguity in the information regarding one or both ends of the target region to be amplified. For instance, if a region shows significant levels of polymorphism in a population, mixtures of primers can be prepared that will amplify alternate sequences. A primer can be labeled, if desired, by incorporating a label detectable by spectroscopic, photochemical, biochemical, immunochemical, or chemical means. For example, useful labels include ^{32}P , fluorescent dyes, electron-dense reagents, enzymes (as commonly used in an ELISA), biotin, or haptens and proteins for which antisera or monoclonal antibodies are available. A label can also be used to "capture" the primer, so as to facilitate the immobilization of either the primer or a primer extension product, such as amplified DNA, on a solid support.

"Probe" refers to an oligonucleotide which binds through complementary base pairing to a subsequence of a target nucleic acid. It will be understood by one of skill in the art that probes will typically substantially bind target sequences lacking complete complementarity with the probe sequence depending upon the stringency of the hybridization conditions. The probes are preferably directly labeled as with isotopes or indirectly labeled such as with biotin to which a streptavidin complex can later bind. By assaying for the presence or absence of the probe, one can detect the presence or absence of the target.

"Subsequence" refers to a sequence of nucleic acids that comprise a part of a longer sequence of nucleic acids.

The term "target region" refers to a region of a nucleic acid to be analyzed and can include a polymorphic region.

The term alkyl, as used herein, unless otherwise specified, refers to a saturated straight, branched, or cyclic, primary, secondary, or tertiary hydrocarbon typically of C_1 to C_{10} , and specifically includes methyl, CF_3 , CCl_3 , CFCl_2 , CF_2Cl , ethyl, CH_2CF_3 , CF_2CF_3 , propyl, isopropyl, cyclopropyl, butyl, isobutyl, *t*-butyl, pentyl, cyclopentyl, isopentyl, neopentyl, hexyl, isohexyl, cyclohexyl, cyclohexylmethyl, 3-methylpentyl,

2,2-dimethylbutyl, and 2,3-dimethylbutyl. The term includes both substituted and unsubstituted alkyl groups, and particularly includes halogenated alkyl groups, and even more particularly fluorinated alkyl groups. Moieties with which the alkyl group can be substituted are selected from the group consisting of halogen (fluoro, chloro, bromo or iodo), hydroxyl, amino, alkylamino, arylamino, alkoxy, aryloxy, nitro, cyano, sulfonic acid, sulfate, phosphonic acid, phosphate, or phosphonate, either unprotected, or protected as necessary, as known to those skilled in the art, for example, as taught in Greene, *et al.*, Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991, hereby incorporated by reference.

The term lower alkyl, as used herein, and unless otherwise specified, refers to a C₁ to C₄ saturated straight, branched, or if appropriate, a cyclic (for example, cyclopropyl) alkyl group, including both substituted and unsubstituted forms. Unless otherwise specifically stated in this application, when alkyl is a suitable moiety, lower alkyl is preferred. Similarly, when alkyl or lower alkyl is a suitable moiety, unsubstituted alkyl or lower alkyl is preferred.

The term alkylamino or arylamino refers to an amino group that has one or two alkyl or aryl substituents, respectively.

The term “protected” as used herein and unless otherwise defined refers to a group that is added to an oxygen, nitrogen, or phosphorus atom to prevent its further reaction or for other purposes. A wide variety of oxygen and nitrogen protecting groups are known to those skilled in the art of organic synthesis.

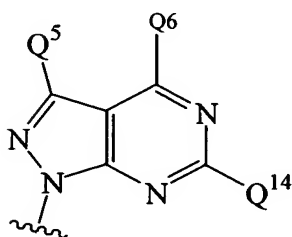
The term aryl, as used herein, and unless otherwise specified, refers to phenyl, biphenyl, or naphthyl, and preferably phenyl. The term includes both substituted and unsubstituted moieties. The aryl group can be substituted with one or more moieties selected from the group consisting of halogen (fluoro, chloro, bromo or iodo), hydroxyl, amino, alkylamino, arylamino, alkoxy, aryloxy, nitro, cyano, sulfonic acid, sulfate, phosphonic acid, phosphate, or phosphonate, either unprotected, or protected as necessary, as known to those skilled in the art, for example, as taught in Greene, *et al.*, Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

The term alkaryl or alkylaryl refers to an alkyl group with an aryl substituent. The term aralkyl or arylalkyl refers to an aryl group with an alkyl substituent.

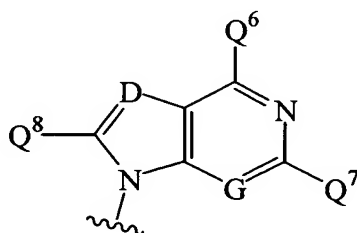
The term halo, as used herein, includes chloro, bromo, iodo, and fluoro.

The term base refers to any purine or pyrimidine base including, but not limited to, guanine, adenine, hypoxanthine, 2,6-diaminopurine, 6-chloropurine, N⁶-alkylpurines, N⁶-acylpurines (wherein acyl is C(O)(alkyl, aryl, alkylaryl, or arylalkyl), N⁶-benzylpurine, N⁶-halopurine, N⁶-vinylpurine, N⁶-acetylenic purine, N⁶-acyl purine, N⁶-hydroxyalkyl purine, N⁶-thioalkyl purine, N²-alkylpurines, N²-alkyl-6-thiopurines, thymine, cytosine, 5-fluorocytosine, 5-methylcytosine, 6-azapyrimidine, including 6-azacytosine, 2- and/or 4-mercaptopyrimidine, uracil, 5-halouracil, including 5-fluorouracil, C⁵-alkylpyrimidines, C⁵-benzylpyrimidines, C⁵-halopyrimidines, C⁵-vinylpyrimidine, C⁵-acetylenic pyrimidine, C⁵-acyl pyrimidine, C⁵-hydroxyalkyl purine, C⁵-amidopyrimidine, C⁵-cyanopyrimidine, C⁵-nitropyrimidine, C⁵-amino-pyrimidine, N²-alkylpurines, N²-alkyl-6-thiopurines, 5-azacytidinyl, 5-aza-uracilyl, triazolopyridinyl, imidazolopyridinyl, pyrrolopyrimidinyl, pyrazolopyrimidinyl,

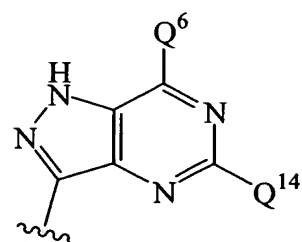
a base of the formula:



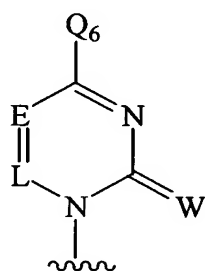
(i)



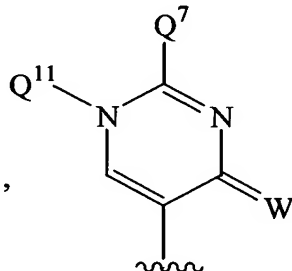
(ii)



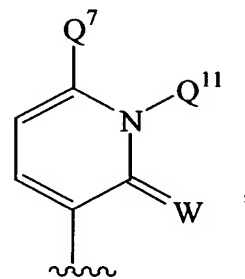
(iii)



(iv)



(v)



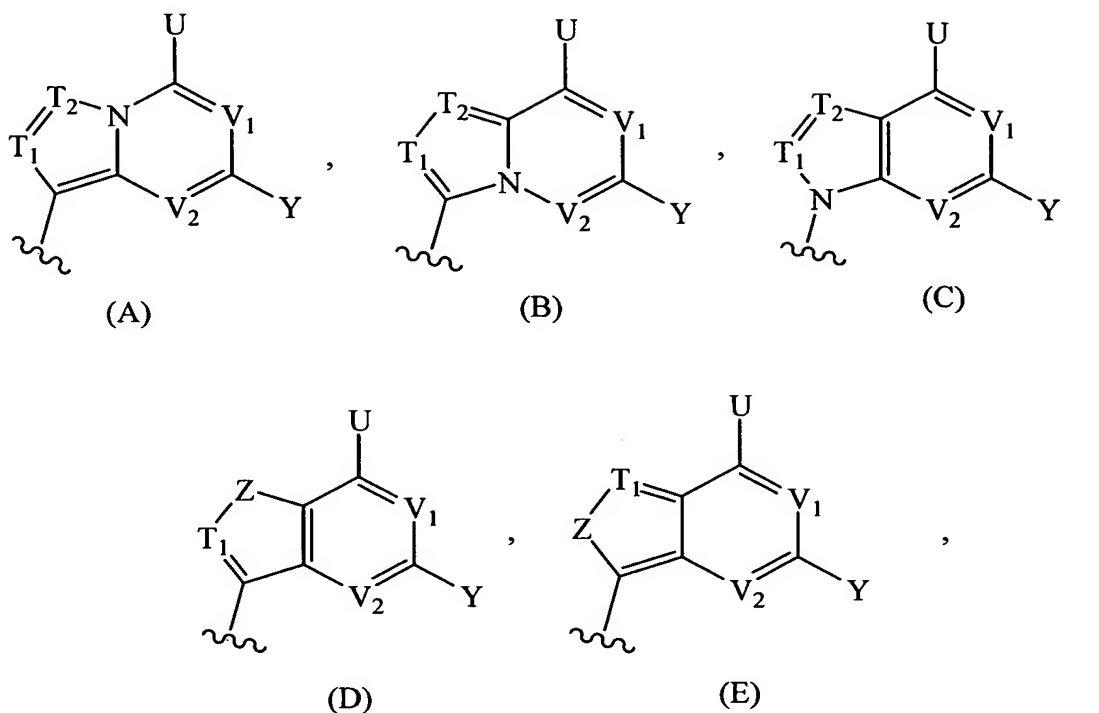
(vi)

wherein:

G and L are each independently CH or N;

- D is N, CH, C-CN, C-NO₂, C-C₁₋₃ alkyl, C-NHCONH₂, C-CONQ¹¹Q¹¹, C-CSNQ¹¹Q¹¹, CCOOQ¹¹, C-C(=NH)NH₂, C-hydroxy, C-C₁₋₃alkoxy, C-amino, C-C₁₋₄alkyl-amino, C-di(C₁₋₄ alkyl)amino, C-halogen, C-(1,3-oxazol-2-yl), C-(1,3-thiazol-2-yl), or C-(imidazol-2-yl); wherein alkyl is unsubstituted or substituted with one to three groups independently selected from halogen, amino, hydroxy, carboxy, and C₁₋₃ alkoxy;
- E is N or CQ⁵;
- W is O, S, or NR;
- R is H, OH, alkyl;
- Q⁶ is H, OH, SH, NH₂, C₁₋₄ alkylamino, di(C₁₋₄ alkyl)amino, C₃₋₆ cycloalkylamino, halogen, C₁₋₄ alkyl, C₁₋₄ alkoxy, or CF₃;
- Q⁵ is H, C₁₋₆ alkyl, C₂₋₆ alkenyl, C₂₋₆ alkynyl, C₁₋₄ alkylamino, CF₃, halogen, N, CN, NO₂, NHCONH₂, CONQ¹¹Q¹¹, CSNQ¹¹Q¹¹, COOQ¹¹, C(=NH)NH₂, hydroxy, C₁₋₃alkoxy, amino, C₁₋₄ alkylamino, di(C₁₋₄ alkyl)amino, halogen, 1,3-oxazol-2-yl, 1,3-thiazol-2-yl, or imidazol-2-yl; wherein alkyl is unsubstituted or substituted with one to three groups independently selected from halogen, amino, hydroxy, carboxy, and C₁₋₃ alkoxy;
- Q⁷ and Q¹⁴ are each independently selected from the group consisting of H, CF₃, OH, SH, OR, SR C₁₋₄ alkyl, amino, C₁₋₄ alkylamino, C₃₋₆ cycloalkylamino, and di(C₁₋₄ alkyl)amino;
- Q¹¹ is independently H or C₁₋₆ alkyl;
- Q⁸ is H, halogen, CN, carboxy, C₁₋₄ alkyloxycarbonyl, N₃, amino, C₁₋₄ alkylamino, di(C₁₋₄ alkyl)amino, hydroxy, C₁₋₆ alkoxy, C₁₋₆ alkylthio, C₁₋₆ alkylsulfonyl, (C₁₋₄ alkyl)₀₋₂aminomethyl, NH₂, CN, NO₂, C₁₋₃ alkyl, NHCONH₂, CONQ¹¹Q¹¹, CSNQ¹¹Q¹¹, COOQ¹¹, C(=NH)NH₂, 1,3-oxazol-2-yl, 1,3-thiazol-2-yl, or imidazol-2-yl, wherein alkyl is unsubstituted or substituted with one to three groups independently selected from halogen, amino, hydroxy, carboxy, and C₁₋₃ alkoxy;

a base of the formula:



wherein:

T₁ and T₂ are independently selected from N, CH, or C-Q¹⁶;

Q¹⁶, U, and Y are independently selected from is H, OH, substituted or unsubstituted alkyl, substituted or unsubstituted alkenyl, substituted or unsubstituted alkynyl, cycloalkyl, CO-alkyl, CO-aryl, CO-alkoxyalkyl, chloro, bromo, fluoro, iodo, OR⁴, NR⁴R⁵ or SR⁵, Br-vinyl, -O-alkyl, -O-alkenyl, -O-alkynyl, -O-aryl, -O-aralkyl, -O-acyl, -O-cycloalkyl, NH₂, NH-alkyl, N-dialkyl, NH-acyl, N-aryl, N-aralkyl, NH-cycloalkyl, SH, S-alkyl, S-acyl, S-aryl, S-cycloalkyl, S-aralkyl, CN, N₃, COOH, CONH₂, CO₂-alkyl, CONH-alkyl, CON-dialkyl, OH, CF₃, CH₂OH, (CH₂)_mOH, (CH₂)_mNH₂, (CH₂)_mCOOH, (CH₂)_mCN, (CH₂)_mNO₂, (CH₂)_mCONH₂, C₁₋₄ alkylamino, di(C₁₋₄ alkyl)amino, C₃₋₆ cycloalkylamino, C₁₋₄ alkoxy, C₁₋₄ alkoxycarbonyl, C₁₋₆ alkylthio, C₁₋₆ alkylsulfonyl, (C₁₋₄ alkyl)₀₋₂aminomethyl, or -NHC(=NH)NH₂;

R⁴ and R⁵ are independently selected from hydrogen, acyl (including lower acyl), or alkyl (including but not limited to methyl, ethyl, propyl and cyclopropyl);

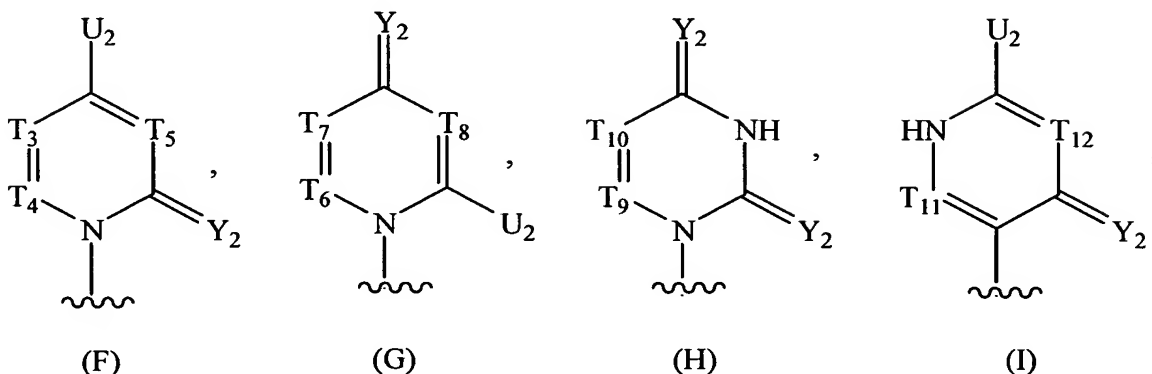
m is 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10;

Z is S, SO, SO₂, C=O, or NQ²⁰;

Q²⁰ is H or alkyl; and

V₁ and V₂ are independently selected from CH or N;

a base of the formula:



wherein:

T₃ and T₄ are independently selected from N or CQ²²;

Q²² is independently selected from H, OH, substituted or unsubstituted alkyl, substituted or unsubstituted alkenyl, substituted or unsubstituted alkynyl, cycloalkyl, CO-alkyl, CO-aryl, CO-alkoxyalkyl, chloro, bromo, fluoro, iodo, OR⁴, NR⁴R⁵ or SR⁵, Br-vinyl, -O-alkyl, -O-alkenyl, -O-alkynyl, -O-aryl, -O-aralkyl, -O-acyl, -O-cycloalkyl, NH₂, NH-alkyl, N-dialkyl, NH-acyl, N-aryl, N-aralkyl, NH-cycloalkyl, SH, S-alkyl, S-acyl, S-aryl, S-cycloalkyl, S-aralkyl, CN, N₃, COOH, CONH₂, CO₂-alkyl, CONH-alkyl, CON-dialkyl, OH, CF₃, CH₂OH, (CH₂)_mOH, (CH₂)_mNH₂, (CH₂)_mCOOH, (CH₂)_mCN, (CH₂)_mNO₂, (CH₂)_mCONH₂, C₁₋₄ alkylamino, di(C₁₋₄ alkyl)amino, C₃₋₆ cycloalkylamino, C₁₋₄ alkoxy, C₁₋₄ alkoxycarbonyl, C₁₋₆ alkylthio, C₁₋₆ alkylsulfonyl, (C₁₋₄ alkyl)₀₋₂aminomethyl, or -NHC(=NH)NH₂;

T₅ is NH;

R⁴ and R⁵ are independently selected from hydrogen, acyl (including lower acyl), or alkyl (including but not limited to methyl, ethyl, propyl and cyclopropyl);

m is 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10;

T₆, T₇, T₈, T₉, T₁₀, T₁₁, and T₁₂ are independently selected from N or CH;

U₂ is H, straight chained, branched or cyclic alkyl, CO-alkyl, CO-aryl, CO-alkoxyalkyl, chloro, bromo, fluoro, iodo, OR⁴, NR⁴R⁵ or SR⁵;

Y₂ is O, S, NH, NR or CQ²⁴Q²⁶ where R is H, OH, or alkyl;

5 Q²⁴ and Q²⁶ are independently selected from H, alkyl, straight chained, branched or cyclic alkyl, CO-alkyl, CO-aryl, CO-alkoxyalkyl, chloro, bromo, fluoro, iodo, OR⁴, NR⁴R⁵ or SR⁵.

Further examples of purine bases include, but are not limited to, guanine, adenine, hypoxanthine, 2,6-diaminopurine, 6-chloropurine, and 6-N-methylamino
10 purine. Functional oxygen and nitrogen groups on the base can be protected as necessary or desired. Suitable protecting groups are well known to those skilled in the art, and include trimethylsilyl, dimethylhexylsilyl, *t*-butyldimethylsilyl, and *t*-butyldiphenylsilyl, trityl, alkyl groups, and acyl groups such as acetyl and propionyl, methanesulfonyl, and *p*-toluenesulfonyl.

15 The term acyl or O-linked ester refers to a group of the formula C(O)R', wherein R' is an straight, branched, or cyclic alkyl (including lower alkyl), amino acid, aryl including phenyl, alkaryl, aralkyl including benzyl, alkoxyalkyl including methoxymethyl, aryloxyalkyl such as phenoxymethyl; or substituted alkyl (including lower alkyl), aryl including phenyl optionally substituted with chloro, bromo, fluoro,
20 iodo, C₁ to C₄ alkyl or C₁ to C₄ alkoxy, sulfonate esters such as alkyl or aralkyl sulphonyl including methanesulfonyl, the mono, di or triphosphate ester, trityl or monomethoxy-trityl, substituted benzyl, alkaryl, aralkyl including benzyl, alkoxyalkyl including methoxymethyl, aryloxyalkyl such as phenoxymethyl. Aryl groups in the esters optimally comprise a phenyl group. In particular, acyl groups include acetyl,
25 trifluoroacetyl, methylacetyl, cyclopropylacetyl, propionyl, butyryl, hexanoyl, heptanoyl, octanoyl, neo-heptanoyl, phenylacetyl, 2-acetoxy-2-phenylacetyl, diphenylacetyl, α -methoxy- α -trifluoromethyl-phenylacetyl, bromoacetyl, 2-nitro-benzeneacetyl, 4-chloro-benzeneacetyl, 2-chloro-2,2-diphenylacetyl, 2-chloro-2-phenylacetyl, trimethylacetyl, chlorodifluoroacetyl, perfluoroacetyl, fluoroacetyl, bromodifluoroacetyl, methoxyacetyl,
30 2-thiopheneacetyl, chlorosulfonylacetyl, 3-methoxyphenylacetyl, phenoxyacetyl, tert-butylacetyl, trichloroacetyl, monochloro-acetyl, dichloroacetyl, 7H-dodecafluoro-

heptanoyl, perfluoro-heptanoyl, 7H-dodeca-fluoroheptanoyl, 7-chlorododecafluoro-heptanoyl, 7-chloro-dodecafluoro-heptanoyl, 7H-dodecafluoroheptanoyl, 7H-dodecafluoroheptanoyl, nona-fluoro-3,6-dioxa-heptanoyl, nonafluoro-3,6-dioxaheptanoyl, perfluoroheptanoyl, methoxybenzoyl, methyl 3-amino-5-phenylthiophene-2-carboxyl, 3,6-dichloro-2-methoxy-benzoyl, 4-(1,1,2,2-tetrafluoro-ethoxy)-benzoyl, 2-bromopropionyl, omega-aminocapryl, decanoyl, n-pentadecanoyl, stearyl, 3-cyclopentylpropionyl, 1-benzene-carboxyl, O-acetylmandelyl, pivaloyl acetyl, 1-adamantane-carboxyl, cyclohexane-carboxyl, 2,6-pyridinedicarboxyl, cyclopropane-carboxyl, cyclobutane-carboxyl, perfluorocyclohexyl carboxyl, 4-methylbenzoyl, chloromethyl isoxazolyl carbonyl, perfluorocyclohexyl carboxyl, crotonyl, 1-methyl-1H-indazole-3-carbonyl, 2-propenyl, isovaleryl, 1-pyrrolidinecarbonyl, 4-phenylbenzoyl.

As used herein, the term “substantially free of” or “substantially in the absence of” refers to a nucleoside composition that includes at least 95% to 98% by weight, and even more preferably 99% to 100% by weight, of the designated enantiomer of that nucleoside. In a preferred embodiment, in the methods and compounds of this invention, the compounds are substantially free of enantiomers.

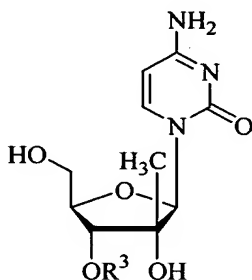
Similarly, the term “isolated” refers to a nucleoside composition that includes at least 95% to 98% by weight, and even more preferably 99% to 100% by weight, of the nucleoside, the remainder comprising other chemical species or enantiomers.

The term host, as used herein, refers to an unicellular or multicellular organism in which the virus can replicate, including cell lines and animals, and preferably a human. Alternatively, the host can be carrying a part of the *Flaviviridae* viral genome, whose replication or function can be altered by the compounds of the present invention. The term host specifically refers to infected cells, cells transfected with all or part of the *Flaviviridae* genome and animals, in particular, primates (including chimpanzees) and humans. In most animal applications of the present invention, the host is a human patient. Veterinary applications, in certain indications, however, are clearly anticipated by the present invention (such as chimpanzees).

II. 2'-Branched Nucleosides and Prodrugs Thereof

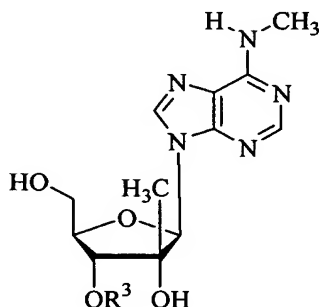
In the broadest embodiment, the present invention provides a method to treat a Flaviviridae infection that is resistant to a 2'-branched nucleoside. In one embodiment, the 2'-branched nucleoside is a purine, or a purine derivative such as a pyrrolo-purine. In a preferred embodiment, the 2'-branched nucleoside is a pyrimidine. Other sub-embodiments include the 2'-alkyl and 2'-methyl-branched pyrimidine nucleosides, including the 2'-branched uracil and 2'-branched cytosine nucleosides. In another embodiment, the 2'-branched nucleoside is a purine. Other sub-embodiments include the 2'-alkyl and 2'-methyl-branched purine nucleosides, including the 2'-branched adenosine nucleosides and the 2'-branched 6-N-methylamino purine nucleosides.

In a specific embodiment, the 2'-branched pyrimidine nucleoside is represented by the compound β -D-2'-CH₃-riboC, which is represented by the formula:



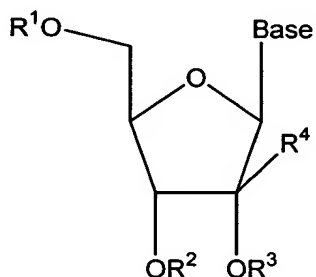
wherein R³ is H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; lipid (including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R³ is H or phosphate when administered *in vivo*.

In another specific embodiment, the 2'-branched purine nucleoside is represented by the compound β -D-2'-CH₃-ribo-6-N-methylaminopurine, which is represented by the formula:



5 wherein R³ is H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; lipid (including a phospholipid); amino acid;
 10 carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R³ is H or phosphate when administered *in vivo*.

In one embodiment of the invention, the 2'-branched nucleoside is of the general formula:



15 or its pharmaceutically acceptable prodrug and/or salt, wherein

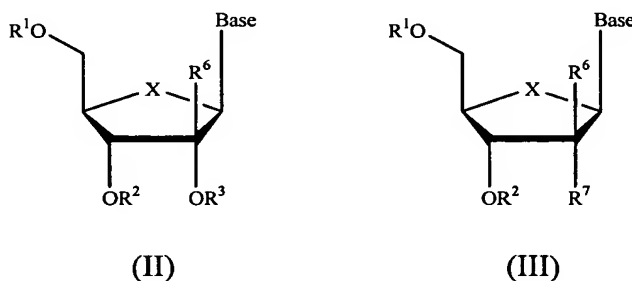
R¹, R², and R³ are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with
 20 one or more substituents as described in the definition of aryl given herein; lipid

(including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R^1 , R^2 or R^3 is independently H or phosphate when administered *in vivo*; and

R^4 is hydrogen, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, $-C(O)O(alkyl)$, $-C(O)O(lower\ alkyl)$, $-O(acyl)$, $-O(lower\ acyl)$, $-O(alkyl)$, $-O(lower\ alkyl)$, $-O(alkenyl)$, chloro, bromo, fluoro, iodo, NO_2 , NH_2 , $-NH(lower\ alkyl)$, $-NH(acyl)$, $-N(lower\ alkyl)_2$, or $-N(acyl)_2$;

and Base is a purine or pyrimidine as further described herein.

In another embodiment of the invention, the 2'-branched nucleoside is of the general formula:



or its pharmaceutically acceptable prodrug and/or salt, wherein

Base is a purine or pyrimidine base as defined herein;

R^1 , R^2 and R^3 are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate); straight chained, branched or cyclic alkyl (including lower alkyl); acyl (including lower acyl); CO-alkyl, CO-aryl, CO-alkoxyalkyl, CO-aryloxyalkyl, CO-substituted aryl, sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; alkylsulfonyl, arylsulfonyl, aralkylsulfonyl, lipid (including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R^1 , R^2 or R^3 is independently H or phosphate when administered *in vivo*;

R^6 is alkyl (including lower alkyl and halogenated alkyl), CH_3 , CF_3 , azido, cyano, alkenyl, alkynyl, Br-vinyl, 2-Br-ethyl, $-C(O)O(alkyl)$, $-C(O)O(lower\ alkyl)$, $-O(acyl)$,

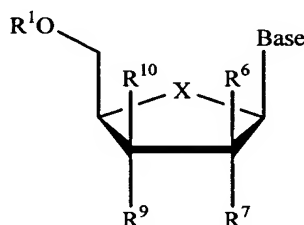
-O(lower acyl), -O(alkyl), -O(lower alkyl), -O(alkenyl), CF₃, chloro, bromo, fluoro, iodo, NO₂, NH₂, -NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂; and

R⁷ is hydrogen, OR³, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, -C(O)O(alkyl), -C(O)O(lower alkyl), -O(acyl), -O(lower acyl), -O(alkyl), -O(lower alkyl), -O(alkenyl), fluorine, chlorine, bromine, iodine, NO₂, NH₂, -NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂; and

X is O, S, SO₂ or CH₂.

and Base is a purine or pyrimidine as further described herein.

In yet another embodiment the invention, the 2'-branched nucleoside is of the general formula:



(IV)

wherein:

Base is a purine or pyrimidine base as defined herein;

R⁶ is alkyl (including lower alkyl and halogenated alkyl), CH₃, CF₃, azido, cyano, alkenyl, alkynyl, Br-vinyl, 2-Br-ethyl, -C(O)O(alkyl), -C(O)O(lower alkyl), -O(acyl), -O(lower acyl), -O(alkyl), -O(lower alkyl), -O(alkenyl), CF₃, fluoro, chloro, bromo, iodo, NO₂, NH₂, -NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂;

R⁷ is OR², hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, halo-vinyl, -C(O)O(alkyl), -C(O)O(lower alkyl), -O(acyl), -O(lower acyl), -O(alkyl), -O(lower alkyl), -O(alkenyl), fluorine, chlorine, bromine, iodine, NO₂, NH₂, -NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂;

R⁹ is hydrogen, OR³, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, -C(O)O(alkyl), -C(O)O(lower alkyl), -O(acyl), -O(lower acyl),

-O(alkyl), -O(lower alkyl), -O(alkenyl), chlorine, bromine, iodine, NO₂, NH₂,
-NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, -N(acyl)₂;

R¹⁰ is H, alkyl (including lower alkyl), fluorine, chlorine, bromine or iodine;

R¹, R² and R³ are independently H, phosphate (including mono-, di- or triphosphate and
a stabilized phosphate); straight chained, branched or cyclic alkyl (including lower
alkyl); acyl (including lower acyl); CO-alkyl, CO-aryl, CO-alkoxyalkyl, CO-
aryloxyalkyl, CO-substituted aryl, sulfonate ester (including alkyl or arylalkyl
sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally
substituted with one or more substituents as described in the definition of aryl given
herein; alkylsulfonyl, arylsulfonyl, aralkylsulfonyl, lipid (including a phospholipid);
amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable
leaving group that provides a compound wherein R¹, R² or R³ is independently H or
phosphate when administered *in vivo*; and

X is O, S, SO₂ or CH₂.

General Synthesis of 2'-Branched Nucleosides:

1. Glycosylation of the nucleobase with an appropriately modified sugar

The key starting material for this process can be an appropriately substituted
sugar with a 2'-OH and 2'-H, with the appropriate leaving group (LG), for example an
acyl group or a chloro, bromo, fluoro or iodo. The sugar can be purchased or can be
prepared by any known means including standard epimerization, substitution, oxidation
and reduction techniques. The substituted sugar can then be oxidized with the
appropriate oxidizing agent in a compatible solvent at a suitable temperature to yield the
2'-modified sugar. Possible oxidizing agents are Jones reagent (a mixture of chromic
acid and sulfuric acid), Collins's reagent (dipyridine Cr(VI) oxide, Corey's reagent
(pyridinium chlorochromate), pyridinium dichromate, acid dichromate, potassium
permanganate, MnO₂, ruthenium tetroxide, phase transfer catalysts such as chromic acid
or permanganate supported on a polymer, Cl₂-pyridine, H₂O₂-ammonium molybdate,
NaBrO₂-CAN, NaOCl in HOAc, copper chromite, copper oxide, Raney nickel,

palladium acetate, Meerwin-Pondorf-Verley reagent (aluminum *t*-butoxide with another ketone) and *N*-bromosuccinimide.

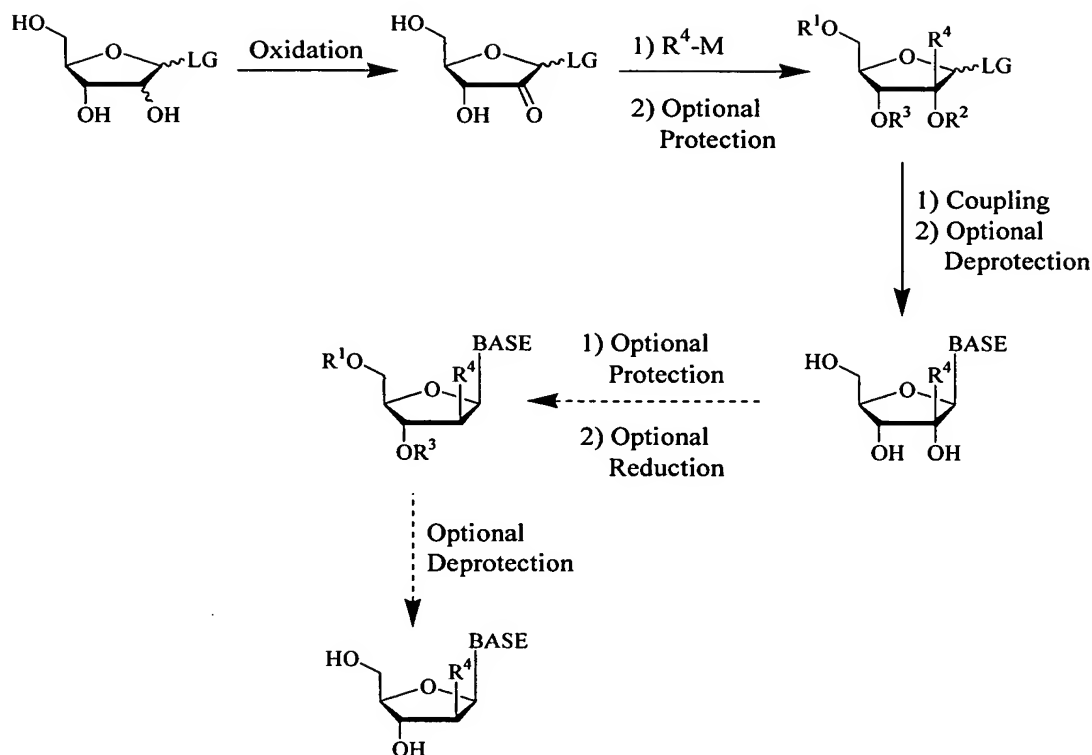
Then coupling of an organometallic carbon nucleophile, such as a Grignard reagent, an organolithium, lithium dialkylcopper or R^4-SiMe_3 (wherein R^4 is defined below) in TBAF with the ketone with the appropriate non-protic solvent at a suitable temperature, yields the 2'-alkylated sugar. The alkylated sugar can be optionally protected with a suitable protecting group, preferably with an acyl or silyl group, by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

The optionally protected sugar can then be coupled to the BASE by methods well known to those skilled in the art, as taught by Townsend Chemistry of Nucleosides and Nucleotides, Plenum Press, 1994. For example, an acylated sugar can be coupled to a silylated base with a lewis acid, such as tin tetrachloride, titanium tetrachloride or trimethylsilyltriflate in the appropriate solvent at a suitable temperature. Alternatively, a halo-sugar can be coupled to a silylated base with the presence of trimethylsilyltriflate.

Subsequently, the nucleoside can be deprotected by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

In a particular embodiment, the 2'-C-branched ribonucleoside is desired. The synthesis of a ribonucleoside is shown in **Scheme 1**. Alternatively, the deoxyribonucleoside can be used. To obtain these nucleosides, the formed ribonucleoside can optionally be protected by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991, and then the 2'-OH can be reduced with a suitable reducing agent. Optionally, the 2'-hydroxyl can be activated to facilitate reduction; i.e. via the Barton reduction.

Scheme 1:



wherein:

LG is a leaving group;

- 5 R¹, R², and R³ are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; lipid (including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R¹, R² or R³ is independently H or phosphate when administered *in vivo*; and
- 10 R⁴ is hydrogen, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, -C(O)O(alkyl), -C(O)O(lower alkyl), -O(acyl), -O(lower acyl), -O(alkyl), -O(lower alkyl), -O(alkenyl), chloro, bromo, fluoro, iodo, NO₂, NH₂, -NH(lower alkyl), -NH(acyl), -N(lower alkyl)₂, or -N(acyl)₂.
- 15

2. Modification of a pre-formed nucleoside

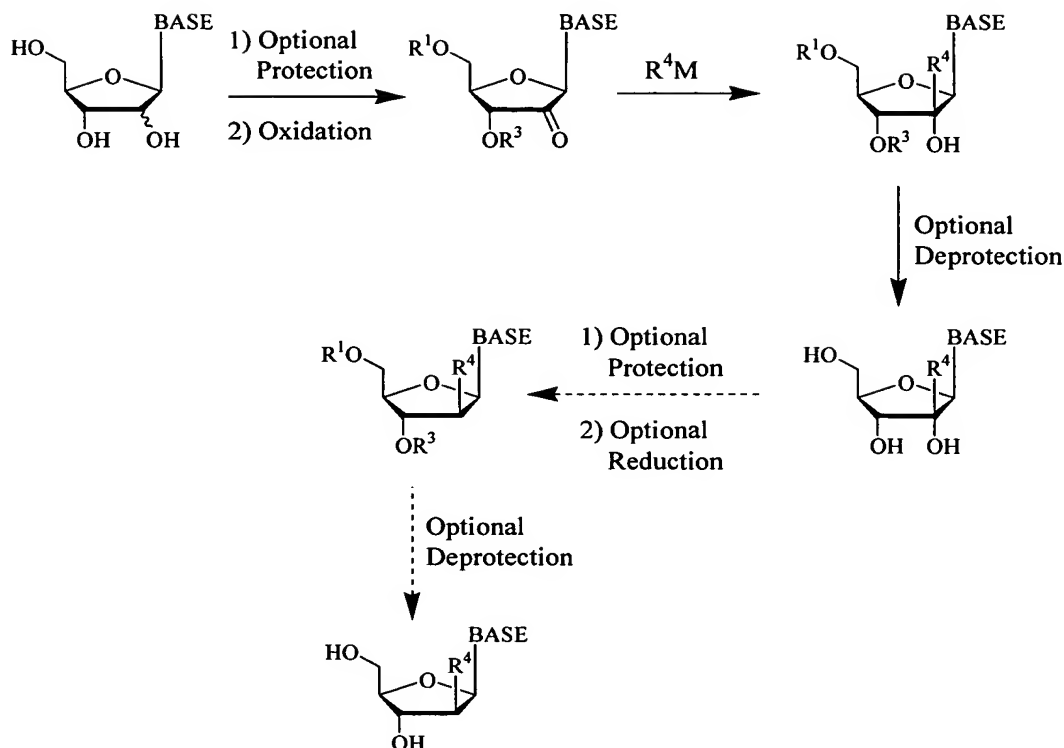
The key starting material for this process can be an appropriately substituted nucleoside with a 2'-OH and 2'-H. The nucleoside can be purchased or can be prepared by any known means including standard coupling techniques. The nucleoside can be optionally protected with suitable protecting groups, preferably with acyl or silyl groups, by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

The appropriately protected nucleoside can then be oxidized with the appropriate oxidizing agent in a compatible solvent at a suitable temperature to yield the 2'-modified sugar. Possible oxidizing agents are Jones reagent (a mixture of chromic acid and sulfuric acid), Collins's reagent (dipyridine Cr(VI) oxide), Corey's reagent (pyridinium chlorochromate), pyridinium dichromate, acid dichromate, potassium permanganate, MnO₂, ruthenium tetroxide, phase transfer catalysts such as chromic acid or permanganate supported on a polymer, Cl₂-pyridine, H₂O₂-ammonium molybdate, NaBrO₂-CAN, NaOCl in HOAc, copper chromite, copper oxide, Raney nickel, palladium acetate, Meerwin-Pondorf-Verley reagent (aluminum *t*-butoxide with another ketone) and *N*-bromosuccinimide.

Subsequently, the nucleoside can be deprotected by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

In a particular embodiment, the 2'-C-branched ribonucleoside is desired. The synthesis of a ribonucleoside is shown in **Scheme 2**. Alternatively, deoxyribonucleoside can be used. To obtain these nucleosides, the formed ribonucleoside can optionally be protected by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991, and then the 2'-OH can be reduced with a suitable reducing agent. Optionally, the 2'-hydroxyl can be activated to facilitate reduction; i.e. via the Barton reduction.

Scheme 2



wherein:

R^1 and R^3 are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; lipid (including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R^1 or R^3 is independently H or phosphate when administered *in vivo*; and

R^4 is hydrogen, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, $-C(O)O(alkyl)$, $-C(O)O(lower\ alkyl)$, $-O(acyl)$, $-O(lower\ acyl)$, $-O(alkyl)$, $-O(lower\ alkyl)$, $-O(alkenyl)$, chloro, bromo, fluoro, iodo, NO_2 , NH_2 , $-NH(lower\ alkyl)$, $-NH(acyl)$, $-N(lower\ alkyl)_2$, or $-N(acyl)_2$.

General Synthesis of 2'-Branched Pyrimidine Nucleoside

1. Glycosylation of the pyrimidine with an appropriately modified sugar

A representative general method for the preparation of 2'-branched pyrimidine nucleoside is outlined in **Scheme 3**. This scheme illustrates the 2'-branched pyrimidine nucleoside in the β -D-ribo configuration. Alternatively, one skilled in the art could modify the general scheme to produce 2'- β -L-pyrimidine nucleoside. The key starting material for this process can be an appropriately substituted sugar with a 2'-OH and 2'-H, with the appropriate leaving group (LG), for example an acyl group or a chloro, bromo, fluoro or iodo. The sugar can be purchased or can be prepared by any known means including standard epimerization, substitution, oxidation and reduction techniques. The substituted sugar can then be oxidized with the appropriate oxidizing agent in a compatible solvent at a suitable temperature to yield the 2'-modified sugar. Possible oxidizing agents are Jones reagent (a mixture of chromic acid and sulfuric acid), Collins's reagent (dipyridine Cr(VI) oxide, Corey's reagent (pyridinium chlorochromate), pyridinium dichromate, acid dichromate, potassium permanganate, MnO₂, ruthenium tetroxide, phase transfer catalysts such as chromic acid or permanganate supported on a polymer, Cl₂-pyridine, H₂O₂-ammonium molybdate, NaBrO₂-CAN, NaOCl in HOAc, copper chromite, copper oxide, Raney nickel, palladium acetate, Meerwin-Pondorf-Verley reagent (aluminum *t*-butoxide with another ketone) and *N*-bromosuccinimide.

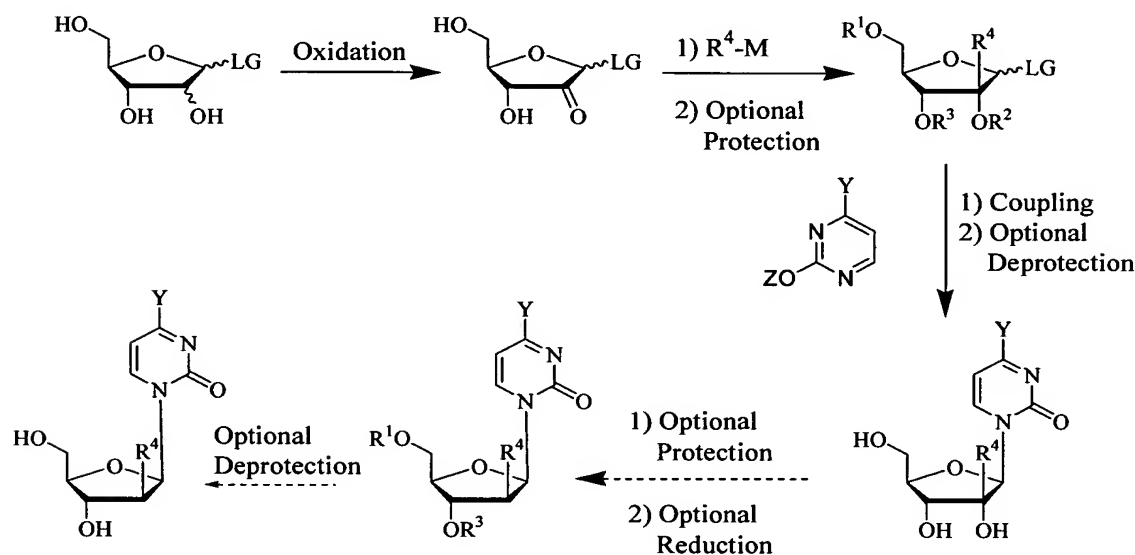
Then coupling of an organometallic carbon nucleophile, such as a Grignard reagent, an organolithium, lithium dialkylcopper or R⁴-SiMe₃ in TBAF with the ketone with the appropriate non-protic solvent at a suitable temperature, yields the 2'-alkylated sugar. The alkylated sugar can be optionally protected with a suitable protecting group, preferably with an acyl or silyl group, by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

The optionally protected sugar can then be coupled to a pyrimidine base by methods well known to those skilled in the art, as taught by Townsend Chemistry of Nucleosides and Nucleotides, Plenum Press, 1994. For example, an acylated sugar can be coupled to a silylated pyrimidine, such as cytidine, with a lewis acid, such as tin

tetrachloride, titanium tetrachloride or trimethylsilyltriflate in the appropriate solvent at a suitable temperature. Alternatively, a halo-sugar can be coupled to a silylated pyrimidine, such as a cytidine, with the presence of trimethylsilyltriflate.

5

Scheme 3



wherein:

R^1 , R^2 , and R^3 are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; lipid (including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R^1 , R^2 or R^3 is independently H or phosphate when administered *in vivo*; and

R^4 is hydrogen, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, $-C(O)O(alkyl)$, $-C(O)O(lower\ alkyl)$, $-O(acyl)$, $-O(lower\ acyl)$, $-O(alkyl)$, $-O(lower\ alkyl)$, $-O(alkenyl)$, chloro, bromo, fluoro, iodo, NO_2 , NH_2 , $-NH(lower\ alkyl)$, $-NH(acyl)$, $-N(lower\ alkyl)_2$, or $-N(acyl)_2$.

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2. *Modification of a pre-formed 2'-branched pyrimidine nucleoside*

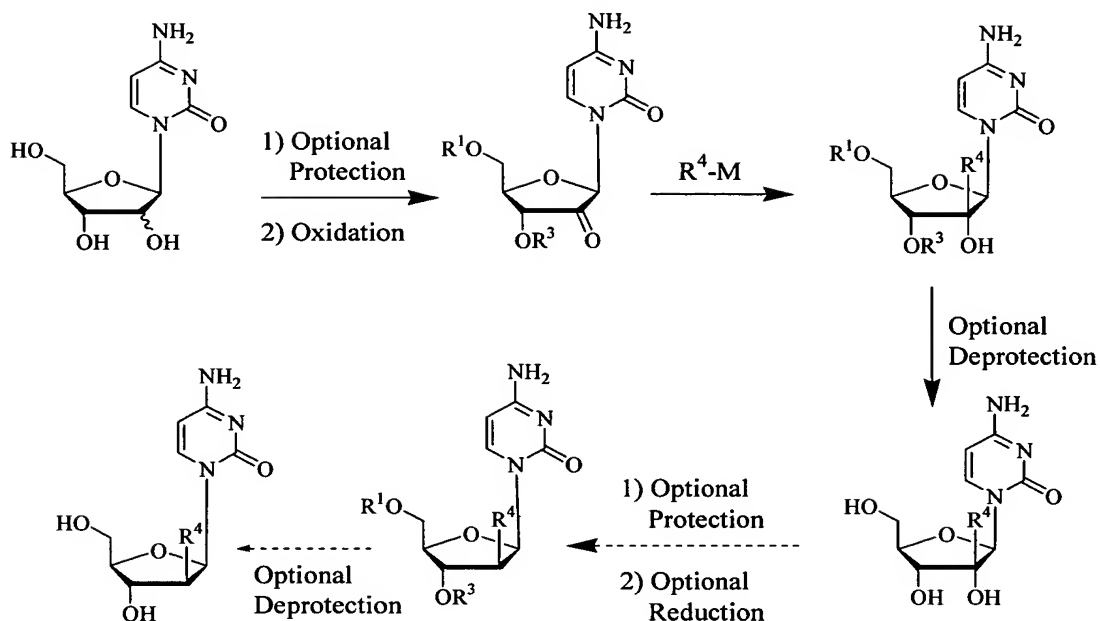
The key starting material for this process can be an appropriately substituted 2'-branched pyrimidine nucleoside with a 2'-OH and 2'-H. The 2'-branched pyrimidine nucleoside can be purchased or can be prepared by any known means including standard coupling techniques. The 2'-branched pyrimidine nucleoside can be optionally protected with suitable protecting groups, preferably with acyl or silyl groups, by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

The appropriately protected 2'-branched pyrimidine nucleoside can then be oxidized with the appropriate oxidizing agent in a compatible solvent at a suitable temperature to yield the 2'-modified sugar. Possible oxidizing agents are Jones reagent (a mixture of chromic acid and sulfuric acid), Collins's reagent (dipyridine Cr(VI) oxide, Corey's reagent (pyridinium chlorochromate), pyridinium dichromate, acid dichromate, potassium permanganate, MnO₂, ruthenium tetroxide, phase transfer catalysts such as chromic acid or permanganate supported on a polymer, Cl₂-pyridine, H₂O₂-ammonium molybdate, NaBrO₂-CAN, NaOCl in HOAc, copper chromite, copper oxide, Raney nickel, palladium acetate, Meerwin-Pondorf-Verley reagent (aluminum *t*-butoxide with another ketone) and *N*-bromosuccinimide.

Subsequently, the 2'-branched pyrimidine nucleoside can be deprotected by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

In a particular embodiment, the 2'-C-branched ribonucleoside is desired. The synthesis of a ribonucleoside is shown in **Scheme 4**. Alternatively, deoxyribonucleoside can be used. To obtain these nucleosides, the formed ribonucleoside can optionally be protected by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991, and then the 2'-OH can be reduced with a suitable reducing agent. Optionally, the 2'-hydroxyl can be activated to facilitate reduction; i.e. via the Barton reduction.

Scheme 4



wherein:

R^1 and R^3 are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; lipid (including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R^1 or R^3 is independently H or phosphate when administered *in vivo*; and

R^4 is hydrogen, hydroxy, alkyl (including lower alkyl), azido, cyano, alkenyl, alkynyl, Br-vinyl, $-C(O)O(alkyl)$, $-C(O)O(lower\ alkyl)$, $-O(acyl)$, $-O(lower\ acyl)$, $-O(alkyl)$, $-O(lower\ alkyl)$, $-O(alkenyl)$, chloro, bromo, fluoro, iodo, NO_2 , NH_2 , $-NH(lower\ alkyl)$, $-NH(acyl)$, $-N(lower\ alkyl)_2$, or $-N(acyl)_2$.

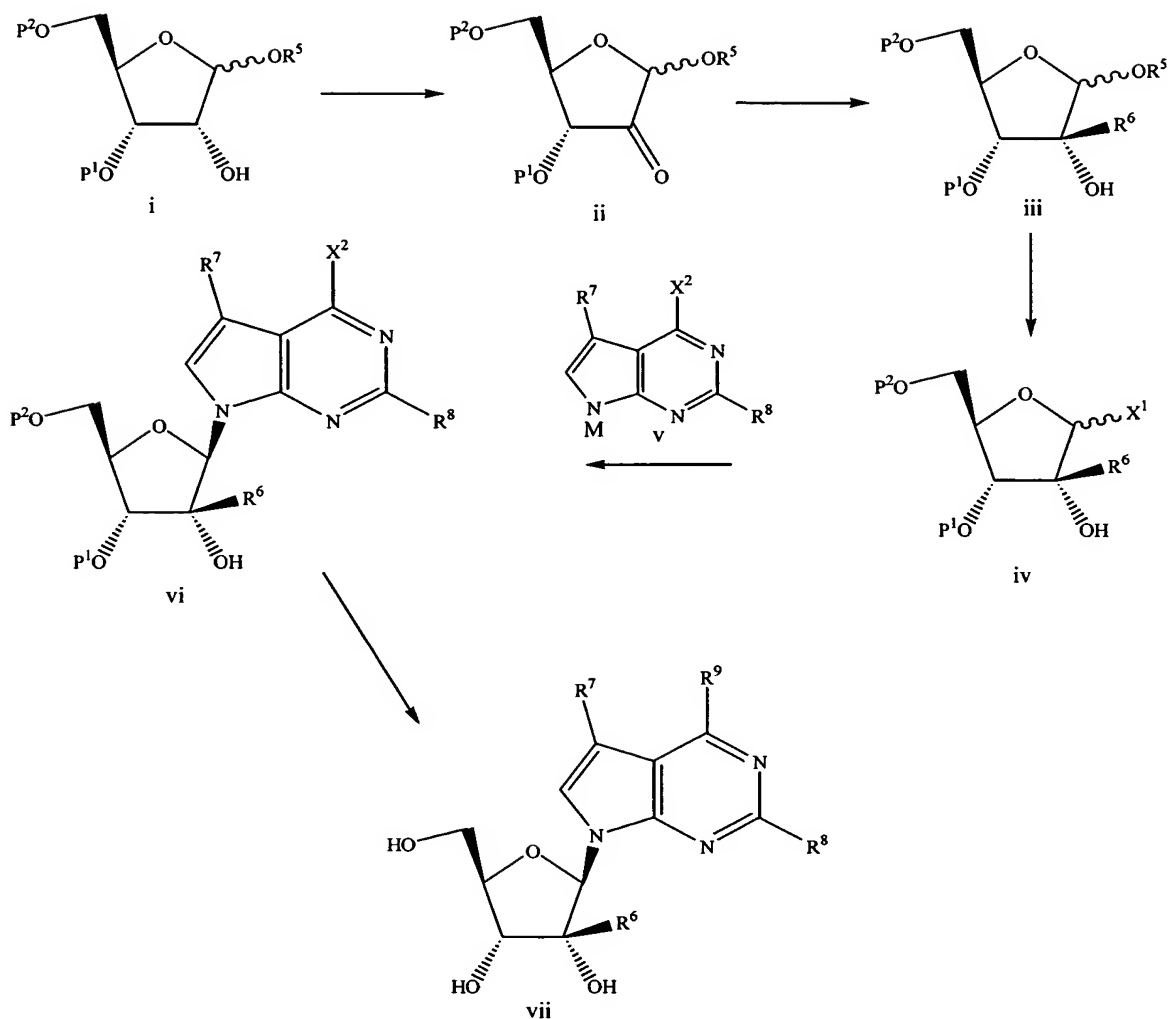
General Synthesis of 2'-Branched Purine Nucleoside

1. Glycosylation of the pyrimidine with an appropriately modified sugar

A representative general method for the preparation of 2'-branched purine nucleoside is outlined in **Scheme 5** below. This scheme illustrates the synthesis of 2'-
5 branched purine nucleoside in the β -D-ribo configuration. Alternative, it is well appreciated to those skilled in the art are able to prepare the β -L-ribo configuration using the appropriate starting material. The starting material is a 3,5-bis protected alkyl furanoside, such as methyl furanoside, of structural formula (i). The C-2 hydroxyl group is then oxidized with a suitable oxidizing agent, such as a chromium trioxide or chromate
10 reagent or Dess-Martin periodinane, or by Swern oxidation, to afford a C-2 ketone of structural formula (ii). Addition of a Grignard reagent, such as an alkyl, alkenyl, or alkynyl magnesium halide (for example, MeMgBr, EtMgBr, vinylMgBr, allylMgBr, and ethynylMgBr) or an alkyl, alkenyl, or alkynyl lithium, such as MeLi, across the carbonyl double bond of (ii) in a suitable organic solvent, such as tetrahydrofuran, diethyl ether,
15 and the like, affords the C-2 tertiary alcohol of structural formula (iii). A good leaving group (such as F, Cl, Br, and I) is next introduced at the C-1 (anomeric) position of the furanose sugar derivative by treatment of the furanoside of formula (iii) with a hydrogen halide in a suitable organic solvent, such as hydrogen bromide in acetic acid, to afford the intermediate furanosyl halide (iv). A C-sulfonate, such as methanesulfonate
20 (MeSO₂O-), trifluoromethanesulfonate (CF₃SO₂O-) or p-toluenesulfonate (-OTs), may also serve as a useful leaving group in the subsequent reaction to generate the glycosidic (nucleosidic) linkage. The nucleosidic linkage is constructed by treatment of the intermediate of structural formula (iv) with the metal salt (such as lithium, sodium, or potassium) of an appropriately substituted 1H-pyrrolo[2,3-d]pyrimidine (v), such as an
25 appropriately substituted 4-halo-1H-pyrrolo[2,3-d]pyrimidine, which can be generated *in situ* by treatment with an alkali hydride (such as sodium hydride), an alkali hydroxide (such as potassium hydroxide), an alkali carbonate (such as potassium carbonate), or an alkali hexamethyldisilazide (such as NaHMDS) in a suitable anhydrous organic solvent, such as acetonitrile, tetrahydrofuran, 1-methyl-2-pyrrolidinone, or N,N-dimethyl-
30 formamide (DMF). The displacement reaction can be catalyzed by using a phase-transfer catalyst, such as TDA-1 or triethylbenzylammonium chloride, in a two-phase system (solid-liquid or liquid-liquid). The optional protecting groups in the protected nucleoside

of structural formula (vi) are then cleaved following established deprotection methodologies, such as those described in T.W. Greene and P.G.M. Wuts, "Protective Groups in Organic Synthesis," P ed., John Wiley & Sons, 1999. Optional introduction of an amino group at the 4-position of the pyrrolo[2,3-d]pyrimidine nucleus is effected by treatment of the 4-halo intermediate (vi) with the appropriate amine, such as alcoholic ammonia or liquid ammonia, to generate a primary amine at the C-4 position (-NH₂), an alkylamine to generate a secondary amine (-NHR), or a dialkylamine to generate a tertiary amine (-NRR'). A 7H-pyrrolo[2,3-d]pyrimidin-4(3H)one compound may be derived by hydrolysis of (vi) with aqueous base, such as aqueous sodium hydroxide. Alcoholysis (such as methanolysis) of 1-6 affords a C-4 alkoxide (-OR), whereas treatment with an alkyl mercaptide affords a C-4 alkylthio (-SR) derivative. Subsequent chemical manipulations well-known to practitioners of ordinary skill in the art of organic/medicinal chemistry may be required to attain the desired compounds of the present invention.

Scheme 5



wherein:

P^1 and P^2 are indenpdently a protecting group; alternatively, P^1 and P^2 can come together to form a cyclic protecting group;

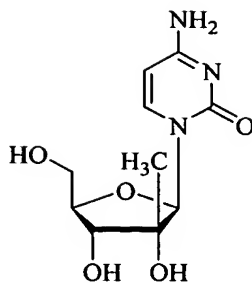
R^5 and R^6 are indenpdently alkyl group;

M is Li, Na, or K;

X^1 and X^2 are indenpdently F, Cl, Br, or I;

R^7 , R^8 , and R^9 are indenpdently hydrogen, hydroxyl, halogen, alkoxy, amino, alkylamino, or alkyl.

Synthesis of β -D-2'-CH₃-riboC



β -D-2'-CH₃-riboC

The following syntheses provided are non-limiting steps to achieve the compound β -D-2'-CH₃-riboC. One of ordinary skill in the art may modify the synthesis in any known manner to achieve the compound β -D-2'-CH₃-riboC.

1. *Glycosylation of the nucleobase with an appropriately modified sugar*

The key starting material for this process can be an appropriately substituted sugar with a 2'-OH and 2'-H, with the appropriate leaving group (LG), for example an acyl group or a chloro, bromo, fluoro or iodo. The sugar can be purchased or can be prepared by any known means including standard epimerization, substitution, oxidation and reduction techniques. The substituted sugar can then be oxidized with the appropriate oxidizing agent in a compatible solvent at a suitable temperature to yield the 2'-modified sugar. Possible oxidizing agents are Jones reagent (a mixture of chromic acid and sulfuric acid), Collins's reagent (dipyridine Cr(VI) oxide, Corey's reagent (pyridinium chlorochromate), pyridinium dichromate, acid dichromate, potassium permanganate, MnO₂, ruthenium tetroxide, phase transfer catalysts such as chromic acid or permanganate supported on a polymer, Cl₂-pyridine, H₂O₂-ammonium molybdate, NaBrO₂-CAN, NaOCl in HOAc, copper chromite, copper oxide, Raney nickel, palladium acetate, Meerwin-Pondorf-Verley reagent (aluminum *t*-butoxide with another ketone) and *N*-bromosuccinimide.

Then coupling of an organometallic carbon nucleophile, such as a Grignard reagent, an organolithium, lithium dialkylcopper or CH₃-SiMe₃ in TBAF with the ketone with the appropriate non-protic solvent at a suitable temperature, yields the 2'-methyl

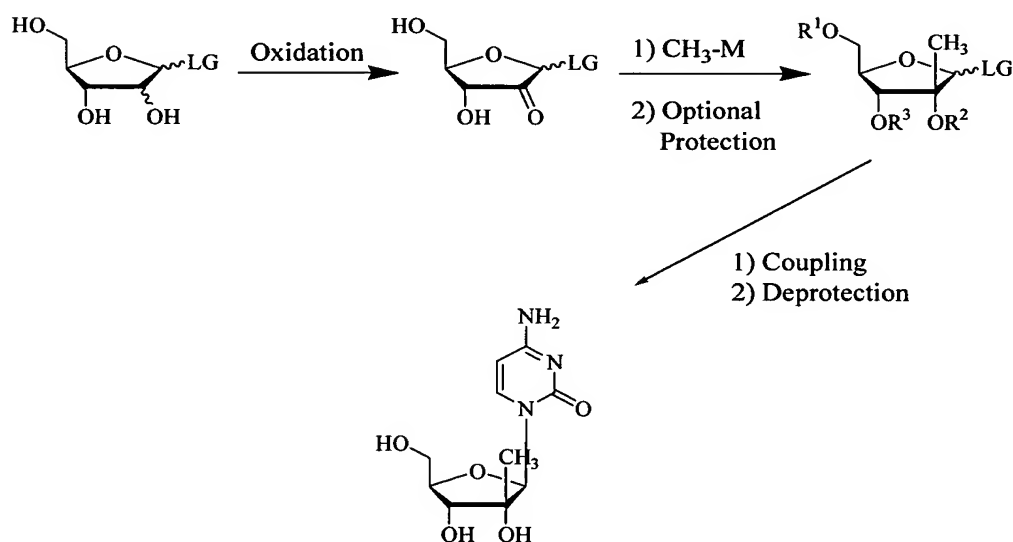
sugar. The methyl sugar can be optionally protected with a suitable protecting group, preferably with an acyl or silyl group, by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

The optionally protected sugar can then be coupled to the BASE by methods well known to those skilled in the art, as taught by Townsend Chemistry of Nucleosides and Nucleotides, Plenum Press, 1994. For example, an acylated sugar can be coupled to a silylated base with a lewis acid, such as tin tetrachloride, titanium tetrachloride or trimethylsilyltriflate in the appropriate solvent at a suitable temperature. Alternatively, a halo-sugar can be coupled to a silylated base with the presence of trimethylsilyltriflate.

Subsequently, the 2'-methyl-nucleoside can be deprotected by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

The synthesis of a 2'-methyl-nucleoside is shown in **Scheme 6**.

Scheme 6



wherein:

LG is a leaving group; and

R¹, R², and R³ are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; lipid (including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R¹, R² or R³ is independently H or phosphate when administered *in vivo*.

2. Modification of a pre-formed 2'-methyl nucleoside

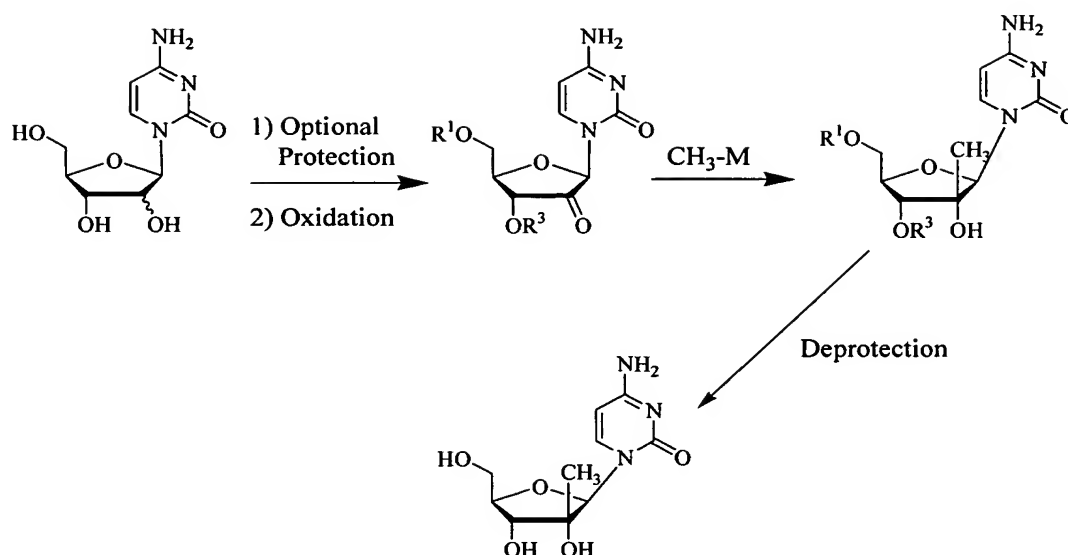
The key starting material for this process can be an appropriately substituted 2'-methyl-cytidine nucleoside with a 2'-OH and 2'-H. The nucleoside can be purchased or can be prepared by any known means including standard coupling techniques. The nucleoside can be optionally protected with suitable protecting groups, preferably with acyl or silyl groups, by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

The appropriately protected 2'-methyl-cytidine nucleoside can then be oxidized with the appropriate oxidizing agent in a compatible solvent at a suitable temperature to yield the 2'-methyl sugar. Possible oxidizing agents are Jones reagent (a mixture of chromic acid and sulfuric acid), Collins's reagent (dipyridine Cr(VI) oxide, Corey's reagent (pyridinium chlorochromate), pyridinium dichromate, acid dichromate, potassium permanganate, MnO₂, ruthenium tetroxide, phase transfer catalysts such as chromic acid or permanganate supported on a polymer, Cl₂-pyridine, H₂O₂-ammonium molybdate, NaBrO₂-CAN, NaOCl in HOAc, copper chromite, copper oxide, Raney nickel, palladium acetate, Meerwin-Pondorf-Verley reagent (aluminum *t*-butoxide with another ketone) and *N*-bromosuccinimide.

Subsequently, the nucleoside can be deprotected by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

The synthesis of a 2'-methyl-cytidine nucleoside is shown in **Scheme 7**.

Scheme 7



wherein:

R^1 and R^3 are independently H, phosphate (including mono-, di- or triphosphate and a stabilized phosphate prodrug); acyl (including lower acyl); alkyl (including lower alkyl); sulfonate ester (including alkyl or arylalkyl sulfonyl including methanesulfonyl); benzyl, wherein the phenyl group is optionally substituted with one or more substituents as described in the definition of aryl given herein; lipid (including a phospholipid); amino acid; carbohydrate; peptide; cholesterol; or a pharmaceutically acceptable leaving group that provides a compound wherein R^1 or R^3 is independently H or phosphate when administered *in vivo*.

Pharmaceutically Acceptable Prodrugs

The term “pharmaceutically acceptable prodrug and/or salt” is used throughout the specification to describe any pharmaceutically acceptable form (such as an ester, phosphate ester, salt of an ester or a related group) of a nucleoside compound which, upon administration to a patient, provides the parent nucleoside compound. Pharmaceutically acceptable salts include those derived from pharmaceutically acceptable inorganic or organic bases and acids. Suitable salts include those derived

from alkali metals such as potassium and sodium, alkaline earth metals such as calcium and magnesium, among numerous other acids well known in the pharmaceutical art. Pharmaceutically acceptable prodrugs refer to a compound that is metabolized, for example hydrolyzed or oxidized, in the host to form the compound of the present invention. Typical examples of prodrugs include compounds that have biologically labile protecting groups on a functional moiety of the active compound. Prodrugs include compounds that can be oxidized, reduced, aminated, deaminated, hydroxylated, dehydroxylated, hydrolyzed, dehydrolyzed, alkylated, dealkylated, acylated, deacylated, phosphorylated, dephosphorylated to produce the active compound. The compounds of this invention possess antiviral activity against a *Flaviviridae*, or are metabolized to a compound that exhibits such activity.

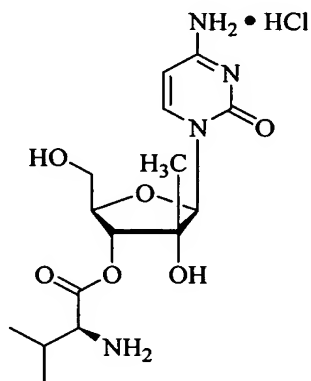
2'-Branched nucleosides, including 2'-branched pyrimidine nucleoside, such as β -D-2'-CH₃-riboC, or related compounds administered as acylated or nucleoside prodrugs can be used in combination or alternation therapy.

Any of the nucleosides described herein or other compounds that contain a hydroxyl or amine function can be administered as a nucleoside prodrug to increase the activity, bioavailability, stability or otherwise alter the properties of the nucleoside. A number of nucleoside prodrug ligands are known. In general, alkylation, acylation or other lipophilic modification of the hydroxyl group of the compound or of the mono, di or triphosphate of the nucleoside will increase the stability of the nucleoside. Examples of substituent groups that can replace one or more hydrogens on the phosphate moiety or hydroxyl are alkyl, aryl, steroids, carbohydrates, including sugars, 1,2-diacylglycerol and alcohols. Many are described in R. Jones and N. Bischofberger, *Antiviral Research*, 27 (1995) 1-17. Any of these can be used in combination with the disclosed nucleosides or other compounds to achieve a desired effect.

The active nucleoside or other hydroxyl containing compound can also be provided as an ether lipid (and particularly a 5'-ether lipid for a nucleoside), as disclosed in the following references: Kucera, L.S., N. Iyer, E. Leake, A. Raben, Modest E.K., D.L.W., and C. Piantadosi. 1990. "Novel membrane-interactive ether lipid analogs that inhibit infectious HIV-1 production and induce defective virus formation." *AIDS Res. Hum. Retro Viruses*. 6:491-501; Piantadosi, C., J. Marasco C.J., S.L. Morris-Natschke,

K.L. Meyer, F. Gumus, J.R. Surles, K.S. Ishaq, L.S. Kucera, N. Iyer, C.A. Wallen, S. Piantadosi, and E.J. Modest. 1991. "Synthesis and evaluation of novel ether lipid nucleoside conjugates for anti-HIV activity." *J. Med. Chem.* **34**:1408.1414; Hosteller, K.Y., D.D. Richman, D.A. Carson, L.M. Stuhmiller, G.M. T. van Wijk, and H. van den Bosch. 1992. "Greatly enhanced inhibition of human immunodeficiency virus type 1 replication in CEM and HT4-6C cells by 3'-deoxythymidine diphosphate dimyristoylglycerol, a lipid prodrug of 3,-deoxythymidine." *Antimicrob. Agents Chemother.* **36**:2025.2029; Hostetler, K.Y., L.M. Stuhmiller, H.B. Lenting, H. van den Bosch, and D.D. Richman, 1990. "Synthesis and antiretroviral activity of phospholipid analogs of azidothymidine and other antiviral nucleosides." *J. Biol. Chem.* **265**:61127. Nonlimiting examples of U.S. patents that disclose suitable lipophilic substituents that can be covalently incorporated into the nucleoside or other hydroxyl or amine containing compound, preferably at the 5'-OH position of the nucleoside or lipophilic preparations, include U.S. Patent Nos. 5,149,794 (Sep. 22, 1992, Yatvin et al.); 5,194,654 (Mar. 16, 1993, Hostetler et al., 5,223,263 (June 29, 1993, Hostetler et al.); 5,256,641 (Oct. 26, 1993, Yatvin et al.); 5,411,947 (May 2, 1995, Hostetler et al.); 5,463,092 (Oct. 31, 1995, Hostetler et al.); 5,543,389 (Aug. 6, 1996, Yatvin et al.); 5,543,390 (Aug. 6, 1996, Yatvin et al.); 5,543,391 (Aug. 6, 1996, Yatvin et al.); and 5,554,728 (Sep. 10, 1996; Basava et al.), all of which are incorporated herein by reference. Foreign patent applications that disclose lipophilic substituents that can be attached to the nucleosides of the present invention, or lipophilic preparations, include WO 89/02733, WO 90/00555, WO 91/16920, WO 91/18914, WO 93/00910, WO 94/26273, WO 96/15132, EP 0 350 287, EP 93917054.4, and WO 91/19721.

2', 3' and 5'-Prodrugs of 2'-branched β -D nucleosides, or their pharmaceutically acceptable salts or pharmaceutically acceptable formulations containing these compounds can be used to treat *Flaviviridae* infections. Specifically, the 3'-valine ester prodrug of β -D-2'-CH₃-riboC represented by the formula:



or its pharmaceutically acceptable salt, can be administered to a subject for the treatment of a *Flaviviridae* infection.

In one embodiment, 2'-branched β -D nucleoside 2'-prodrug includes biologically cleavable moieties at the 2' and/or 5' positions. Preferred moieties are natural or synthetic D or L amino acid esters, including D or L-valyl, though preferably L-amino acid esters, such as L-valyl, and alkyl esters including acetyl. Therefore, this invention specifically includes 2'-D or L-amino acid ester and 2',5'-D or L-diamino acid ester, preferably L-amino acid ester, of 2'-branched β -D or β -L nucleosides with any desired purine or pyrimidine base, wherein the parent drug optionally has an EC₅₀ of less than 15 micromolar, and even more preferably less than 10 micromolar; 2'-(alkyl or aryl) ester or 2',5'-di(alkyl or aryl) ester of 2'-branched β -D or β -L nucleosides with any desired purine or pyrimidine base, wherein the parent drug optionally has an EC₅₀ of less than 10 or 15 micromolar; and prodrugs of 2',5'-diesters of 2'-branched β -D or β -L nucleosides wherein (i) the 2' ester is a natural or synthetic D or L-amino acid ester, though preferably an L-amino acid ester, and the 5'-ester is an alkyl or aryl ester; (ii) both esters are independently natural or synthetic D or L-amino acid ester, though preferably both are L-amino acid esters; (iii) both esters are independently alkyl or aryl esters; and (iv) the 2' ester is independently an alkyl or aryl ester and the 5'-ester is a natural or synthetic D or L-amino acid ester, though preferably an L-amino acid ester, wherein the parent drug optionally has an EC₅₀ of less than 10 or 15 micromolar.

Examples of prodrugs falling within the invention are 2'-D or L-valine ester of β -D-2'-methyl-cytidine; β -D-2',6-dimethyl-cytidine; 2'-L-valine ester of β -D-2',6-dimethyl-thymidine; 2'-L-valine ester of β -D-2',8-dimethyl-adenosine; 2'-L-valine ester of β -D-2',8-dimethyl-guanosine; 2'-L-valine ester of β -D-2',6-dimethyl-5-fluorocytidine; 2'-L-valine ester of β -D-2',6-dimethyl-uridine; 2'-acetyl ester of β -D-2',6-dimethyl-cytidine; 2'-acetyl ester of β -D-2',6-dimethyl-thymidine; 2'-acetyl ester of β -D-2',8-dimethyl-adenosine; 2'-acetyl ester of β -D-2',8-dimethyl-guanosine; 2'-acetyl ester of β -D-2',6-dimethyl-5-fluoro-cytidine; and 2'-esters of β -D-2',6-dimethyl-(cytidine, 5-fluorocytidine, uridine or thymidine) or 2'-esters of β -D-2'-methyl-cytidine or β -D-2',8-dimethyl-(guanosine, adenosine or inosine) wherein (i) the 2' ester is an amino acid ester; or (ii) the 2' ester is an alkyl or aryl ester.

Additional examples of prodrugs falling within the invention are 2',5'-L-divalinal ester of β -D-2'-methyl cytidine; β -D-2',6-dimethyl-cytidine (dival-2',6-diMe-LdC); 2',5'-L-divalinal ester of β -D-2',6-dimethyl-thymidine; 2',5'-L-divalinal ester of β -D-2',8-dimethyl-adenosine; 2',5'-L-divalinal ester of β -D-2',8-dimethyl-guanosine; 2',5'-L-divalinal ester of β -D-2',6-dimethyl-5-fluoro-cytidine; 2',5'-L-divalinal ester of β -D-2',6-dimethyl-uridine; 2',5'-diacetyl ester of β -D-2',6-dimethyl-cytidine; 2',5'-diacetyl ester of β -D-2',6-dimethyl-thymidine; 2',5'-diacetyl ester of β -D-2',8-dimethyl-adenosine; 2',5'-diacetyl ester of β -D-2',8-dimethyl-guanosine; 2',5'-diacetyl ester of β -D-2',6-dimethyl-5-fluoro-cytidine; and 2',5'-diesters of β -D-2',6-dimethyl-(cytidine, 5-fluorocytidine, uridine or thymidine) or 2',5'-diesters of β -D-2'-methyl cytidine or β -D-2',8-dimethyl-(guanosine, adenosine or inosine) wherein (i) the 2' ester is an amino acid ester and the 5'-ester is an alkyl or aryl ester; (ii) both esters are amino acid esters; (iii) both esters are independently alkyl or aryl esters; or (iv) the 2' ester is an alkyl or aryl ester and the 5'-ester is an amino acid ester.

In another embodiment, the 2'-branched β -D nucleoside 3'-prodrug includes biologically cleavable moieties at the 3' and/or 5' positions. Preferred moieties are natural or synthetic D or L amino acid esters, such as valyl, though preferably L-amino acids, such as L-valyl, and alkyl esters including acetyl. Therefore, this invention specifically includes 3'-L-amino acid ester and 3',5'-L-diamino acid ester of a 2'-branched β -D or β -L nucleosides with any desired purine or pyrimidine base, wherein the

parent drug optionally has an EC₅₀ of less than 15 micromolar, and even more preferably less than 10 micromolar; 3'-(alkyl or aryl) ester or 3',5'-L-di(alkyl or aryl) ester of 2'-branched β-D or β-L nucleosides with any desired purine or pyrimidine base, wherein the parent drug optionally has an EC₅₀ of less than 10 or 15 micromolar; and prodrugs of 3',5'-diesters of 2'-branched β-D or β-L nucleosides wherein (i) the 3' ester is a natural or synthetic D or L amino acid ester and the 5'-ester is an alkyl or aryl ester; (ii) both esters are natural or synthetic D or L-amino acid esters; (iii) both esters are independently alkyl or aryl esters; and (iv) the 3' ester is independently an alkyl or aryl ester and the 5'-ester is a natural or synthetic D or L-amino acid ester, wherein the parent drug optionally has an EC₅₀ of less than 10 or 15 micromolar.

Examples of prodrugs falling within the invention are 3'-L-valine ester of β-D-2'-methyl-cytidine; β-D-2',6-dimethyl-cytidine; 3'-L-valine ester of β-D-2',6-dimethyl-thymidine; 3'-L-valine ester of β-D-2',8-dimethyl-adenosine; 3'-L-valine ester of β-D-2',8-dimethyl-guanosine; 3'-L-valine ester of β-D-2',6-dimethyl-5-fluorocytidine; 3'-L-valine ester of β-D-2',6-dimethyl-uridine; 3'-acetyl ester of β-D-2',6-dimethyl-cytidine; 3'-acetyl ester of β-D-2',6-dimethyl-thymidine; 3'-acetyl ester of β-D-2',8-dimethyl-adenosine; 3'-acetyl ester of β-D-2',8-dimethyl-guanosine; 3'-acetyl ester of β-D-2'-methyl-cytidine; 3'-acetyl ester of β-D-2',6-dimethyl-5-fluoro-cytidine; and 3'-esters of β-D-2',6-dimethyl-(cytidine, 5-fluorocytidine, uridine or thymidine) or 3'-esters of β-D-2',8-dimethyl-(guanosine, adenosine or inosine) wherein (i) the 3' ester is an amino acid ester; or (ii) the 3' ester is an alkyl or aryl ester.

Additional examples of prodrugs falling within the invention are 3',5'-L-divalinal ester of β-D-2'-methyl-cytidine; β-D-2',6-dimethyl-cytidine (dival-2',6-diMe-L-dC); 3',5'-L-divalinal ester of β-D-2',6-dimethyl-thymidine; 3',5'-L-divalinal ester of β-D-2',8-dimethyl-adenosine; 3',5'-L-divalinal ester of β-D-2',8-dimethyl-guanosine; 3',5'-L-divalinal ester of β-D-2',6-dimethyl-5-fluoro-cytidine; 3',5'-L-divalinal ester of β-D-2',6-dimethyl-uridine; 3',5'-diacetyl ester of β-D-2',6-dimethyl-cytidine; 3',5'-diacetyl ester of β-D-2',6-dimethyl-thymidine; 3',5'-diacetyl ester of β-D-2',8-dimethyl-adenosine; 3',5'-diacetyl ester of β-D-2',8-dimethyl-guanosine; 3',5'-diacetyl ester of β-D-2',6-dimethyl-5-fluoro-cytidine; and 3',5'-diesters of β-D-2',6-dimethyl-(cytidine, 5-fluorocytidine, or β-D-2'-methyl-cytidine, uridine or thymidine) or 3',5'-diesters of β-D-2',8-

dimethyl-(guanosine, adenosine or inosine) wherein (i) the 3' ester is an amino acid ester and the 5'-ester is an alkyl or aryl ester; (ii) both esters are amino acid esters; (iii) both esters are independently alkyl or aryl esters; or (iv) the 3' ester is an alkyl or aryl ester and the 5'-ester is an amino acid ester.

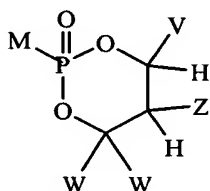
5 In another embodiment, the prodrug of 2'-branched β -D nucleoside includes biologically cleavable moieties at the 2', 3' and/or 5' positions. Preferred moieties are natural or synthetic D or L amino acid esters, including D or L-valyl, though preferably L-amino acid esters, such as L-valyl, and alkyl esters including acetyl. Therefore, this invention specifically includes 2',3'-L or D-diamino acid ester and 2',3',5'-L or D-
10 triamino acid ester of 2'-branched β -D or β -L nucleosides, preferably L-amino acid, with any desired purine or pyrimidine base, wherein the parent drug optionally has an EC_{50} of less than 15 micromolar, and even more preferably less than 10 micromolar; 2',3'-di(alkyl or aryl) ester or 2',3',5'-L-tri(alkyl or aryl) ester of 2'-branched β -D or β -L nucleosides with any desired purine or pyrimidine base, wherein the parent drug
15 optionally has an EC_{50} of less than 10 or 15 micromolar; and prodrugs of 2',3'-diesters of 2'-branched β -D or β -L nucleosides wherein (i) the 2' ester is an amino acid ester and the 3'-ester is an alkyl or aryl ester; (ii) both esters are amino acid esters; (iii) both esters are independently alkyl or aryl esters; and (iv) the 2' ester is independently an alkyl or aryl ester and the 3'-ester is an amino acid ester, wherein the parent drug optionally has
20 an EC_{50} of less than 10 or 15 micromolar. Further, 2',3',5'-triesters of 2'-branched β -D or β -L nucleosides wherein (i) all three esters are amino acid esters; (ii) all three esters are independently alkyl or aryl esters; (iii) the 2' ester is an amino acid ester, the 3' ester is an amino acid ester and the 5'-ester is an alkyl or aryl ester; (iv) the 2' ester is an amino acid ester, the 3' ester is an alkyl or aryl ester and the 5'-ester is an alkyl or aryl
25 ester; (v) the 2' ester is an alkyl or aryl ester, the 3' ester is an alkyl or aryl ester and the 5'-ester is an amino acid ester; (vi) the 2' ester is an alkyl or aryl ester, the 3' ester is an amino acid ester and the 5'-ester is an amino acid ester; (vii) the 2' ester is an alkyl or aryl ester, the 3' ester is an amino acid ester and the 5'-ester is an alkyl or aryl ester; and (viii) the 2' ester is an amino acid ester, the 3' ester is an alkyl or aryl ester and the 5'-
30 ester is an amino acid ester; wherein the parent drug optionally has an EC_{50} of less than 10 or 15 micromolar.

Examples of prodrugs falling within the invention include 2',3'-L-divaline ester of β -D-2'-methyl-cytidine; β -D-2',6-dimethyl-cytidine (dival-2',6-diMe-L-dC); 2',3'-L-divaline ester of β -D-2',6-dimethyl-thymidine; 2',3'-L-divaline ester of β -D-2',8-dimethyl-adenosine; 2',3'-L-divaline ester of β -D-2',8-dimethyl-guanosine; 2',3'-L-divaline ester of β -D-2',6-dimethyl-5-fluoro-cytidine; 2',3'-L-divaline ester of β -D-2',6-dimethyl-uridine; 2',3'-diacetyl ester of β -D-2',6-dimethyl-cytidine; 2',3'-diacetyl ester of β -D-2',6-dimethyl-thymidine; 2',3'-diacetyl ester of β -D-2',8-dimethyl-adenosine; 2',3'-diacetyl ester of β -D-2',8-dimethyl-guanosine; 2',3'-diacetyl ester of β -D-2',6-dimethyl-5-fluoro-cytidine; and 2',3'-diesters of β -D-2',6-dimethyl-(cytidine, 5-fluorocytidine, uridine or thymidine) or 2',3'-diesters of β -D-2',8-dimethyl-(guanosine, adenosine or inosine) wherein (i) the 2' ester is an amino acid ester and the 3'-ester is an alkyl or aryl ester; (ii) both esters are amino acid esters; (iii) both esters are independently alkyl or aryl esters; or (iv) the 2' ester is an alkyl or aryl ester and the 3'-ester is an amino acid ester.

Additional examples of prodrugs falling within the invention include 2',3',5'-L-trivaline ester of β -D-2'-methyl-cytidine; β -D-2',6-dimethyl-cytidine (trival-2',6-diMe-L-dC); 2',3',5'-L-trivaline ester of β -D-2',6-dimethyl-thymidine; 2',3',5'-L-trivaline ester of β -D-2',8-dimethyl-adenosine; 2',3',5'-L-trivaline ester of β -D-2',8-dimethyl-guanosine; 2',3',5'-L-trivaline ester of β -D-2',6-dimethyl-5-fluoro-cytidine; 2',3',5'-L-trivaline ester of β -D-2',6-dimethyl-uridine; 2',3',5'-triacetyl ester of β -D-2',6-dimethyl-cytidine; 2',3',5'-triacetyl ester of β -D-2',6-dimethyl-thymidine; 2',3',5'-triacetyl ester of β -D-2',8-dimethyl-adenosine; 2',3',5'-triacetyl ester of β -D-2',8-dimethyl-guanosine; 2',3',5'-triacetyl ester of β -D-2',6-dimethyl-5-fluoro-cytidine; and 2',3',5'-triesters of β -D-2',6-dimethyl-(cytidine, 5-fluorocytidine, uridine or thymidine) and 2',3',5'-triesters of β -D-2',8-dimethyl-(guanosine, adenosine or inosine) wherein (i) all three esters are amino acid esters; (ii) all three esters are independently alkyl or aryl esters; (iii) the 2' ester is an amino acid ester, the 3' ester is an amino acid ester and the 5'-ester is an alkyl or aryl ester; (iv) the 2' ester is an amino acid ester, the 3' ester is an alkyl or aryl ester and the 5'-ester is an alkyl or aryl ester; (v) the 2' ester is an alkyl or aryl ester, the 3' ester is an alkyl or aryl ester and the 5'-ester is an amino acid ester; (vi) the 2' ester is an alkyl or aryl ester, the 3' ester is an amino acid ester and the 5'-ester is an amino acid ester; (vii) the 2' ester is an alkyl or aryl ester, the 3' ester is an amino acid ester and the

5'-ester is an alkyl or aryl ester; and (viii) the 2' ester is an amino acid ester, the 3' ester is an alkyl or aryl ester and the 5'-ester is an amino acid ester.

Further examples of prodrugs falling within the invention include prodrugs disclosed in U.S. Patent Nos. 6,284,748 and 6,312,662. In particular, the prodrugs of the present invention include compounds of the structure



wherein:

V, W and W' are independently selected from the group consisting of --H, alkyl, aralkyl, alicyclic, aryl, substituted aryl, heteroaryl, substituted heteroaryl, 1-alkenyl, and 1-alkynyl; or

together V and Z are connected via an additional 3-5 atoms to form a cyclic group containing 5-7 atoms, optionally 1 heteroatom, substituted with hydroxy, acyloxy, alkoxycarbonyloxy, or aryloxycarbonyloxy attached to a carbon atom that is three atoms from both O groups attached to the phosphorus; or

together V and Z are connected via an additional 3-5 atoms to form a cyclic group, optionally containing 1 heteroatom, that is fused to an aryl group at the beta and gamma position to the O attached to the phosphorus; or

together V and W are connected via an additional 3 carbon atoms to form an optionally substituted cyclic group containing 6 carbon atoms and substituted with one substituent selected from the group consisting of hydroxy, acyloxy, alkoxycarbonyloxy, alkylthiocarbonyloxy, and aryloxycarbonyloxy, attached to one of said carbon atoms that is three atoms from an O attached to the phosphorus;

together Z and W are connected via an additional 3-5 atoms to form a cyclic group, optionally containing one heteroatom, and V must be aryl, substituted aryl, heteroaryl, or substituted heteroaryl;

together W and W' are connected via an additional 2-5 atoms to form a cyclic group, optionally containing 0-2 heteroatoms, and V must be aryl, substituted aryl, heteroaryl, or heteroaryl;

Z is selected from the group consisting of $-\text{CHR}^2\text{OH}$, $-\text{CHR}^2\text{OC}(\text{O})\text{R}^3$, $-\text{CHR}^2\text{OC}(\text{S})\text{R}^3$, $-\text{CHR}^2\text{OC}(\text{S})\text{OR}^3$, $-\text{CHR}^2\text{OC}(\text{O})\text{SR}^3$, $-\text{CHR}^2\text{OCO}_2\text{R}^3$, $-\text{OR}^2$, $-\text{SR}^2$, $-\text{CHR}^2\text{N}_3$, $-\text{CH}_2\text{aryl}$, $-\text{CH}(\text{aryl})\text{OH}$, $-\text{CH}(\text{CH}=\text{CR}^2)\text{OH}$, $-\text{CH}(\text{C}\equiv\text{CR}^2)\text{OH}$, $-\text{R}^2$, $-\text{NR}^2_2$, $-\text{OCOR}^3$, $-\text{OCO}_2\text{R}^3$, $-\text{SCOR}^3$, $-\text{SCO}_2\text{R}^3$, $-\text{NHCOR}^2$, $-\text{NHCO}_2\text{R}^3$, $-\text{CH}_2\text{NHaryl}$, $-(\text{CH}_2)_p\text{-OR}^{12}$, and $-(\text{CH}_2)_p\text{-SR}^{12}$;

p is an integer 2 or 3;

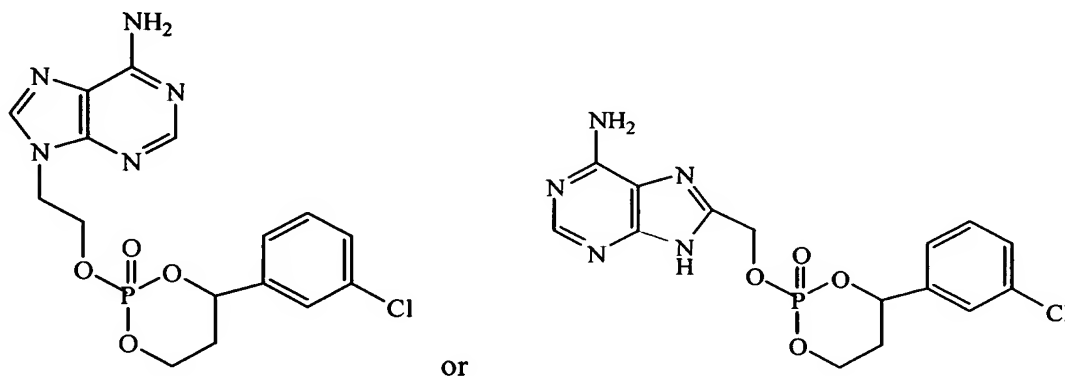
R^2 is selected from the group consisting of $\text{R}^{\text{sup.3}}$ and $-\text{H}$;

R^3 is selected from the group consisting of alkyl, aryl, alicyclic, and aralkyl;

R^{12} is selected from the group consisting of $-\text{H}$, and lower acyl;

M is selected from the group that attached to PO_3^{2-} , $\text{P}_2\text{O}_6^{3-}$ or $\text{P}_3\text{O}_9^{4-}$ is a the 2'-branched nucleoside, and is attached to the phosphorus via a carbon, oxygen, sulfur or nitrogen atom.

In one non-limiting example, the prodrug is attached to the nucleoside as in the following compounds



General Synthesis of 2' and/or 3'-Prodrugs

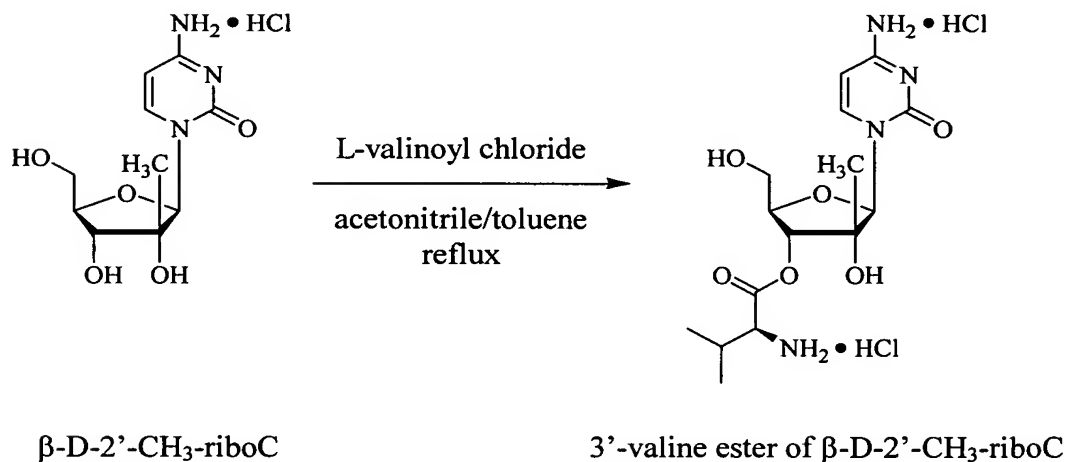
The key starting material for this process is an appropriately substituted 2'-branched β -D nucleosides. The branched nucleoside can be purchased or can be

prepared by any known means including the techniques disclosed herein. The branched nucleoside can be optionally protected with a suitable protecting group, preferably with a silyl group, by methods well known to those skilled in the art, as taught by Greene *et al.* Protective Groups in Organic Synthesis, John Wiley and Sons, Second Edition, 1991.

5 The protected branched nucleoside can then be coupled with a suitable acyl donor, such as an acyl chloride and/or an acyl anhydride with the appropriate protic or aprotic solvent at a suitable temperature, to give the 2' and/or 3' prodrug of 2'-branched β -D nucleoside. Alternatively, the protected branched nucleoside can then be coupled with a suitable acyl, such as a carboxylic acid, such as alkanolic acid and/or amino acid residue, optionally with a suitable coupling agent, with the appropriate aprotic solvent at a
10 suitable temperature, to give the 2' and/or 3' prodrug of 2'-branched β -D nucleoside. Possible coupling reagents are any reagents that promote coupling, including but are not limiting to, Mitsunobu reagents (e.g. diisopropyl azodicarboxylate and diethyl azodicarboxylate) with triphenylphosphine or various carbodiimides.

15 For example, simple amino-alcohols can be esterified using acid chlorides in refluxing acetonitrile-benzene mixture (See **Scheme 8** below: *Synthetic Communications*, 1978, 8(5), 327-333; hereby incorporated by reference). Alternatively, esterification can be achieved using an anhydride, as described in *J. Am. Chem. Soc.*, 1999, 121(24), 5661-5664.

Scheme 8



III. Detection of the β -D-2'-CH₃-riboC Induced Mutation in a *Flaviviridae* Genome

In one embodiment, a method is provided for treating a patient infected with *Flaviviridae* comprising:

- 5 (i) administering an effective amount of β -D-2'-CH₃-riboC or a prodrug, such as the 3' valine ester prodrug of β -D-2'-CH₃-riboC, or a pharmaceutically acceptable salt thereof;
- (ii) identifying viral resistance to β -D-2'-CH₃-riboC in the patient;
- 10 (iii) administering an effective amount of one or more drugs that in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that are associated with such a mutation.
- 15

In another embodiment, a method is provided for treating a patient infected with HCV comprising:

- 20 (i) administering an effective amount of β -D-2'-CH₃-riboC or a prodrug, such as the 3' valine ester prodrug of β -D-2'-CH₃-riboC, or a pharmaceutically acceptable salt thereof;
- (ii) identifying viral resistance to β -D-2'-CH₃-riboC in the patient;
- 25 (iii) administering an effective amount of one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine at position 282 to a different amino acid, such as threonine, in the highly conserved consensus sequence, XXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that is associated with such a mutation.

In one embodiment, a method is provided for treating a host infected with BVDV comprising:

- (i) administering an effective amount of β -D-2'-CH₃-riboC or a prodrug, such as the 3' valine ester prodrug of β -D-2'-CH₃-riboC, or a pharmaceutically acceptable salt thereof;
- (ii) identifying viral resistance to β -D-2'-CH₃-riboC in the host;
- (iii) administering an effective amount of one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine at position 405 to a different amino acid, such as threonine, in the highly conserved consensus sequence, XXRXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that is associated with such a mutation.

In another embodiment, the invention provides a method for treating a patient infected with *Flaviviridae* comprising:

- (i) administering an effective amount of β -D-2'-CH₃-riboC or a prodrug, such as the 3' valine ester prodrug of β -D-2'-CH₃-riboC, or a pharmaceutically acceptable salt thereof;
- (ii) identifying viral resistance to β -D-2'-CH₃-riboC in the patient;
- (iii) administering an effective amount of interferon.

In certain embodiments, identification of viral resistance to β -D-2'-CH₃-riboC in the patient can be determined by a phenotypic analysis of viral plaque growth. In another embodiment, identification of viral resistance to β -D-2'-CH₃-riboC in the patient can be determined by the replication fitness of the virus. In a further embodiment, identification of viral resistance to β -D-2'-CH₃-riboC in the patient can be determined by detecting the presence of cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or cytidine at nucleotide 8443 of HCV.

In one embodiment, the present invention includes a method for treating a patient infected with *Flaviviridae* comprising:

- 5 (i) administering an effective amount of β -D-2'-CH₃-riboC or a prodrug, such as the 3' valine ester prodrug of β -D-2'-CH₃-riboC, or a pharmaceutically acceptable salt thereof;
- (ii) obtaining a viral culture sample from the patient;
- (iii) culturing the sample and comparing the plaque growth between the sample and wild type virus;
- 10 (iv) determining whether the plaque growth of the sample is smaller than the plaque growth of the wildtype, which indicates resistance to β -D-2'-CH₃-riboC;
- (v) administering an effective amount of interferon to those patients that are resistant to β -D-2'-CH₃-riboC.

15 In another embodiment, the invention provides a method for treating a patient infected with *Flaviviridae* comprising:

- (i) administering an effective amount of β -D-2'-CH₃-riboC or a prodrug, such as the 3' valine ester prodrug of β -D-2'-CH₃-riboC, or a pharmaceutically acceptable salt thereof;
- 20 (ii) obtaining a viral sample from the patient;
- (iii) determining the replication fitness of the viral;
- (iv) determining whether the replication fitness of the sample is less than the replication fitness of the wild-type virus, which indicates resistance to β -D-2'-CH₃-riboC;
- 25 (v) administering an effective amount of interferon to those patients that are resistant to β -D-2'-CH₃-riboC.

In one embodiment, viral plaque growth and/or viral replication fitness can be quantitated by a viral plaque assay. In other embodiments, other assays, such as the infectious center assay, virus-inducible reporter assay, transformation assay, end point dilution assay, or RT-PCR technology can be used to quantitate viral titers (Flint et al. Principles of Virology (ASM) Chapter 2; Wagner & Hewlett. Basic Virology (Blackwell), Chapters 9 & 10).

A plaque assay can be conducted to quantitate viral plaque growth and/or replication fitness. A dilute solution of the virus can be applied to a culture dish that contains a monolayer of host cells. The cells can be overlaid with a semisolid layer (such as a viscous medium, for example, agar) to prevent virus diffusion from one infected cell to another. After incubation the 'plaques' can be recognized, and the number of infective virus particles in the original suspension estimated. One method to recognize the plaques is through the use of antibody staining methods to detect viral antigens within infected cells in the monolayer. These infected cells can then be visualized using a chromagen or a fluorescent label on the virus-specific antibody. The plaques can be observed for phenotypic analysis and/or counted to determine viral titers. Virus titers can be calculated in focus forming units (FFU)/mL, using the following equation: $T_{\text{FFU/mL}} = N \times 5 \times D$; where T is a virus titer in FFU/mL; N is a number of plaques per well; and D is a dilution factor for the corresponding virus sample. (For example, if 12 plaques were found in a well corresponding to 10^{-5} dilution of virus sample, then $T = 12 \times 5 \times 10^5 = 6 \times 10^6$ PFU/mL) and viral replication fitness, which is the overall replicative ability to produce infected progeny in a defined host environment, can then be determined.

Another aspect of the invention is a method for detecting the presence of the nucleotide 1214 G to C mutation of the RNA polymerase region of BVDV (causing a mutation from Serine to Threonine at amino acid 405). Since it is recognized that Ser₄₀₅, the amino acid position of the BVDV putative functional NS5B domain B, is highly conserved among all hepac-, pesti- and flavivirus genomes (Figure 11; Lai et al. J Virol., 1999, 73, 10129-36), the corresponding serine residues of the putative functional NS5B domain B of other *Flaviviridae*s that are mutated can be detected according to the embodiments of the present invention. For example, Ser₄₀₅ of the RNA polymerase domain of BVDV corresponds to Ser₂₈₂ of the RNA polymerase domain of HCV.

Therefore, the embodiments of the present invention also encompass a G to C mutation of nucleotide 8443 of the HCV genome (which corresponds to nucleotide 1214 of the BVDV RNA polymerase region).

In one embodiment, the invention provides a process for detecting a mutation that indicates β -D-2'-CH₃-riboC resistance, which includes contacting a sample containing a *Flaviviridae* nucleic acid sequence with an oligonucleotide probe having a sequence complementary to a section of the *Flaviviridae* genome that includes the mutation; and then determining if the oligonucleotide hybridizes to the viral nucleic acid.

In other embodiments, the invention provides a method for treating a patient infected with *Flaviviridae* comprising:

- (i) administering an effective amount of β -D-2'-CH₃-riboC or a prodrug, such as the 3' valine ester prodrug of β -D-2'-CH₃-riboC, or a pharmaceutically acceptable salt thereof;
- (ii) assaying the blood of the patient to test for seroconversion from wildtype to mutant virus;
- (iii) administering an effective amount of interferon.

In yet another embodiment, the invention provides a method for assaying a sample suspected of containing a β -D-2'-CH₃-riboC -resistant *Flaviviridae* comprising:

- (i) contacting a sample containing a *Flaviviridae* nucleic acid sequence with a detectable oligonucleotide probe having a sequence complementary to the cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or nucleotide 8443 of the HCV genome;
- (ii) allowing the probe to hybridize to the sequence;
- (iii) detecting the hybridization of the probe to cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or nucleotide 8443 of the HCV genome;.

In a further embodiment, the invention provides a method for assaying a sample suspected of containing a Thr instead of a Ser in the highly conserved consensus sequence, XXRSGXXXXT, of domain B of the RNA polymerase region of a *Flaviviridae*, which indicates that the virus is hypersensitive to interferon treatment, comprising:

- 5 (i) contacting a sample suspected of containing a *Flaviviridae* nucleic acid sequence with a detectable oligonucleotide probe having a sequence complementary a codon that encodes Thr in the position of Ser in the conserved consensus sequence, XXRSGXXXXT, of domain B of the RNA polymerase region of a *Flaviviridae*;
- 10 (ii) allowing the probe to hybridize to the sequence;
- (iii) detecting the hybridization of the probe to the sequence.

15 In another embodiment, the invention provides a method for assaying a sample suspected of containing a Thr instead of a Ser at amino acid position 405 or a cytidine at nucleotide 1214 of the RNA polymerase region of BVDV, which indicates that the virus is hypersensitive to interferon treatment, comprising:

- (i) contacting a sample suspected of containing a BVDV nucleic acid sequence with a detectable oligonucleotide probe having a sequence complementary to the cytidine at nucleotide 1214 of the RNA polymerase region;
- 20 (ii) allowing the probe to hybridize to the sequence;
- (iii) detecting the hybridization of the probe to cytidine at nucleotide 1214 of the RNA polymerase region of BVDV.

25 In another embodiment, the invention provides a method for assaying a sample suspected of containing a Thr instead of a Ser in the highly conserved consensus sequence, XXRSGXXXXT, of domain B of the RNA polymerase region at the highly conserved at amino acid position 282 or a cytidine at nucleotide 8433 of the HCV genome, which indicates that the virus is hypersensitive to interferon treatment, comprising:

- (i) contacting a sample suspected of containing a HCV nucleic acid sequence with a detectable oligonucleotide probe having a sequence complementary to the cytidine at nucleotide 8443;
- (ii) allowing the probe to hybridize to the sequence;
- 5 (iii) detecting the hybridization of the probe to cytidine at nucleotide 8443 of the HCV genome.

Oligonucleotide Probes

Oligonucleotide probes are provided that are capable of detecting the presence of a 2'-branched pyrimidine nucleoside-induced mutation of *Flaviviridae*. The probes are complementary to sequences of viral nucleic acids that include the mutation. These probes can be used in processes and kits. The oligonucleotide probes can detect the nucleotide cytidine at nucleotide 1214 of the RNA polymerase region of BVDV or at nucleotide 8443 of the RNA polymerase region of HCV, or other nucleotides of *Flaviviridae* that encode the conserved serine of the domain B of RNA polymerase within a *Flaviviridae* genome (**Figure 11**).

The oligonucleotide probes are preferably at least 14 nucleotides in length, and in a preferred embodiment, are at least 15, 20, 25 or 30 nucleotides in length. It is generally not preferred to use a probe that is greater than approximately 25 or 30 nucleotides in length. In one embodiment, the oligonucleotide probe can be designed to identify the guanine to cytidine base change at nucleotide 1214 of the RNA polymerase region of a BVDV. In another embodiment, the oligonucleotide probe can be designed to identify the guanine to cytidine base change at nucleotide 8443 of HCV. The oligonucleotide probe can be designed such that the mutated region is located in the interior section of the hybridized segment, or alternatively can be on either the 3' or 5' end of the hybridized segment. It is preferred that the mutated region be located near the middle of the probe to allow efficient hybridization.

Table 2 below provides illustrative embodiments of BVDV nucleotide sequences that include nucleotide position 1214 of the RNA polymerase region, alternatively referred to as nucleotide position 11,136 (see Genbank accession number AJ133739; Vassilev and Donis (2000) *Virus Res* 69(2) 95-107). Given these sequences, one of

ordinary skill using standard algorithms can construct oligonucleotide probes that are complementary or substantially complementary to the nucleotide sequences below. The rules for complementary pairing are well known: cytidine (“C”) always pairs with guanine (“G”) and thymine (“T”) or uracil (“U”) always pairs with adenine (“A”). It is
5 recognized that it is not necessary for the probe to be 100% complementary to the target nucleic acid sequence, as long as the probe sufficiently hybridizes and can recognize the diagnostic nucleotide. A certain degree of base pair mismatch can generally be tolerated.

Table 2: Nonlimiting examples of nucleic acid sequences with a single point mutation at nucleotide 1214 (also referred to as position 11,136: see Genebank accession number AJ133739; Vassilev and Donis, Virus Res **2000**, 69(2), 95-107) of the RNA polymerase region of BVDV

		405 S	405 T
AAGTATATATAAGAAATGGGCAGAGAGGG	AGC ACC	GGCCAGCCAGACACAAAGTGTCTGGCAACAG.	(Seq. ID No. 1)
AAGTATATATAAGAAATGGGCAGAGAGGG	ACC		(Seq. ID No. 2)
AGTATATATAAGAAATGGGCAGAGAGGG	ACC	G.	(Seq. ID No. 3)
GTATATATAAGAAATGGGCAGAGAGGG	ACC	GG.	(Seq. ID No. 4)
TATATATAAGAAATGGGCAGAGAGGG	ACC	GGC.	(Seq. ID No. 5)
ATATATAAGAAATGGGCAGAGAGGG	ACC	GGCC.	(Seq. ID No. 6)
TATATAAGAAATGGGCAGAGAGGG	ACC	GGCCA.	(Seq. ID No. 7)
ATATAAGAAATGGGCAGAGAGGG	ACC	GGCCAG.	(Seq. ID No. 8)
TATAAGAAATGGGCAGAGAGGG	ACC	GGCCAGC.	(Seq. ID No. 9)
ATAAGAAATGGGCAGAGAGGG	ACC	GGCCAGCC.	(Seq. ID No. 10)
TAAGAAATGGGCAGAGAGGG	ACC	GGCCAGCCA.	(Seq. ID No. 11)
AAGAAATGGGCAGAGAGGG	ACC	GGCCAGCCAG.	(Seq. ID No. 12)
AGAAATGGGCAGAGAGGG	ACC	GGCCAGCCAGA.	(Seq. ID No. 13)
GAAATGGGCAGAGAGGG	ACC	GGCCAGCCAGAC.	(Seq. ID No. 14)
AAATGGGCAGAGAGGG	ACC	GGCCAGCCAGACA.	(Seq. ID No. 15)
AATGGGCAGAGAGGG	ACC	GGCCAGCCAGACAC.	(Seq. ID No. 16)
ATGGGCAGAGAGGG	ACC	GGCCAGCCAGACACA.	(Seq. ID No. 17)
TGGGCAGAGAGGG	ACC	GGCCAGCCAGACACAA.	(Seq. ID No. 18)
GGGCAGAGAGGG	ACC	GGCCAGCCAGACACAAG.	(Seq. ID No. 19)
GGCAGAGAGGG	ACC	GGCCAGCCAGACACAAGT.	(Seq. ID No. 20)
GCAGAGAGGG	ACC	GGCCAGCCAGACACAAGTG.	(Seq. ID No. 21)
CAGAGAGGG	ACC	GGCCAGCCAGACACAAGTGC.	(Seq. ID No. 22)
AGAGAGGG	ACC	GGCCAGCCAGACACAAGTGT.	(Seq. ID No. 23)
GAGAGGG	ACC	GGCCAGCCAGACACAAGTGTG.	(Seq. ID No. 24)
AGAGGG	ACC	GGCCAGCCAGACACAAGTGTGG.	(Seq. ID No. 25)
GAGGG	ACC	GGCCAGCCAGACACAAGTGTGGC.	(Seq. ID No. 26)
AGGG	ACC	GGCCAGCCAGACACAAGTGTGGCA.	(Seq. ID No. 27)
GGG	ACC	GGCCAGCCAGACACAAGTGTGGCAA.	(Seq. ID No. 28)
GG	ACC	GGCCAGCCAGACACAAGTGTGGCAAC.	(Seq. ID No. 29)
G	ACC	GGCCAGCCAGACACAAGTGTGGCAACA.	(Seq. ID No. 30)
		GGCCAGCCAGACACAAGTGTGTGGCAACAG.	(Seq. ID No. 31)

Therefore, in one embodiment, the oligonucleotide has 1, 2, 3, 4, 5 or 6 mismatches in complementarity to the *Flaviviridae* nucleotide sequence.

Other aspects of the present invention provide a method to treat a *Flaviviridae* infection by administering a therapeutically effective amount of a 2'-branched nucleoside, for example, a 2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC, or its pharmaceutically acceptable prodrug and/or salt, to a human in need of therapy, in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a threonine in the highly conserved consensus sequence, **XXRSGXXXT**, of domain B of the RNA polymerase region, and/or one or more drugs that is associated with such mutation. The codons ACA, ACG or ACU, which also encode Threonine, can be substituted for the codon ACC (in bold) in **Table 2** above to detect the presence of a Threonine in domain B of the RNA polymerase region of BVDV.

Another aspect of the present invention provides a method to treat and/or to substantially cure a *Flaviviridae* infection in a host infected with a *Flaviviridae* that contains a Serine to Threonine mutation in the highly conserved consensus sequence, **XXRSGXXXT**, of domain B of the RNA polymerase region by administering a therapeutically effective amount of interferon. Therefore, in other embodiments of the present invention, the codons ACA, ACG or ACU, which also encode Threonine, can be substituted for the codon ACC (in bold) in **Table 2** above, for example to detect the presence of a Threonine at residue 405 of the RNA polymerase region of BVDV.

Table 3 below provides illustrative embodiments of HCV nucleotide sequences that include nucleotide position 8443 of the HCV genome (Genbank accession number AJ238799; Lohmann et al. (1999) Science 285(5424)110-113). Nucleotide position 8443 of the HCV genome corresponds to nucleotide position 11,136 of the BVDV genome and represents the conserved Serine residue of the RNA polymerase of *Flaviviridae* (Ser₄₀₅ of the BVDV genome, which corresponds to Ser₂₈₂ of the HCV genome (see **Figure 11**)) that is mutated due to treatment with β -D-2'-CH₃-riboC. As stated above, given these sequences, one of ordinary skill using standard algorithms can construct oligonucleotide probes that are complementary or substantially complementary to the nucleotide sequences below.

Table 3: Nonlimiting examples of nucleic acid sequences encompassing a single point mutation at nucleotide 8443 (Genebank accession number AJ238799; Lohmann et al. (1999) Science 285(5424)110-113) of the RNA polymerase region of HCV

	AGC	282 S
AGAACTGCGGCTATGCGCGGTGCGCGCG	ACC	282 T
		(Seq. ID No. 32)
AGAACTGCGGCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 33)
GAACTGCGGCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 34)
AACTGCGGCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 35)
ACTGCGGCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 36)
CTGCGGCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 37)
TGCGGCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 38)
GCGGCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 39)
CGGCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 40)
GGCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 41)
GCTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 42)
CTATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 43)
TATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 44)
ATGCGCGGTGCGCGCG	ACC	(Seq. ID No. 45)
TGCGCGGTGCGCGCG	ACC	(Seq. ID No. 46)
CGCGCGGTGCGCGCG	ACC	(Seq. ID No. 47)
GCGCGGTGCGCGCG	ACC	(Seq. ID No. 48)
CCGCGGTGCGCGCG	ACC	(Seq. ID No. 49)
CGGTGCGCGCG	ACC	(Seq. ID No. 50)
GGTGGCGCGCG	ACC	(Seq. ID No. 51)
GTGGCGCGCG	ACC	(Seq. ID No. 52)
TGGCGCGCG	ACC	(Seq. ID No. 53)
GCGCGCG	ACC	(Seq. ID No. 54)
CCGCGCG	ACC	(Seq. ID No. 55)
CGCGCG	ACC	(Seq. ID No. 56)
GCGCG	ACC	(Seq. ID No. 57)
CGCG	ACC	(Seq. ID No. 58)
GCG	ACC	(Seq. ID No. 59)
CG	ACC	(Seq. ID No. 60)
G	ACC	(Seq. ID No. 61)
	ACC	(Seq. ID No. 62)
	GGTGACTGACGACCGAGCTGCGGTAATAC.	

Therefore, in one embodiment, the oligonucleotide has 1, 2, 3, 4, 5 or 6 mismatches in complementarity to the *Flaviviridae* nucleotide sequence.

Other aspects of the present invention provide a method to treat a *Flaviviridae* infection by administering a therapeutically effective amount of a 2'-branched nucleoside, for example, a 2'-branched pyrimidine nucleoside, for example β -D-2'-CH₃-riboC, or its pharmaceutically acceptable prodrug and/or salt, to a human in need of therapy, in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in a *Flaviviridae* at a location other than a mutation of a nucleotide that results in a change from serine to a threonine in the highly conserved consensus sequence, **XXRXSGXXXT**, of domain B of the RNA polymerase region, and/or one or more drugs that is associated with such mutation. As before, the codons ACA, ACG or ACU, which also encode Threonine, can be substituted for the codon ACC (in bold) in **Table 3**, for example to detect the presence of a Threonine in domain B of the RNA polymerase region of HCV.

Another aspect of the present invention provides a method to treat and/or to substantially cure a *Flaviviridae* infection in a host infected with a *Flaviviridae* that contains a Serine to Threonine mutation in the highly conserved consensus sequence, **XXRXSGXXXT**, of domain B of the RNA polymerase region by administering a therapeutically effective amount of interferon. The codons ACA, ACG or ACU, which also encode Threonine, can be substituted for the codon ACC (in bold) in **Table 3**, as above, for example to detect the presence of a Threonine at residue 282 of the RNA polymerase region of HCV.

In another embodiment, the invention provides an oligonucleotide primer for amplifying a *Flaviviridae* nucleic acid sequence. In one embodiment, the oligonucleotide is at least 14 nucleotides in length and hybridizes under sequence-specific, stringent hybridization conditions to a nucleotide sequence that contains the mutation.

Oligonucleotide sequences used as the hybridizing region of a primer can also be used as the hybridizing region of a probe. Suitability of a primer sequence for use as a probe depends on the hybridization characteristics of the primer. Similarly, an oligonucleotide used as a probe can be used as a primer.

It will be apparent to those of skill in the art that, provided with these embodiments, that specific primers and probes can be prepared by, for example, the addition of nucleotides to either the 5'- or 3'-ends, which nucleotides are complementary to the target sequence or are not complementary to the target sequence. So long as primer compositions serve as a point of initiation for extension on the target sequences, and so long as the primers and probes comprise at least 14 consecutive nucleotides contained within those exemplified embodiments, such compositions are within the scope of the invention.

The probe(s) herein can be selected by the following non-limiting criteria, which are not considered exclusive or determinative: (1) the probes are selected from the region of the *Flaviviridae* genome that contains the mutation.; (2) the probes lack homology with any sequences of viral genomes that would be expected to compromise the test; and (3) the probes lack secondary structure formation in the amplified nucleic acid that, for example, can interfere with nucleic acid extension by an amplification enzyme such as *E. coli* DNA polymerase, such as the portion of the DNA polymerase referred to as the Klenow fragment. Prevention of secondary structure formation can be accomplished by employing up to about 15% by weight, preferably 5-10% by weight, dimethyl sulfoxide (DMSO) in the amplification medium and/or increasing the amplification temperatures to 30°-40 °C.

Further, the probe can have an approximate 50% content of guanine and cytidine, and may not contain multiple consecutive adenine and thymine residues at the 3'-end of the primer which can result in less stable hybrids.

The probes of the invention can be about 10 to 30 nucleotides long, preferably at least 14, 15, 20, 25, or 30 nucleotides in length. The nucleotides as used in the present invention can be ribonucleotides, deoxyribonucleotides and modified nucleotides such as inosine or nucleotides containing modified groups that do not essentially alter their hybridization characteristics. Probe sequences are represented throughout the specification as single stranded DNA oligonucleotides from the 5'- to the 3'-end. Any of the probes can be used as such, or in their complementary form, or in their RNA form (wherein T is replaced by U).

The probes according to the invention can be prepared by cloning of recombinant plasmids containing inserts including the corresponding nucleotide sequences, optionally by cleaving the latter from the cloned plasmids through the use of adequate nucleases and recovering them, e.g. by fractionation according to molecular weight. The probes
5 according to the present invention can also be synthesized chemically, for instance, by the conventional phosphotriester or phosphodiester methods or automated embodiments thereof. In one such automated embodiment diethylphosphoramidites are used as starting materials and can be synthesized as described by Beaucage et al., Tetrahedron Letters (1981), 22:1859-1862. One method for synthesizing oligonucleotides on a modified
10 solid support is described in U.S. Pat. No. 4,458,066. It is also possible to use a primer which has been isolated from a biological source (such as a restriction endonuclease digest).

The oligonucleotides used as primers or probes can also comprise nucleotide analogues such as phosphorothiates (Matsukura et al., 1967), alkylphosphorothiates
15 (Miller et al., 1979), peptide nucleic acids (Nielsen et al., 1991; Nielsen et al., 1993), morpholino nucleic acids, locked nucleic acids, pseudocyclic oligonucleobases, 2'-O-4'-C-ethylene bridged nucleic acids or can contain intercalating agents (Asseline et al., 1984).

For designing probes with desired characteristics, the following useful guidelines
20 known to the person skilled in the art can be applied. Because the extent and specificity of hybridization reactions are affected by a number of factors, manipulation of one or more of those factors will determine the exact sensitivity and specificity of a particular probe, whether perfectly complementary to its target or not. The importance and effect of various assay conditions, explained further herein, are known to those skilled in the art.

The stability of the probe to target nucleic acid hybrid should be chosen to be compatible with the assay conditions. This can be accomplished by avoiding long AT-rich sequences, by terminating the hybrids with GC base pairs, and by designing the probe with an appropriate T_m . The beginning and end of the probe should be chosen so
25 that the length and %GC result in a T_m of about 2-10 °C higher than the temperature at which the final assay will be performed. The base composition of the probe is significant because G-C base pairs exhibit greater thermal stability due to additional
30 hydrogen bonding as compared to A-T base pairs. Thus, hybridization involving

complementary nucleic acids of higher G-C content will be stable at higher temperatures. Conditions such as ionic strength and incubation temperature under which the probe will be used should also be taken into account when designing the probe. It is known that hybridization can increase as the ionic strength of the reaction mixture increases, and that the thermal stability of the hybrids can increase with increasing ionic strength. On the other hand, chemical reagents, such as formamide, urea, DIVISO and alcohols, which disrupt hydrogen bonds, can increase the stringency of hybridization. Destabilization of hydrogen bonds by such reagents can greatly reduce the T_m . In general, optimal hybridization for synthetic oligonucleotide probes of about 10-50 bases in length occurs approximately 5 °C below the melting temperature for a given duplex. Incubation at temperatures below the optimum can allow mismatched base sequences to hybridize and can therefore result in reduced specificity. It is desirable to have probes that hybridize only under conditions of high stringency, in which only highly complementary nucleic acid hybrids will form and/or hybrids without a sufficient degree of complementarity will not form. Accordingly, the stringency of the assay conditions determines the degree of complementarity needed between two nucleic acid strands that form the hybrid. The degree of stringency is chosen, for example, to maximize the difference in stability between the hybrid formed with the target and the non-target nucleic acid. In the present case, single base pair changes need to be detected, which requires conditions of very high stringency.

The length of the target nucleic acid sequence and the length of the probe sequence should also be considered. In some cases, there can be several sequences from a particular region, varying in location and length, which will yield probes with the desired hybridization characteristics. In other cases, one sequence can be significantly better than another which differs merely by a single base.

While it is possible for nucleic acids that are not perfectly complementary to hybridize, the longest stretch of perfectly complementary base sequences normally will determine hybrid stability. While oligonucleotide probes of different lengths and base compositions can be used, preferably oligonucleotide probes of this invention are between about 14 and 30 bases in length and optionally further have a sufficient sequence length that is perfectly complementary to the target nucleic acid sequence.

Regions in the target DNA or RNA that are known to form strong internal structures inhibitory to hybridization are less preferred. In one embodiment, probes with extensive self-complementarity are avoided. As explained above, hybridization is the association of two single strands of complementary nucleic acids to form a hydrogen bonded double strand. It is implicit that if one of the two strands is wholly or partially involved in a hybrid that it will be less able to participate in formation of a new hybrid. Intramolecular and intermolecular hybrids may be formed within the molecules of a single probe if there is sufficient self-complementarity. Such structures can be avoided through careful probe design. By designing a probe so that a substantial portion of the sequence of interest is single stranded, the rate and extent of hybridization can be greatly increased. Computer programs are available to search for this type of interaction. However, in certain instances, it may not be possible to avoid this type of interaction.

Specific primers and sequence specific oligonucleotide probes can be used in a polymerase chain reaction that enables amplification and detection of the viral genomic sequences.

One aspect of the invention relates to specific oligonucleotide primers. The invention provides compositions comprising an oligonucleotide primer for amplifying an *Flaviviridae* nucleic acid wherein said primer is suitable for amplifying a nucleic acid subsequence from a *Flaviviridae* mutation. For example, the primer can be capable of detecting a G to C nucleotide change at nucleotide 1214 of the RNA polymerase region of BVDV. In another example, the primer can be capable of detecting a G to C nucleotide change at nucleotide 8443 of HCV genome.

Amplification and Detection of a *Flaviviridae* Mutation

Another aspect of the invention relates to methods for amplifying and detecting the presence of a *Flaviviridae* mutation.

DNA or RNA can be extracted from a bodily sample, such as blood or tissue material, such as liver, by a variety of techniques known in the art. An unpure sample, for example samples taken from plasma, serum or blood, can be treated with an amount of a reagent effective to open the cells, fluids, tissues, viral capsids or animal cell membranes of the sample, and to expose and/or separate the strand(s) of the nucleic

acid(s), before amplification. This lysing and nucleic acid denaturing step to expose and separate the strands will allow amplification to occur much more readily.

In one embodiment, the invention provides a process to detect a mutation wherein a sample suspected of containing *Flaviviridae* nucleic acid sequence(s) is amplified; the amplified sequence is contacted with an oligonucleotide probe having a sequence complementary to the nucleotide sequence of the mutation; and the sequence is detected by hybridizing the probe to the sequence. In one embodiment, amplification is achieved by the use of the polymerase chain reaction method. In another embodiment, the mutation is a nucleotide change from G to C at position 1214 of the RNA polymerase region of the BVDV genome. In yet another embodiment, the mutation is a nucleotide change from G to C at position 8443 of the HCV genome.

Amplification

The amplification method used can be the polymerase chain reaction (PCR; Saiki et al., 1988), ligase chain reaction (LCR; Landgren et al., 1988; Wu & Wallace, 1989; Barany, 1991), nucleic acid sequence-based amplification (NASBA; Guatelli et al., 1990; Compton, 1991), transcription-based amplification system (TAS; Kwoh et al., 1989), strand displacement amplification (SDA; Duck, 1990; Walker et al., 1992), amplification by means of Q9 replicase (Lizardi et al., 1988; Lomeli et al., 1989), or any other suitable method to amplify nucleic acid molecules known in the art.

Polymerase Chain Reaction

The PCR process for amplification is generally well known in the art (See, for example, U.S. Pat. Nos. 4,683,202 and 4,683,194). The amplification process can involve an enzymatic chain reaction for preparing, in exponential quantities relative to the number of reaction steps involved, a specific nucleic acid sequence, given that the ends of the required sequence are known in sufficient detail that oligonucleotide primers can be synthesized that will hybridize to them, and that a small amount of the sequence is available to initiate the chain reaction. One primer is complementary to the negative (-) strand and the other is complementary to the positive (+) strand. Annealing the primers to denatured nucleic acid followed by extension with an enzyme such as the large

fragment of DNA Polymerase I (Klenow) and nucleotides results in newly synthesized (+) and (-) strands containing the target sequence. Because these newly synthesized sequences are also templates for the primers, repeated cycles of denaturing, primer annealing and extension results in exponential accumulation of the region defined by the primer. The product of the chain reaction will be a discrete nucleic acid duplex with termini corresponding to the ends of the specific primers employed.

Any specific nucleic acid sequence can be produced by the present process. It is only necessary that a sufficient number of bases at both ends of the sequence be known in sufficient detail so that two oligonucleotide primers can be prepared which will hybridize to different strands of the desired sequence and at relative positions along the sequence such that an extension product synthesized from one primer, when it is separated from its template (complement), can serve as a template for extension of the other primer into a nucleic acid of defined length. The greater the knowledge about the bases at both ends of the sequence, the greater can be the specificity of the primers for the target nucleic acid sequence, and thus the greater the efficiency of the process. It will be understood that the word primer as used hereinafter can refer to more than one primer, particularly in the case where there is some ambiguity in the information regarding the terminal sequence(s) of the fragment to be amplified. For instance, in the case where a nucleic acid sequence is inferred from protein sequence information a collection of primers containing sequences representing all possible codon variations based on degeneracy of the genetic code can be used for each strand. One primer from this collection can be substantially conserved with the end of the desired sequence to be amplified.

A specific nucleic acid sequence is produced by using the diagnostic marker nucleic acid containing that sequence as a template. If the target nucleic acid sequence of the sample contains two strands, it is necessary to separate the strands of the nucleic acid before they can be used as the template, either as a separate step or simultaneously with the synthesis of the primer extension products. This strand separation can be accomplished using any suitable denaturing techniques, including physical, chemical or enzymatic means, wherein the word "denaturing", as used herein, includes all such means. One physical method of separating the strands of the nucleic acid involves heating the nucleic acid unit until it is denatured. Typical heat denaturation can involve

temperatures ranging from about 80 - 150 °C for times ranging from about 1 to 10 minutes. Strand separation can also be induced by an enzyme from the class of enzymes known as helicases or the enzyme RecA, which has helicase activity, and in the presence of riboATP is known to denature DNA. Reaction conditions suitable for separating the strands of nucleic acids with helicases are described by Kuhn Hoffmann-Berling, CSH-Quantitative Biology, 43:63 (1978), and techniques for using RecA are reviewed in C. Radding, Ann. Rev. Genetics, 16:405-37 (1982).

If the original nucleic acid containing the sequence to be amplified is single stranded, its complement is synthesized by adding one or two oligonucleotide primers to it. If an appropriate single primer is added, a primer extension product is synthesized in the presence of the primer, an agent for polymerization, and the four nucleoside triphosphates described below. The product will be partially complementary to the single-stranded nucleic acid and will hybridize with the nucleic acid strand to form a duplex of unequal length strands that can then be separated into single strands as described above to produce two single separated complementary strands. Alternatively, two appropriate primers can be added to the single-stranded nucleic acid and the reaction carried out.

If the original nucleic acid constitutes the sequence to be amplified, the primer extension product(s) produced will be completely or substantially completely complementary to the strands of the original nucleic acid and will hybridize with them to form a duplex of equal length strands to be separated into single-stranded molecules.

When the complementary strands of the nucleic acid or acids are separated, whether the nucleic acid was originally double or single stranded, the strands are ready to be used as a template for the synthesis of additional nucleic acid strands. This synthesis is performed under conditions allowing hybridization of primers to templates to occur. Generally it occurs in a buffered aqueous solution, to obtain, for example, a pH range of 7-9. A molar excess (for genomic nucleic acid, usually about 10^8 : 1 primer: template) of the two oligonucleotide primers can be added to the buffer containing the separated template strands. It is understood, however, that the amount of complementary strand cannot be known if the process is used for diagnostic applications, so that the amount of primer relative to the amount of complementary strand cannot be determined with certainty. As a practical matter, however, the amount of primer added will generally be

in molar excess over the amount of complementary strand (template) when the sequence to be amplified is contained in a mixture of complicated long-chain nucleic acid strands. A large molar excess is preferred to improve the efficiency of the process.

5 The deoxyribonucleoside triphosphates dATP, dCTP, dGTP and TTP are also added to the synthesis mixture, either separately or together with the primers, in adequate amounts and the resulting solution is heated to about 90-100 °C for from about 1 to 10 minutes, for example from about 1 to 4 minutes. After this heating period the solution is allowed to cool to room temperature, which is preferable for the primer hybridization. To the cooled mixture is added an appropriate agent for effecting the primer extension reaction (called herein "agent for polymerization"), and the reaction is allowed to occur under conditions known in the art. The agent for polymerization can also be added together with the other reagents if it is heat stable. This synthesis reaction can occur from room temperature to a temperature above which the agent for polymerization no longer functions. Thus, for example, if DNA polymerase is used as the agent, the temperature is generally no greater than about 40 °C. Most conveniently the reaction occurs at room temperature.

20 The agent for polymerization can be any compound or system that can function to accomplish the synthesis of primer extension products, including enzymes. Suitable enzymes for this purpose include, for example, E. coli DNA polymerase I, Klenow fragment of E. coli DNA polymerase I, T4 DNA polymerase, other available DNA polymerases, polymerase muteins, reverse transcriptase(s), and other enzymes, including heat-stable enzymes (i.e., those enzymes that perform primer extension after being subjected to temperatures sufficiently elevated to cause denaturation), which can facilitate combination of the nucleosides in the proper manner to form the primer extension products that are complementary to each nucleic acid strand. Generally, the synthesis will be initiated at the 3'-end of each primer and proceed in the 5' direction along the template strand until synthesis terminates, producing molecules of different lengths. Alternatively, agents for polymerization that initiate synthesis at the 5'-end and proceed toward the 3'-end, using the same process as described above, can be used.

30 The newly synthesized strand and its complementary nucleic acid strand will form a double-stranded molecule under the hybridizing conditions described above if the target sequence is present, and this hybrid is used in the succeeding steps of the process.

In the next step, the sample treated under hybridizing conditions is subjected to denaturing conditions using any of the procedures described above to provide single-stranded molecules if the target sequence is present.

5 New nucleic acid is synthesized on the single-stranded molecules. Additional agents for polymerization, nucleosides and primers can be added if necessary for the reaction to proceed under the conditions prescribed above. Again, the synthesis will be initiated at one end of each of the oligonucleotide primers and will proceed along the single strands of the template to produce additional nucleic acid. After this step, half of the extension product will consist of the specific nucleic acid sequence bounded by the
10 two primers.

The steps of denaturing and extension product synthesis can be repeated as often as needed to amplify the target nucleic acid sequence to the extent necessary for detection. As will be described in further detail below, the amount of the specific nucleic acid sequence produced will accumulate in an exponential fashion.

15 When it is desired to produce more than one specific nucleic acid sequence from the first nucleic acid or mixture of nucleic acids, an appropriate number of different oligonucleotide primers are utilized. For example, if two different specific nucleic acid sequences are to be produced, four primers are utilized. Two of the primers are specific for one of the specific nucleic acid sequences and the other two primers are specific for
20 the second specific nucleic acid sequence. In this manner, each of the two different specific sequences can be produced exponentially by the present process.

The present invention can be performed in a step-wise fashion where after each step new reagents are added, or simultaneously or in a single-step manner, where all reagents are added at the initial step, or partially step-wise and partially simultaneous,
25 where fresh reagent is added after a given number of steps. If a method of denaturation, for example heat, is employed, which can inactivate the agent for polymerization, as in the case of a heat-labile enzyme, then it is necessary to replenish the agent after every strand separation step. The simultaneous method can be utilized when an enzymatic means is used for the strand separation step. In the simultaneous procedure, the reaction
30 mixture can contain, in addition to the nucleic acid strand(s) with the desired sequence, the strand-separating enzyme (e.g., helicase), an appropriate energy source for the strand-separating enzyme (e.g. rATP), the four nucleoside triphosphates, the oligonucleotide

primers in molar excess, and the agent for polymerization (e.g., Klenow fragment of E. coli DNA polymerase I).

If heat is used for denaturation in a simultaneous process, a heat-stable agent such as a thermostable polymerase can be employed that will operate at an elevated temperature, for example from about 50-105 °C depending on the agent, at which temperature the nucleic acid will consist of single and double strands in equilibrium. For smaller lengths of nucleic acid, lower temperatures of about 40-50 °C can be employed. The upper temperature range will depend on the temperature at which the enzyme will degrade or the temperature above which an insufficient level of primer hybridization will occur. Such a heat-stable enzyme is described, e.g., by A. S. Kaledin et al., *Biokhimiya*, 45, 644-651 (1980). For this constant temperature reaction to succeed, the primers have their 3' ends within 6-8 base pairs of each other. Each step of the process will occur sequentially notwithstanding the initial presence of all the reagents. Additional materials can be added as necessary. After the appropriate length of time has passed to produce the desired amount of the specific nucleic acid sequence, the reaction can be halted by inactivating the enzymes in any known manner or by separating the components of the reaction.

The amplification can also be carried out using a temperature-cycling reaction wherein the temperature is increased incrementally to allow for extension, annealing and denaturation using a heat-stable enzyme.

The process of the present invention can be conducted continuously. In one embodiment of an automated process, the reaction can be cycled through a denaturing region, a reagent addition region, and a reaction region. In another embodiment, the enzyme used for the synthesis of primer extension products can be immobilized in a column. Other reaction components can be continuously circulated by a pump through the column and a heating coil in series, thus the nucleic acids produced can be repeatedly denatured without inactivating the enzyme.

Flaviviridae genotyping using PCR techniques is commonly known in the art. Further, after PCR has been performed on samples suspected of containing a *Flaviviridae*, the *Flaviviridae* genome can be sequenced.

Detection of Hybridization of the Probe and Target Sequence

Suitable assay formats for detecting hybrids formed between probes and target nucleic acid sequences in a sample are known in the art (Sambrook et al., 1985). Detection of hybridization can be accomplished whether or not the nucleic acid has been amplified.

One method of detection is through the use of a labeled probe capable of hybridizing with the unamplified or amplified nucleic acid sequence and determining if the probe has hybridized. Such probe necessarily contains the nucleotide that is suspected of being mutated, such as 1214 of the RNA polymerase of BVDV or nucleotide 8443 of the HCV genome.

Oligonucleotides can be labeled by incorporating a label detectable by spectroscopic, photochemical, biochemical, immunochemical, or chemical means. Useful labels include ^{32}P , fluorescent dyes, electron-dense reagents, enzymes (as commonly used in ELISAs), biotin, or haptens and proteins for which antisera or monoclonal antibodies are available.

The nucleic acid can be detected by analyzing it by Northern or Southern blots with or without radioactive probes. In one embodiment, a small sample of DNA from, e.g., peripheral blood suspected of containing *Flaviviridae*, is analyzed via a Southern blot technique using oligonucleotide probes to detect the specific nucleic acid viral marker. In another embodiment, a small sample of DNA from, e.g., peripheral blood suspected of containing *Flaviviridae*, is first amplified and then analyzed via a Southern blot technique using oligonucleotide probes to detect the specific nucleic acid viral marker.

Another method involves the oligomer restriction technique (such as described in U.S. Pat. No. 4,683,194). In this procedure, an amplified nucleic acid is denatured and hybridized in solution to a labeled oligonucleotide probe which hybridizes specifically to the target sequence (i.e., spans the particular conserved region contained by the primers) and spans at least one restriction site of interest. The duplex formed between target and probe will reconstitute the restriction site, and when cleaved with restriction enzyme, such as, e.g., BglI, PvuII, or HinfI, releases a labeled probe fragment which can be resolved from the full-length probe by gel electrophoresis. The resulting gel is then

autoradiographed. Analysis of the amplified product by this method can be rapid, i.e., within a few hours.

Another method that can be used to analyze an amplified product is the dot blot method. In a dot-blot method, amplified target DNA is immobilized on a solid support, such as a nylon membrane. The membrane-target complex is incubated with labeled probe under suitable hybridization conditions, unhybridized probe is removed by washing under suitably stringent conditions, and the membrane is monitored for the presence of bound probe.

An alternate format is a "reverse" dot-blot format, in which an amplified target DNA is labeled and the probes are immobilized on a solid support, such as a nylon membrane (see Saiki et al., 1989, Proc. Natl. Acad. Sci. USA 86:6230, and PCT Patent Publication No. 89/11548). The target DNA is typically labeled during amplification by the incorporation of one or more labeled primers. One or both of the primers can be labeled. The membrane-probe complex is incubated with the labeled amplified target DNA under suitable hybridization conditions, unhybridized target DNA is removed by washing under suitably stringent conditions, and the filter is then monitored for the presence of bound target DNA.

Alternatively, the reverse dot-blot assay can be carried out using a solid support having a plurality of probe hybridization sites or wells. For example, a microwell plate is particularly useful in large scale clinical applications of the present methods. Probes can be immobilized to a microwell plate either by passive binding or through a protein intermediate, such as bovine serum albumin (BSA), which adheres to microwell plates. Reverse dot-blot methods carried out in a microwell plate are described in U.S. Pat. No. 5,232,829, and Loeffelholz et al, 1992, J. Clin. Microbiol. 30(11):2847-2851. In another embodiment of the invention, a reverse dot-blot assay is carried out using microwell plates, and the primers are labeled with biotin, as described in Levenson and Chang, 1989, in PCR Protocols: A Guide to Methods and Applications, (Innis et al., eds., Academic Press. San Diego) pages 99-112. The probes are conjugated with BSA (see Tung et al., 1991, Bioconjugate Chem. 2:464-465, incorporated herein by reference) and immobilized on a microwell plate. Following amplification using the labeled primers and hybridization with the immobilized probes, the amplified nucleic acid is detected by first binding the biotin to avidin-horseradish peroxidase (A-HRP) or streptavidin-horseradish

peroxidase (SAHRP), which is then detected by carrying out a reaction in which the HRP catalyzes a color change of a chromogen.

In an alternative method of immobilizing hybridization duplexes for detection, BSA-conjugated probes are bound to magnetic microparticles. The bound probes are hybridized in solution to labeled amplification product. Following hybridization, probe-target duplexes are removed from the solution magnetically, and the magnetically immobilized hybridization duplexes are then detected.

Another method of detection is referred to as a 5'-nuclease assay in which the labeled detection probes are added during the PCR amplification process. The probes are modified so as to prevent them from acting as primers for DNA synthesis. Any probe which hybridizes to target DNA during each synthesis step, i.e., during primer extension, is degraded by the 5' to 3' exonuclease activity of the DNA polymerase, e.g., Taq DNA polymerase. The degradation product from the probe is then detected. Thus, the presence of probe breakdown product indicates both that hybridization between probe and target DNA occurred and that the amplification reaction occurred. See also for example, U.S. Pat. No. 5,210,015.

The assay formats described above typically utilize labeled oligonucleotides to facilitate detection of the hybrid duplexes. Oligonucleotides can be labeled by any of the previously mentioned techniques, such as incorporating a label detectable by spectroscopic, photochemical, biochemical, immunochemical, or chemical means. Useful labels include ³² P, fluorescent dyes, electron-dense reagents, enzymes (as commonly used in ELISAS), biotin, or haptens and proteins for which antisera or monoclonal antibodies are available.

An alternative method for detecting the amplification of a *Flaviviridae* nucleic acid is by monitoring the increase in the total amount of double-stranded DNA in the reaction mixture (as described in Higuchi et al., 1992, Bio/Technology 10:413-417; Higuchi et al., 1993, Bio/Technology 11:1026-1030; and European Patent Publication Nos. 487,218 and 512,334). The detection of double-stranded target DNA relies on the increased fluorescence that ethidium bromide (EtBr) and other DNA binding labels exhibit when bound to double-stranded DNA. The increase of double-stranded DNA resulting from the synthesis of target sequences results in a detectable increase in fluorescence.

Yet another method useful for detecting *Flaviviridae* mutations is through reverse hybridization assays. This is especially useful if a multitude of probes are involved. In one embodiment the selected set of probes are immobilized to a solid support in known distinct locations (dots, lines or other figures). In another embodiment the selected set of probes can be immobilized to a membrane strip in a line fashion. Said probes can be immobilized individually or as mixtures to delineated locations on the solid support. In a specific embodiment, a line probe assay can be used to screen for *Flaviviridae* genotypes containing the mutation of the present invention. The line probe assay involves multiple probes that are immobilized in parallel lines on a membrane, then a reverse hybridization of amplified nucleic acid fragments is performed. The hybrid can then be detected via a biotin-streptavidin coupling with a non-radioactive color developing system. See, for example, WO 97/40193.

Flaviviridae genotyping techniques can also be used to analyze the presence of *Flaviviridae* mutations. For example, sequence based phylogenetic analysis, differential hybridization, PCR or fragment length polymorphism can be used.

Detection of *Flaviviridae* Protein/Peptide Markers

In another embodiment, the invention provides a process for detecting viral markers diagnostic for long term 2'-branched nucleoside therapy failure for *Flaviviridae* infection wherein a sample containing *Flaviviridae* protein, peptides, or peptide fragments is analyzed for such viral markers. The proteins, peptides, or peptide fragments correlated with therapy failure can be detected by any generally applicable protein detection technology known in the art, including western blot, two dimensional gel electrophoresis, enzyme linked immunosorbent assays (ELISA), enhanced chemiluminescence (ECL), immunohistochemistry, ELI-Spot assays, peptide sequencing, or antibody based protein array technology. For example, protein expression of *Flaviviridae* viral markers diagnostic for 2'-branched nucleoside treatment can be analyzed with classical immunohistological methods. In these, the specific recognition is provided by the primary antibody (polyclonal or monoclonal) to the specific viral marker, but the secondary detection system can utilize fluorescent, enzyme, or other conjugated secondary antibodies. As a result, an immunohistological staining of the *Flaviviridae* infected tissue section for pathological examination is obtained.

Another method for detecting proteins, peptides, or peptide fragments that are diagnostic for long term response of *Flaviviridae* carriers to 2'-branched nucleoside therapy is the western blot. In brief, a sample containing *Flaviviridae* protein, peptide, or peptide fragments is separated via means of an electrophoretic gel. The separated proteins are then transferred to a medium such as nitrocellulose. Antibodies having a detectable label such as streptavidin-alkaline phosphatase reactive to specific *Flaviviridae* amino acid sequences correlated to 2'-branched nucleoside failure are then contacted onto the nitrocellulose medium containing the *Flaviviridae* amino acid sequences. Reactive antibodies will bind with the corresponding *Flaviviridae* amino acid sequence, and can be detected using a reagent such as nitoblu tetrazolium and 5-bromo-4-chloro-3-indlyl phosphate (BCIP). See, for example, Jalkanen, M., et al., J. Cell. Biol. 101:976-985 (1985); Jalkanen, M., et al., J. Cell. Biol. 105:3087-3096 (1987).

Alternatively, reactive antibodies present in the sera of *Flaviviridae* carriers can be used to detect the presence of *Flaviviridae* viral markers that are diagnostic of 2'-branched nucleoside treatment. Any known antibody assay technique known by those skilled in the art, including enzyme immunoassay (EIA), can be used. For example, in one embodiment a sample from an *Flaviviridae* carrier containing *Flaviviridae* specific antibodies is contacted with a solid support array containing specific *Flaviviridae* peptides correlated with 2'-branched nucleoside therapy success and/or failure. The reactive antibodies are then detected using rabbit anti-human IgG antibodies labeled with streptavidin-alkaline phosphatase and a reagent containing exposed to nitoblu tetrazolium and 5-bromo-4-chloro-3-indlyl phosphate.

Another method for detecting proteins, peptides, or peptide fragments that are diagnostic for long term response of *Flaviviridae* carriers to 2'-branched nucleoside therapy is by sequencing the protein, peptide, or peptide fragment using techniques known to one of ordinary skill in the art (see, for example, Matsudaira, P., J Biol Chem 262: 10035-10038, (1987); Salinovich, O. and Montelano, R., Anal. Biochem. 156: 341, (1986); Tarr, G.E.: Manual Edman Sequencing System. In: Shively, J.E., (ed.) Methods of Protein Microcharacterization. The Humana Press Inc., Clifton, NJ, 1986, pp. 155-194; and Fernandez, J., Andrews, L. and Mische, S., Anal. Biochem. 218: 112-117, (1994)). For example, one could use the Edman technique to determine the amino acid sequence of the peptide. In brief, the Edman chemistry removes amino acid residues

from the N-terminus of a protein/peptide, one at a time in sequence. Each cycle of Edman chemistry, needed for the removal of each amino acid residue, consists of three steps: a coupling with phenyl isothiocyanate (PITC) under mildly alkaline conditions to form a phenylthiocarbamyl (PTC)-peptide; cleavage to release the first residue as its anilinothiazolinone (ATZ)-amino acid derivative; conversion of the ATZ derivative to a more stable phenylthiohydantoin (PTH)-amino acid derivative. The PTH-amino acid residue, removed in each cycle of Edman degradation, is identified by small or micro bore RP-HPLC. A full description of the process and possible pitfalls is given by Tarr, G.E.: Manual Edman Sequencing System. In: Shively, J.E., (ed.) Methods of Protein Microcharacterization. The Humana Press Inc., Clifton, NJ, 1986, pp. 155- 194. Alternatively, if a sample yields no N-terminal sequence, the N-terminal residue is blocked, degraded during preparative procedures, or for steric reasons unavailable to the Edman chemistry reagents, then the sample can be subjected to controlled specific proteolysis, where the peptides are fractionated and then analyzed. This fractionation approach is described by Fernandez, J., Andrews, L. and Mische, S.: An improved procedure for enzymatic digestion of polyvinylidene difluoride-bound proteins for internal sequence analysis. Anal. Biochem. 218: 112-117, 1994.

Arrays

Another aspect of the present invention provides the use of DNA, RNA or peptide arrays to detect *Flaviviridae* nucleic acid viral markers. Such arrays include DNA macroarrays, DNA microarrays, and DNA microchips. DNA arrays, for example, have been described in U.S. Pat. No. 5,837,832, U.S. Pat. No. 5,807,522, U.S. Pat. No. 6,007,987, U.S. Pat. No. 6,110,426, WO 99/05324, 99/05591, WO 00/58516, WO 95/11995, WO 95/35505A1, WO 99/42813, JP10503841T2, GR3030430T3, ES2134481T3, EP804731B1, DE69509925C0, CA2192095AA, AU2862995A1, AU709276B2, AT180570, EP 1066506, and AU 2780499. Such arrays can be incorporated into computerized methods for analyzing hybridization results when the arrays are contacted with prepared sample nucleotides, for example, as described in PCT Publication WO 99/05574, and U.S. Pat. Nos. 5,754,524; 6,228,575; 5,593,839; and 5,856,101. Methods for screening for disease markers are also known to the art, for example, as described in U.S. Pat. Nos. 6,228,586; 6,160,104; 6,083,698; 6,268,398;

6,228,578; and 6,265,174. Further descriptions of DNA array methods can, for example be found in: Shoemaker D.D. et al., *Nature* 409(6822):922-927 (2001); Kane M.D., et al., *Nucleic Acids Res* 28(22):4552-7 (2000); Taton TA, et al., *Science*. 289(5485):1757-60 (2000); Jörg Reichert et al., *Anal. Chem.*, 72(24):6025 -6029 (2000); Reinke V, *Mol Cell* 6(3):605-16 (2000); Marx J. *Science* 289:1670-1672 (2000); Lockhart D.J. et al., *Nature* 405(6788):827-836 (2000); Cortese J.D., *The Scientist* 14[17]:25 (2000); Cortese J.D., *The Scientist* 14[11]:26 (2000); Fritz J. et al., *Science*. 288(5464):316-8 (2000); Mark Schena (Ed.), *Microarray Biochip Technology*, Eaton Publishing Company, Distributed by TeleChem / arrayit.com; Scherf U., et al., *Nat Genet.*24(3):236-44 (2000); Ross D.T. et al., *Nat Genet.* 24(3):227-35 (2000); Walt D.R., *Science* 287: 451-452 (2000); Afshari C.A. et al., *Cancer Res* 59(19):4759-60 (1999); Gwynne P. and Page G., *Science*, 1999 August 6. (special advertising supplement; has a list of microarray-related companies); Baldwin D. et al., *Curr Opin Plant Biol* 2(2):96-103 (1999); Pollack J.R. et al., *Nat Genet* 23(1):41-6 (1999); Khan J. et al., *Electrophoresis* 20(2):223-9 (1999); Gerhold D. et al., *Trends Biochem Sci* 24(5):168-73 (1999); Ekins R. and Chu F.W., *Trends in Biotechnology* 17:217-218 (1999); Nuwaysir, E.F. et al., *Molecular Carcinogenesis* 24:153-159 (1999); Sinclair, B. *The Scientist*, 13(11):18-20 (1999); The Chipping Forecast, *Nature Genetics* (January 1999 Supplement); Schena, M. and Davis, R.W. *Genes, Genomes and Chips*. In *DNA Microarrays: A Practical Approach* (ed. M. Schena), Oxford University Press, Oxford, UK, 1999; Marton M.J. et al., *Nat Med.* 4(11):1293-301 (1998); Wang D.G. et al., *Science* 280(5366):1077-82 (1998); Schena, M. and R.W. Davis. *Parallel Analysis with Biological Chips*. in *PCR Methods Manual* (eds. M. Innis, D. Gelfand, J. Sninsky), Academic Press, San Diego, 1998; Lemieux, B. et al., *Molecular Breeding* 4:277-289 (1998); Schena, M. et al., *Trends in Biotechnology* 16:301-306 (1998); Service, R.F., *Science* 282(5388):396-399 (1998); Service, R.F., *Science* 282(5388):399-401 (1998); Kricka, L., *Nature Biotechnology* 16:513 (1998); Housman, D., *Nature Biotechnology* 16(6):492-493 (1998); Ramsay, G., *Nature Biotechnology* 16(1):40-44 (1998); Marshall, A. et al., *Nature Biotechnology* 16(1):27-31 (1998); Kononen J. et al., *Nat. Med.* 4(7):844-847 (1998); Blanchard, A.P. (1998) *Synthetic DNA Arrays*; in *Genetic Engineering*, Vol. 20, pp. 111-123, edited by J.K. Setlow, Plenum Press, New York; Proudnikov D. et al., *Anal Biochem* 259(1):34-41 (1998); Chen J.J. et al., *Genomics* 51(3):313-24 (1998); Wallace R. W., *Molecular Medicine Today* 3:384-389 (1998); Covacci, A. et al., *Drug Development Research*

41:180-192 (1997); Forozan, F. et al., Trends in Genetics 13:405-409 (1997); Blanchard, A.P. & L. Hood, Nature Biotechnology 14:1649 (1996); Blanchard, A.P. et al., Biosensors & Bioelectronics 11:687-690 (1996); DeRisi J. et al., Nat Genet 14(4):457-60 (1996); Shalon D. et al., Genome Res 6(7):639-45 (1996); Schena M. et al., Proc Natl Acad Sci U S A 93(20):10614-9 (1996); and Schena M. et al., Science 270(5235):467-70 (1995).

Probes on an array can be of varying lengths, including, but not limited to, as short as about 10-30 nucleotides long or as long as an entire *Flaviviridae* gene or *Flaviviridae* clone, which can be up to several kilobases. In addition, sequences of the various lengths as those described in **Tables 2** and **3** (Seq ID Nos. 1-62) can be used as probes. The array can be designed such that all probes on the array can hybridize to their corresponding genes at about the same hybridization stringency. Probes for arrays should be unique at the hybridization stringencies used. A unique probe is only able to hybridize with one type of nucleic acid per target. A probe is not unique if at the hybridization stringency used, it hybridizes with nucleic acids derived from two different genes, i.e. related genes, or non-homologous sequences. The homology of the sequence of the probe to the gene and the hybridization stringency used help determine whether a probe is unique when testing a selected sample. Probes also may not hybridize with different nucleic acids derived from the same gene, i.e., splice variants. Since the splice variants of interest are known, several different probe sequences can be chosen from the target gene sequence of interest for an array, such that each probe can only hybridize to nucleic acid derived from one of the splice variants. In one embodiment, arrays containing Seq ID Nos. 1-62 are used at hybridization conditions allowing for selective hybridization. At conditions of selective hybridization, probes hybridize with nucleic acid from only one identified sequence. At conditions of selective hybridization, probes hybridize with nucleic acid from only one identified sequence. In another embodiment, arrays containing any *Flaviviridae* sequence of interest are used at hybridization conditions allowing for selective hybridization. At conditions of selective hybridization, probes hybridize with nucleic acid from only one identified sequence.

In one embodiment, the use of the microarray first requires amplification of genes of interest, such as by reverse transcription of mRNA or total RNA followed by polymerase chain reaction using methods known in the art. As the nucleic acid is copied,

it is tagged with a label that can be used in the detection and quantitation methods known in the art. The nucleic acid can be labeled with radioactive or non-radioactive labels, but preferably contain fluorescent labels. The labeled nucleic acid is introduced to the microarray containing the sequence probes of interest and allowed to react for a period of time. Thereafter, the substrate is washed free of extraneous materials, leaving the nucleic acids on the target bound to the fixed probe molecules allowing for detection and quantitation by methods known in the art such as by autoradiograph, liquid scintillation counting, and/or fluorescence. As improvements are made in hybridization and detection techniques, they can be readily applied by one of ordinary skill in the art. As is well known in the art, if the probe molecules and target molecules hybridize by forming a strong non-covalent bond between the two molecules, it can be reasonably assumed that the probe and target nucleic acid are essentially completely complementary if the annealing and washing steps are carried out under conditions of high stringency. The detectable label provides a means for determining whether hybridization has occurred. By obtaining an image of the array with a detection and quantitation method known in the art such as autoradiography, liquid scintillation counting, or fluorescence it can be determined if and to what extent *Flaviviridae* gene sequences are present, by comparing intensities at specific locations on the array. High quantitation signals indicate that a particular sequence is present in a prepared sample, and an absent quantitation signal shows that a particular sequence is not present. The presence of various gene sequences under different conditions can be directly compared, such as prior to 2'-branched nucleoside treatment and during 2'-branched nucleoside treatment. Similarly, it can be determined what sequences are present in response to certain stimuli such as a 2'-branched nucleoside.

In one embodiment, the *Flaviviridae* sequence profile of a patient can be tracked over time using DNA array technologies. In an alternative embodiment, a patient with *Flaviviridae* receiving 2'-branched nucleoside as a modality, or other anti-*Flaviviridae* modality, can be monitored, over time, for changes in the aforementioned *Flaviviridae* genomic sequences in response to the treatment.

Arrays containing Seq ID Nos. 1-62, or any other identified *Flaviviridae* sequence of interest, can be made by any array synthesis method known in the art, such as spotting technology or solid phase synthesis via photolithography. Arrays can also be

printed on solid substrates, e.g., glass microscope slides. Before printing, slides are prepared to provide a substrate for binding, as known in the art. Arrays can be printed using any printing techniques and machines known in the art. Printing involves placing the probes on the substrate, attaching the probes to the substrate, and blocking the substrate to prevent non-specific hybridization, as known in the art. Preferably the arrays of this invention are synthesized by solid phase synthesis using a combination of photolithography and combinatorial chemistry. Some of the key elements of probe selection and array design are common to the production of all arrays. Strategies to optimize probe hybridization, for example, are invariably included in the process of probe selection. Hybridization under particular pH, salt, and temperature conditions can be optimized by taking into account melting temperatures and by using empirical rules that correlate with desired hybridization behaviors (as described in Keller, G. H., and M. M. Manak (1987) DNA Probes, Stockton Press, New York, N.Y., pp. 169-170, hereby incorporated by reference). Computer models can be used for predicting the intensity and concentration-dependence of probe hybridization.

Moderate to high stringency conditions for hybridization are known in the art. An example of high stringency conditions for a blot are hybridizing at 68° C in 5×SSC/5× Denhardt's solution/0.1% SDS, and washing in 0.2×SSC/0.1% SDS at room temperature. An example of conditions of moderate stringency are hybridizing at 68° C in 5×SSC/5× Denhardt's solution/0.1% SDS and washing at 42° C in 3×SSC. The parameters of temperature and salt concentration can be varied to achieve the desired level of sequence identity between a probe and a target nucleic acid. See, for example, Ausubel et al. (1995) Current Protocols in Molecular Biology, John Wiley & Sons, NY, N.Y., for further guidance on hybridization conditions. The melting temperature is described by the following formula (Beltz, G. A. et al., [1983] Methods of Enzymology, R. Wu, L. Grossman and K. Moldave [Eds.] Academic Press, New York 100:266-285). $T_m = 81.5^{\circ} \text{C} + 16.6 \log[\text{Na}^+] + 0.41(+G+C) - 0.61(\% \text{ formamide}) - 600/\text{length of duplex in base pairs}$.

Nucleic acids useful in this invention can be created by Polymerase Chain Reaction (PCR) amplification. PCR products can be confirmed by agarose gel electrophoresis. PCR is a repetitive, enzymatic, primed synthesis of a nucleic acid sequence. This procedure is well known and commonly used by those skilled in this art

(see, for example, Mullis, U.S. Pat. Nos. 4,683,195, 4,683,202, and 4,800,159; Saiki et al., Science 230:1350-1354 (1985)). PCR is used to enzymatically amplify a DNA fragment of interest that is flanked by two oligonucleotide primers that hybridize to opposite strands of the target sequence. The primers are oriented with the 3'-ends pointing towards each other. Repeated cycles of heat denaturation of the template, annealing of the primers to their complementary sequences, and extension of the annealed primers with a DNA polymerase result in the amplification of the segment defined by the 5'-ends of the PCR primers. Since the extension product of each primer can serve as a template for the other primer, each cycle essentially doubles the amount of DNA template produced in the previous cycle. This results in the exponential accumulation of the specific target fragment, up to several million-fold in a few hours. By using a thermostable DNA polymerase such as the Taq polymerase, which is isolated from the thermophilic bacterium *Thermus aquaticus*, the amplification process can be completely automated. Other enzymes that can be used are known to those skilled in the art.

Alternatively, probes made of peptide nucleic acids (PNAs) can be used as substitutes for probes made of oligonucleotides for the same uses as described above. The substitution of PNAs for oligonucleotides is well known in the art: The synthesis of peptide nucleic acids via preformed monomers has been described, for example, in PCT patent applications WO 92/20702 and WO 92/20703. Recent advances have also been reported on the synthesis, structure, biological properties, and uses of PNAs. See, for example, PCT Patent application WO 93/12129, U.S. Pat. No. US6617422 to Nielsen P.E. et al., U.S. Pat. No. 5,539,083 to Cook et al., U.S. Patent application US20030059789A1, U.S. Pat. No. US6475721 to Kleiber et al., Egholm et al., Nature :365, 566-568 (1993), Nielsen et al., Science 254:1497-1500 (1991); and Egholm et al., J. Am. Chem. Soc., 114:1895-1897 (1992).

Kits

A suitable test kit for use in an assay to determine the resistance status of a *Flaviviridae* sample to 2'-Branched nucleoside which makes use of a methodology according to one aspect of the invention, comprises (1) an oligonucleotide being

complementary to a region of the wild-type DNA sequence (or its corresponding RNA) or to a region of the mutant DNA sequence as described herein; (2) materials required for polymerization of the nucleic acid from the 3'-end of the oligonucleotide; and (3) a means for determining the presence of an oligonucleotide primer extended product. Polymerization materials include appropriate enzymes, buffers, washing solutions, labels and substrates for the label, if necessary. If PCR is used to amplify nucleic acid then additional materials such as appropriate oligonucleotide primers that will amplify a region of the wild-type DNA sequence (or its corresponding RNA) or a region of the mutant DNA sequence as described herein (or its corresponding RNA) and dNTP's (deoxynucleoside triphosphates) should be included. Instructions for conducting the assay can also be included.

A suitable test kit for use in an assay to determine the sensitivity of a *Flaviviridae* sample to interferon which makes use of a methodology according to another aspect of the invention comprises an oligonucleotide being complementary to a region of the wild-type DNA sequence (or its corresponding RNA) or to the pertinent region of the mutant DNA sequence, along with materials required to permit hybridization. Such materials include appropriate buffers, washing solutions, labels, and substrates for the labels, if necessary. In one embodiment, the oligonucleotide is labeled. If PCR is used to amplify nucleic acid prior to hybridisation then additional materials such as appropriate oligonucleotide primers that will amplify a region of the wild-type DNA sequence (or its corresponding RNA) or a region of the mutant DNA sequence, appropriate enzymes and dNTP's (deoxynucleotide triphosphates) should be included. Instructions for conducting the assay can also be included.

In another embodiment, the invention provides a kit for the detection of a marker of resistance to long term 2'-branched nucleoside treatment of a *Flaviviridae* infection. The kit can contain a compartment which contains an oligonucleotide probe which binds substantially to a nucleic acid subsequence of the *Flaviviridae* that contains the diagnostic marker. Alternatively, the kit contains peptide nucleic acid (PNA) or other antisense mimic probe in substitution for the oligonucleotide. The kit can also contain reagents to detect the hybridization of the probe to the *Flaviviridae* nucleic acid viral marker. The present invention also includes kits that can contain a primer for the PCR amplification of *Flaviviridae* nucleic acids. A kit can also contain a means for detecting

amplified *Flaviviridae* nucleic acids, such as an oligonucleotide or peptide nucleic acid probe. In some cases, the probe is fixed to an appropriate support membrane. Other optional components of the kit include, for example, an agent to catalyze the synthesis of primer extension products, the substrate nucleoside triphosphates, means used to label (for example, an avidin-enzyme conjugate and enzyme substrate and chromogen if the label is biotin), the appropriate buffers for PCR or hybridization reactions, and instructions for carrying out the present method.

In addition, the kit can have a container which includes a positive control containing one or more nucleic acids with a sequence of the *Flaviviridae* viral genome correlated with therapy failure and/or a container including a negative control without such nucleic acids. Moreover, the kit can have a container for a restriction enzyme capable of cleaving a nucleic acid containing the target sequence at a site contained in a sequence in the probe.

The invention also provides a kit for the detection and/or genetic analysis of one or more viral markers of *Flaviviridae* that are correlated with therapy failure that can be present in a biological sample comprising the following components: (i) when appropriate, a means for releasing, isolating or concentrating the nucleic acids present in the sample; (ii) when appropriate, at least one suitable primer pair; (iii) at least two probes as defined above, possibly fixed to a solid support; (iv) a hybridization buffer, or components necessary for producing said buffer; (v) a wash solution, or components necessary for producing said solution; (vi) when appropriate, a means for detecting the hybrids resulting from the preceding hybridization; (vii) when appropriate, a means for attaching said probe to a known location on solid support; and/or (viii) instructions for carrying out the present method.

Furthermore, the invention also provides for a kit that contains peptide or peptide fragments corresponding to viral markers correlated with 2'-branched nucleoside therapy failure that can be used in an immunoassay to detect the presence of reactive antibodies in a sample. The peptide can be in a stabilized solution or lyophilized. Such kit can contain an appropriate solution for hydrolyzing a lyophilized peptide. The kit can also contain an appropriate solid medium for blotting the aforementioned peptide on. The kit can also contain an appropriate reagent for detecting the presence of reactive antibodies to the peptide, such as an anti-human IgG antibody labeled with streptavidin-alkaline

phosphatase. Furthermore, the kit can contain a detection agent such as nitobluetrazolium and 5-bromo-4-chloro-3-indlyl phosphate (BCIP).

Alternatively, the kit can contain antibodies reactive to specific peptide sequences associated with 2'-branched nucleoside therapy.

5

IV. Treatment of *Flaviviridae* Infections

Combination or Alternation Treatment with anti-*Flaviviridae* Agents

Drug-resistant variants of *Flaviviridae* can emerge after prolonged treatment with an antiviral agent. Drug resistance most typically occurs by mutation of a gene that encodes for an enzyme used in viral replication. The efficacy of a drug against *Flaviviridae* infection can be prolonged, augmented, or restored by administering the compound in combination or alternation with a second, and perhaps third, antiviral compound that induces a different mutation from that caused by the principle drug. Combination therapy induces multiple simultaneous stresses on the virus. The pharmacokinetics, biodistribution, or other parameters of the drug can be altered by such combination or alternation therapy.

The present invention provides methods to achieve optimal treatment of a *Flaviviridae* infection through administration of a 2'-branched nucleoside, or a pharmaceutically acceptable prodrug and/or salt thereof, to a human in need of therapy in combination and/or alternation with one or more drugs that directly or indirectly induce a mutation in the viral genome at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XRXSGXXXT, of domain B of the RNA polymerase region, and/or one or more drugs that is associated with such mutation.

25

Interferon Treatment of Mutant *Flaviviridae* Infections

Another aspect of the present invention provides a method to treat and/or to substantially cure a *Flaviviridae* infection in a host infected with a *Flaviviridae* that contains a Serine to Threonine mutation at the conserved serine amino acid residue of

Domain B of the RNA polymerase region of a Flaviviridae (**Figure 11**) by administering a therapeutically effective amount of interferon. In one embodiment, a method is provided to treat and/or to substantially cure a BVDV infection that contains a serine to threonine mutation at amino acid position 405 of the RNA polymerase region by administering a therapeutically effective amount of interferon. In another embodiment, a method is provided to treat and/or to substantially cure a HCV infection that contains a serine to threonine mutation at amino acid position 282 of the RNA polymerase region. of HCV by administering a therapeutically effective amount of interferon. In a specific embodiment, interferon alpha-2b is administered to treat and/or to substantially cure the *Flaviviridae* infection.

In a further embodiment, a method is provided to treat and/or to substantially cure a *Flaviviridae* infection in a host suspected of being infected with BVDV comprising: (i) obtaining a viral sample from the host; (ii) identifying whether the *Flaviviridae* in the sample contains a Threonine at amino acid residue 405 of the RNA polymerase; and (iii) administering an effective amount of interferon to the host infected with a *Flaviviridae* that contains a Serine to Threonine mutation at amino acid 405 of the RNA polymerase region.

In another embodiment, a method is provided to treat and/or to substantially cure a *Flaviviridae* infection in a host suspected of being infected with HCV comprising: (i) obtaining a viral sample from the host; (ii) identifying whether the *Flaviviridae* in the sample contains a Threonine at amino acid residue 282 of the RNA polymerase; and (iii) administering an effective amount of interferon to the host infected with a *Flaviviridae* that contains a Serine to Threonine mutation at amino acid 282 of the RNA polymerase region.

Interferons include: Intron-A (interferon alpha-2b) by Schering, PEG-INTRON (pegylated interferon alpha-2b) by Schering, Roferon-A (interferon alfa-2a) by Roche, PEGASYS (pegylated interferon alfa-2a) by Roche, INFERGEN (interferon alfacon-1) by InterMune, OMNIFERON (natural interferon) by Viragen, ALBUFERON by Human Genome Sciences, REBIF (interferon beta-1a) by Ares-Serono, Omega Interferon by BioMedicine, Oral Interferon Alpha by Amarillo Biosciences, and Interferon gamma-1b by InterMune.

Identification of the Serine to Threonine mutation at amino acid position 405 of the RNA polymerase region of BVDV, or amino acid position 282 of the RNA polymerase region of HCV, can be accomplished by detecting the presence of a mutation in the *Flaviviridae* genome that would allow for an amino acid change from Serine to Threonine. In one embodiment, the presence of cytidine at nucleotide 1214 of the RNA polymerase region of BVDV (where nucleotide 1214 of the RNA polymerase region corresponds to nucleotide 11,136 of the BVDV genome) can be used to detect the amino acid change. In other embodiments, the following double mutations can be detected: 1214 (G to C) and 1215 (C to A); 1214 (G to C) and 1215 (C to G); or 1214 (G to C) and 1215 (C to U), which result in an amino acid change from serine to threonine at position 405 of the RNA polymerase region of BVDV. In another embodiment, the presence of cytidine at nucleotide 8443 of the HCV genome can be used to detect the amino acid change. In other embodiments, the following double mutations can be detected: 8443 (G to C) and 8444 (C to A); 8443 (G to C) and 8444 (C to G); or 8443 (G to C) and 8444 (C to U), which result in an amino acid change from serine to threonine at position 282 of the RNA polymerase region of HCV. The mutations can be detected using any of the detection techniques described above, such as labeled probes, reverse hybridization assays, southern blots or any other detection technique known to one skilled in the art.

V. Preparation of Pharmaceutical Compositions

Any host, including a human, exhibiting an infection caused by a *Flaviviridae* virus can be treated by administering to the patient an effective amount of a 2'-branched nucleoside or a pharmaceutically acceptable prodrug and/or salt thereof, such as β -D-2'-CH₃-riboC or its 3' valine ester prodrug, in the presence of a pharmaceutically acceptable carrier or dilutant, for any of the indications or modes of administration as described in detail herein in combination or alternation with a drug that induces a mutation in the viral genome at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XXRSGXXXXT, of domain B of the RNA polymerase region. The 2'-branched nucleoside, such as β -D-2'-CH₃-riboC, or a pharmaceutically acceptable prodrug and/or salt thereof can be administered alone or in combination or alternation with other antiviral agents as described herein. The active materials can be administered by any

appropriate route, for example, orally, parenterally, intravenously, intradermally, subcutaneously, or topically, in liquid or solid form.

A preferred dose of a compound will be in the range from about 1 to 50 mg/kg, preferably 1 to 20 mg/kg, of body weight per day, and more generally 0.1 to about 100 mg per kilogram body weight of the recipient per day. The effective dosage range of the pharmaceutically acceptable salts and prodrugs can be calculated based on the weight of the parent nucleoside to be delivered. If the prodrug and/or salt exhibits activity in itself, the effective dosage can be estimated using the weight of the prodrug and/or salt, or by other means known to those skilled in the art.

A compound can be conveniently administered in unit any suitable dosage form, including but not limited to one containing 7 to 3000 mg, preferably 70 to 1400 mg, of active ingredient per unit dosage form. For example, an oral dosage of 50-1000 mg of the active ingredient is usually convenient.

Ideally the active ingredient should be administered to achieve peak plasma concentrations of the active compound of from about 0.2 to 70 μ M, preferably about 1.0 to 10 μ M. This can be achieved, for example, by the intravenous injection of a 0.1 to 5% solution of the active ingredient, optionally in saline, or administered as a bolus of the active ingredient.

The concentration of active compound in the drug composition will depend on absorption, inactivation and excretion rates of the drug as well as other factors known to those of skill in the art. It is to be noted that dosage values will also vary with the severity of the condition to be alleviated. It is to be further understood that for any particular subject, specific dosage regimens should be adjusted over time according to the individual need and the professional judgment of the person administering or supervising the administration of the compositions, and that the concentration ranges set forth herein are exemplary only and are not intended to limit the scope or practice of the claimed composition. The active ingredient can be administered at once, or can be divided into a number of smaller doses to be administered at varying intervals of time.

A preferred mode of administration of the active compound is oral. Oral compositions will generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral

therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition.

The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring. When the dosage unit form is a capsule, it can contain, in addition to material of the above type, a liquid carrier such as a fatty oil. In addition, dosage unit forms can contain various other materials which modify the physical form of the dosage unit, for example, coatings of sugar, shellac, or other enteric agents.

The compound can be administered as a component of an elixir, suspension, syrup, wafer, chewing gum or the like. A syrup can contain, in addition to the active compounds, sucrose as a sweetening agent and certain preservatives, dyes and colorings and flavors.

The compound, or a pharmaceutically acceptable prodrug and/or salt thereof, can also be mixed with other active materials that do not impair the desired action, or with materials that supplement the desired action, such as antibiotics, antifungals, anti-inflammatories, or other antivirals, including other nucleoside compounds. Solutions or suspensions used for parenteral, intradermal, subcutaneous, or topical application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. The parental preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

If administered intravenously, preferred carriers are physiological saline or phosphate buffered saline (PBS).

In a preferred embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation.

Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) are also preferred as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811, which is incorporated herein by reference in its entirety. For example, liposome formulations can be prepared by dissolving appropriate lipid(s), such as stearyl phosphatidyl ethanolamine, stearyl phosphatidyl choline, arachadoyl phosphatidyl choline, and/or cholesterol, in an inorganic solvent that is then evaporated, leaving behind a thin film of dried lipid on the surface of the container. An aqueous solution of the active compound or its monophosphate, diphosphate, and/or triphosphate derivatives is then introduced into the container. The container is swirled by hand to free lipid material from the sides of the container and to disperse lipid aggregates, thereby forming the liposomal suspension.

The active compound(s) are included in the pharmaceutically acceptable carrier or diluent in an amount sufficient to deliver to a patient a therapeutically effective amount of compound to inhibit viral replication *in vivo*, especially *Flaviviridae* replication, without causing serious toxic effects in the treated patient. By "inhibitory amount" is meant an amount of active ingredient sufficient to exert an inhibitory effect as measured by, for example, an assay such as the ones described herein.

Controlled Release Formulations

The field of biodegradable polymers has developed rapidly since the synthesis and biodegradability of polylactic acid was reported by Kulkarni et al., in 1966

(“Polylactic acid for surgical implants,” *Arch. Surg.*, 93:839). Examples of other polymers which have been reported as useful as a matrix material for delivery devices include polyanhydrides, polyesters such as polyglycolides and polylactide-co-glycolides, polyamino acids such as polylysine, polymers and copolymers of polyethylene oxide, acrylic terminated polyethylene oxide, polyamides, polyurethanes, polyorthoesters, polyacrylonitriles, and polyphosphazenes. See, for example, U.S. Patent Nos. 4,891,225 and 4,906,474 to Langer (polyanhydrides), 4,767,628 to Hutchinson (polylactide, polylactide-co-glycolide acid), and 4,530,840 to Tice, et al. (polylactide, polyglycolide, and copolymers). See also U.S. patent No. 5,626,863 to Hubbell, et al which describes photopolymerizable biodegradable hydrogels as tissue contacting materials and controlled release carriers (hydrogels of polymerized and crosslinked macromers comprising hydrophilic oligomers having biodegradable monomeric or oligomeric extensions, which are end capped monomers or oligomers capable of polymerization and crosslinking); and WO 97/05185 to Focal, Inc. directed to multiblock biodegradable hydrogels for use as controlled release agents for drug delivery and tissue treatment agents.

Degradable materials of biological origin, such as crosslinked gelatin, are well known. Hyaluronic acid has been crosslinked and used as a degradable swelling polymer for biomedical applications (U.S. Patent 4,957,744 to Della Valle et. al.; (1991) “Surface modification of polymeric biomaterials for reduced thrombogenicity,” *Polym. Mater. Sci. Eng.*, 62:731-735)).

Many dispersion systems are currently in use, or being explored for use, as carriers of substances, and particularly of biologically active compounds. Dispersion systems used for pharmaceutical and cosmetic formulations can be categorized as either suspensions or emulsions. Suspensions are defined as solid particles ranging in size from a few micrometers up to hundreds of microns, dispersed in a liquid medium using suspending agents. Solid particles include microspheres, microcapsules, and nanospheres. Emulsions are defined as dispersions of one liquid in another, stabilized by an interfacial film of emulsifiers such as surfactants and lipids. Emulsion formulations include water in oil and oil in water emulsions, multiple emulsions, microemulsions, microdroplets, and liposomes. Microdroplets are unilamellar phospholipid vesicles that consist of a spherical lipid layer with an oil phase inside, as defined in U.S. Patent Nos.

4,622,219 and 4,725,442 issued to Haynes. Liposomes are phospholipid vesicles prepared by mixing water-insoluble polar lipids with an aqueous solution. The unfavorable entropy caused by mixing the insoluble lipid in the water produces a highly ordered assembly of concentric closed membranes of phospholipid with entrapped aqueous solution.

U.S. Patent No. 4,938,763 to Dunn, et al., discloses yet another method for drug delivery by forming an implant *in situ* by dissolving a nonreactive, water insoluble thermoplastic polymer in a biocompatible, water soluble solvent to form a liquid, placing the liquid within the body, and allowing the solvent to dissipate to produce a solid implant. The polymer solution can be placed in the body via syringe. The implant can assume the shape of its surrounding cavity. In an alternative embodiment, the implant is formed from reactive, liquid oligomeric polymers which contain no solvent and which cure in place to form solids, usually with the addition of a curing catalyst.

A number of patents disclose drug delivery systems that can be used to administer a 2'-branched nucleoside, or pharmaceutically acceptable prodrug and/or salt thereof, in combination and/or alternation with a drug that induces a mutation in the viral genome at a location other than a mutation of a nucleotide that results in a change from serine to a different amino acid in the highly conserved consensus sequence, XXRSGXXXT, of domain B of the RNA polymerase region. U.S. Patent No. 5,749,847 discloses a method for the delivery of nucleotides into organisms by electrophoration. U.S. Patent No. 5,718,921 discloses the use of microspheres comprising a polymer and drug dispersed therein as a delivery system. U.S. Patent No. 5,629,009 discloses a delivery system for the controlled release of bioactive factors. U.S. Patent No. 5,578,325 discloses the use of nanoparticles and microparticles of non-linear hydrophilic hydrophobic multiblock copolymers for drug delivery. U.S. Patent No. 5,545,409 discloses a delivery system for the controlled release of bioactive factors. U.S. Patent No. 5,494,682 discloses the use of ionically cross-linked polymeric microcapsules as a drug delivery system.

U.S. Patent No. 5,728,402 to Andrx Pharmaceuticals, Inc. describes a controlled release formulation that includes an internal phase that comprises the active drug, its salt or prodrug, in admixture with a hydrogel forming agent, and an external phase which comprises a coating that resists dissolution in the stomach. U.S. Patent Nos. 5,736,159

and 5,558,879 to Andrx Pharmaceuticals, Inc. disclose controlled release formulations for drugs with little water solubility in which a passageway is formed *in situ*. U.S. Patent No. 5,567,441 to Andrx Pharmaceuticals, Inc. discloses a once-a-day controlled release formulation. U.S. Patent No. 5,508,040 discloses a multiparticulate pulsatile drug delivery system. U.S. Patent No. 5,472,708 discloses a pulsatile particle based drug delivery system. U.S. Patent No. 5,458,888 describes a controlled release tablet formulation which can be made using a blend having an internal drug containing phase and an external phase which comprises a polyethylene glycol polymer which has a weight average molecular weight of from 3,000 to 10,000. U.S. Patent No. 5,419,917 discloses methods for the modification of the rate of release of a drug from a hydrogel which is based on the use of an effective amount of a pharmaceutically acceptable ionizable compound that is capable of providing a substantially zero-order release rate of drug from the hydrogel. U.S. Patent No. 5,458,888 discloses a controlled release tablet formulation.

U.S. Patent No. 5,641,745 to Elan Corporation, plc discloses a controlled release pharmaceutical formulation which comprises the active drug in a biodegradable polymer to form microspheres or nanospheres. The biodegradable polymer is suitably poly-D,L-lactide or a blend of poly-D,L-lactide and poly-D,L-lactide-co-glycolide. U.S. Patent No. 5,616,345 to Elan Corporation plc describes a controlled absorption formulation for once a day administration that includes the active compound in association with an organic acid, and a multi-layer membrane surrounding the core and containing a major proportion of a pharmaceutically acceptable film-forming, water insoluble synthetic polymer and a minor proportion of a pharmaceutically acceptable film-forming water soluble synthetic polymer. U.S. Patent No. 5,641,515 discloses a controlled release formulation based on biodegradable nanoparticles. U.S. Patent No. 5,637,320 discloses a controlled absorption formulation for once a day administration. U.S. Patent Nos. 5,580,580 and 5,540,938 are directed to formulations and their use in the treatment of neurological diseases. U.S. Patent No. 5,533,995 is directed to a passive transdermal device with controlled drug delivery. U.S. Patent No. 5,505,962 describes a controlled release pharmaceutical formulation.

The following examples illustrate various embodiments of the invention and are not intended to be limiting in any respect.

EXAMPLES

Example 1:

Isolation of β -D-2'-CH₃-riboC -resistant BVDV

5 Persistent BVDV infection was established in MDBK cell line (ATCC, Manassas, VA, Catalog #: CCL-22) by *in vitro* infection of naive cells with a noncytophatic (ncp) BVDV (strain I-N-dIns; Dr. R. Donis, U. of Nebraska, Lincoln, NE). The multiplicity of infection (MOI) was 0.01. Cells were passaged twice per week (splitting ratio 1:15) until the stable high-level of infection ($10^6 - 10^7$ focus forming units
10 (FFU) per mL) was achieved, as was determined by a focus assay. Next, the persistently infected cells were grown in 6-well culture plates with 8 μ M or without β -D-2'-CH₃-riboC (Idenix Pharmaceuticals). Cell cultures were passaged every three to four days by splitting with a ratio of 1:15 to 1:20. After eight passages, cell cultures grown in the presence of β -D-2'-CH₃-riboC were expanded to T-75 culture flask, freeze/thawed twice,
15 and used as a virus stock of β -D-2'-CH₃-riboC-resistant BVDV for further characterization. The virus titers in cell cultures were monitored at the end of each passage by the virus focus assay.

To conduct the virus focus assay, MDBK cells were seeded onto 6-well plates containing 2×10^5 cells per well and grown at 37 °C/5% CO₂ for at least 5 hours before
20 use. The test samples (culture supernatants combined with cell monolayers) were frozen/thawed twice, serially diluted by 10-fold in medium, and used to inoculate test cells in 6-well plates at 0.2 mL per well. The inoculum was removed after 1 hour of adsorption, and the cells were overlaid with 3 mL of 0.5% agarose in complete growth medium (1X DMEM (Cellgro), supplemented with 8% horse serum, penicillin,
25 streptomycin, L-glutamine, sodium pyruvate, and 25 mM HEPES). After 3 days of incubation at 37°C/5% CO₂, the plates were fixed for 1 hour with 3 mL of 7.4% formaldehyde in PBS, and washed with PBS. The cell monolayers were permeabilized with 1 mL of PBS-0.25% Triton X-100 per well for 10 minutes, and incubated with 0.5

mL of goat anti-BVDV antiserum (VM DR, Inc.; diluted 1:1000 in PBS-0.25% Triton X-100) for 1 hour. The antiserum was then removed, and cell monolayers were washed with PBS (twice for 15 min) and incubated with 0.5 mL of peroxidase-conjugated donkey anti-goat antibody (diluted 1:1000 in PBS-0.25% Triton X-100) for another 1 hour. After the antibody was removed, the cell monolayers were washed with PBS (twice for 15 min), and incubated with 0.5 mL of diaminobenzidine (DAB) peroxidase substrate solution (Vector Laboratories) at room temperature until virus foci become visible (approximately 15 minutes). All incubations were carried out with rocking. The staining was stopped by washing with water and the plates were allowed to air dry. Virus titers were calculated in FFU/mL, using the following equation: $T_{\text{FFU/mL}} = N \times 5 \times D$; where T is a virus titer in FFU/mL; N is a number of foci per well; and D is a dilution factor for the corresponding virus sample. (For example, if 12 foci were found in a well corresponding to 10^{-5} dilution of virus sample, then $T = 12 \times 5 \times 10^5 = 6 \times 10^6$ FFU/mL).

Typically, virus titers reached 10^6 - 10^7 FFU/mL after 2-3 passages, and did not change significantly after further passaging over at least 2 months. When such a persistently infected cell line was treated with 8 μM β -D-2'-CH₃-riboC, the virus titer declined rapidly and the virus was no longer detectable after two passages (**Figure 1**). However, after additional passaging in the presence of the inhibitor, virus reappeared in culture (typically, at passage 3 to 5), and virus titer reached plateau at 10^5 FFU/mL, about ten-fold lower than that of untreated culture (**Figure 1**). This 10-fold difference in virus titers was observed even after 28 days of treatment. This experiment was repeated three times and similar results were obtained. The phenotype of the reappeared virus was remarkably different from the initial wild type virus: it yielded much smaller foci, typically, 3 to 10 times smaller in diameter than those of the wild type virus (**Figure 2**). This phenotype did not change after prolonged passaging in culture in the presence of the inhibitor for at least 72 days, but, it quickly reverted to the wild type phenotype (large foci) after the discontinuation of the treatment.

Taken together, these data demonstrate that the wild type virus disappeared from cell culture after treatment, and the β -D-2'-CH₃-riboC-resistant virus variant demonstrated lesser replication fitness in tissue culture.

Example 2:

Virus growth kinetics

The growth kinetics of both wild type and the β -D-2'-CH₃-riboC-resistant BVDV were compared. MDBK cells were seeded onto 6-well plates (2×10^5 cells per well) and grown at 37°C/5% CO₂ overnight. Cells were infected with BVDV I-N-dIns or the β -D-2'-CH₃-riboC-resistant mutant, I-N-dIns β -D-2'-CH₃-riboC-R, at a multiplicity of infection of 0.1. After a 1-hour adsorption, the inoculum was removed, and cells were washed with PBS and then overlaid with 2 mL of fresh growth medium. For BVDV I-N-dIns β -D-2'-CH₃-riboC-R, duplicate wells were prepared in the presence or absence of 8 μ M β -D-2'-CH₃-riboC. Cell cultures were incubated at 37°C/5% CO₂. At 0 (the end of the adsorption period), 6, 12, 24, 36, 48, 60, 72 hours post-infection, the cultures were frozen/thawed twice, and virus titers were quantified by the focus assay as described above.

At 12 hours post-infection, the wild type virus progeny reached a significant level of over 10^4 FFU/mL, consistent with the BVDV complete life cycle being 8-14 hours. In contrast, the progeny of the resistant virus variant was still undetectable at that point (**Figure 3**). The replication of the resistant virus was first detected at 24 hours post-infection. At 36 hours post-infection, the replication of the resistant virus was still about 100-fold less efficient than that of the wild type virus. These data clearly demonstrate that the β -D-2'-CH₃-riboC-resistant BVDV replicates significantly slower than the wild type virus, especially at the early stages of the infection. These data are also consistent with the results presented in **Figures 1 and 2**.

Example 3

Evaluation of the resistance to β -D-2'-CH₃-riboC

The selected BVDV variant (I-N-dIns β -D-2'-CH₃-riboC-R) is more resistant to β -D-2'-CH₃-riboC than the wild type BVDV since it can stably replicate to a reasonably high levels in MDBK cells in the presence of the compound for a long period of time (at least 72 days) without changes in the phenotype nor in the virus titer levels. To

quantitate this resistance, a virus yield reduction assay was performed, using both the wild type and the variant virus.

To conduct the virus yield reduction assay, MDBK cells were seeded onto 24-well plates (1×10^5 cells per well) and grown at $37^\circ\text{C}/5\% \text{ CO}_2$ overnight. Cells were infected with BVDV at a multiplicity of infection of 0.1. After 1-hour adsorption, the inoculum was removed, and cells were washed with PBS and then overlaid with 1 mL of fresh growth medium, containing serial 2-fold dilutions of the test compound (0 – 32 μM for $\beta\text{-D-2'-CH}_3\text{-riboC}$ and 0-800 IU/mL for Intron®A). After incubation at $37^\circ\text{C}/5\% \text{ CO}_2$ for 48 hours, the plates were frozen/thawed at -70°C twice to lyse the cell cultures. The virus titer in the cell culture was quantified by focus assay as described above. The 50%, 90%, and 4-log effective concentrations (Mean values \pm Standard Deviation) for the test compound were based on duplicate wells. The EC_{50} , EC_{90} and $\text{EC}_{4\text{-log}}$ values were derived by curve fitting using XLFit software. The EC_{50} , EC_{90} , and $\text{EC}_{4\text{-log}}$ are concentrations of test compound, which will reduce viral titer by 50%, 90%, and 99.99%, respectively.

The generation of the infectious wild type BVDV particles was very efficiently inhibited by $\beta\text{-D-2'-CH}_3\text{-riboC}$, with an EC_{50} and EC_{90} values of $0.59 \pm 0.12 \mu\text{M}$ and $1.49 \pm 0.28 \mu\text{M}$, respectively (**Figure 4** and **Table 4**). At $\beta\text{-D-2'-CH}_3\text{-riboC}$ concentration of $7.14 \pm 1.26 \mu\text{M}$ the wild type virus yield was reduced by 4 logs, and at 16 μM virus titers dropped below the detection limit ($<10 \text{ FFU/mL}$). In contrast, no effect on the resistant virus yield was observed at the highest $\beta\text{-D-2'-CH}_3\text{-riboC}$ concentration tested (32 μM). Thus, the I-N-dIns $\beta\text{-D-2'-CH}_3\text{-riboC}$ -R virus was at least 54-fold more resistant to the inhibitor than the wild type virus, based on the EC_{50} values obtained by the virus yield reduction assay ($>32 \mu\text{M}$ versus $0.59 \pm 0.12 \mu\text{M}$).

Table 4: Results of BVDV Yield Reduction Assay

Compound	BVDV strain	Biotype ¹	EC ₅₀ (μM)	EC ₉₀ (μM)	EC _{4 log} (μM)
β-D-2'-CH ₃ -riboC	I-N-dIns	ncp	0.59 ± 0.12	1.49 ± 0.28	7.14 ± 1.26
	I-N-dIns β-D-2'-CH ₃ -riboC-R	ncp	>32	>32	>32
	I-NADL	cp	0.68 ± 0.08	1.73 ± 0.11	8.22 ± 0.05
	I-NADL S3674T	cp	>32	>32	>32
IFN	I-N-dIns	ncp	2.64 ± 1.40	119 ± 34.1	>800
	I-N-dIns β-D-2'-CH ₃ -riboC-R	ncp	0.19 ± 0.04	3.15 ± 0.72	>800

¹ cp = cytopathic; ncp = noncytopathic

Example 4

5 Nucleic acid sequence analysis: Identification of the genetic mutation(s) responsible for the β-D-2'-CH₃-riboC-resistant phenotype.

Based on the nature of the inhibitor, i.e. the nucleoside analog, the viral polymerase was considered as a plausible molecular target. Thus, we begun with the sequencing of the NS5B region for both wild type and the β-D-2'-CH₃-riboC-resistant BVDV. The viral RNA was extracted from the tissue culture lysates after 8 passages of treatment with or without β-D-2'-CH₃-riboC (**Figure 1**), the entire NS5B region was subjected to the RT-PCR and sequencing. Viral RNA was extracted from cell cultures using QIAamp® Viral RNA Mini Kit (QIAGEN) according to the manufacture's protocol. The whole NS5B region was transcribed and amplified using QIAGEN® OneStep RT-PCR Kit. PCR products were purified using QIAquick® PCR Purification Kit (QIAGEN) and sequenced using the ABI PRISM® Sequencing protocol on an automated ABI DNA Sequencer (Perkin-Elmer) at the Tufts Core Facility, Boston, MA.

Each region was sequenced in both directions using at least two independent RT-PCR products. No mutations were found in the wild type virus when compared to the

previously published sequence of the BVDV (strain I-N-dIns) full-length genome (Vassilev, V. B. and R. O. Donis. (2000) *Virus Res.* 69 (2): 95-107). Bovine viral diarrhea virus (BVDV) induced apoptosis correlates with increased intracellular viral RNA accumulation. Only one nucleotide substitution was found in the I-N-dIns β -D-2'-CH₃-riboC-R virus: 1214G to C, changing amino acid residue Ser to Thr at position 405. Interestingly, this amino acid position is located to the putative functional NS5B domain B (**Figure 5**), identified by mutational analysis (Lai V. C., Kao C.C., Ferrari E., Park J., Uss A.S., Wright-Minogue J., Hong Z., and J.Y. Lau. "Mutational analysis of bovine viral diarrhea virus RNA-dependent RNA polymerase" *J Virol.*, **1999**, 73, 10129-36). This domain is also found in the NS5B region of HCV genome, as well as in genomes of other flaviviruses. Moreover, the amino acid position Ser₄₀₅ is highly conserved among all pesti- and flavivirus genomes.

Example 5

Hypersensitivity to Intron A

Comparison of the wild type I-N-dIns virus and I-N-dIns β -D-2'-CH₃-riboC-R variant was conducted for their sensitivity to the Intron A in *de novo*-infected MDBK cells using the virus yield reduction assay, as described above. Again, we found a remarkable difference between the two viruses. The wild type virus was moderately inhibited by the Intron A with an EC₉₀ value of 119 ± 34.1 μ M and an approximate 1.5 log reduction in virus yield at the highest drug concentration tested (**Figure 6**). In contrast, the I-N-dIns β -D-2'-CH₃-riboC-R variant virus was found to be much more sensitive to Intron A, with an EC₉₀ value of 3.15 ± 0.72 μ M and the maximum reduction in the viral yield of nearly 4 log (**Figure 6**). Based on comparison of the EC₉₀ values, the β -D-2'-CH₃-riboC-resistant virus was approximately 40 times more sensitive to the Intron A than the wild type BVDV.

Example 6

Combination treatment with β -D-2'-CH₃-riboC and Intron A

The effect of Intron A, alone or in combination with β -D-2'-CH₃-riboC, on the wild type BVDV was further studied in the persistently infected MDBK cells. In one experimental setting, the virus titers were determined after 7 days (two passages) of single or double treatment with several inhibitor concentrations. The results of this experiment, presented in **Tables 5A** and **5B**, and also in **Figures 7** and **8**, can be summarized as follows. β -D-2'-CH₃-riboC alone strongly inhibited BVDV (strain I-N-dIns) propagation in a dose-dependent manner under described experimental conditions. Treatment with 8 μ M β -D-2'-CH₃-riboC reduced virus titer 6.2 logs (**Figure 7**). Interferon alpha-2b alone had minimal antiviral effect (0.1 log-reduction in virus titers). Single treatment with 2 μ M β -D-2'-CH₃-riboC or 2000 IU/mL interferon alpha-2b reduced viral titers 1.61 logs and 0.1 log, respectively. The effect of a combination treatment with the same concentrations was 2.22 logs, which was 0.51 log higher than the calculated additive effect (1.71 log). Single treatment with 4 μ M β -D-2'-CH₃-riboC or 2000 IU/mL interferon alpha-2b reduced viral titers 2.06 logs and 0.1 log, respectively (**Table 5B**, **Figure 8**). The effect of a combination treatment with same concentrations was 4.56 logs, which was 2.4 logs higher than the calculated additive effect (2.16 log). Thus, β -D-2'-CH₃-riboC and interferon alpha-2b acted synergistically to inhibit BVDV, especially when β -D-2'-CH₃-riboC was used at a concentration of 4 μ M.

Table 5A. Effect of β -D-2'-CH₃-riboC and interferon alpha-2b on BVDV (strain I-N-dIns) titers in persistently infected MDBK cells. Numbers represent BVDV titer values in FFU/mL.

	0 μ M β -D-2'-CH ₃ - riboC	2 μ M β -D-2'-CH ₃ - riboC	4 μ M β -D-2'-CH ₃ - riboC	8 μ M β -D-2'-CH ₃ - riboC
0 IU/mL Interferon alpha-2b	4.03x106 \pm 2.34x106	1.25x105 \pm 3.54x104	3.58x104 \pm 1.06x103	2.50x100 \pm 2.89x100
5 IU/mL Interferon alpha-2b	6.44x106 \pm 3.15x106	2.63x105 \pm 7.42x104	1.00x104 \pm 3.54x103	1.25x100 \pm 2.50x100
50 IU/mL Interferon alpha-2b	8.85x106 \pm 4.53x106	2.13x105 \pm 6.72x104	5.75x102 \pm 4.84x102	0.00x100 \pm 0.00x100

	0 μ M β -D-2'-CH ₃ - rib C	2 μ M β -D-2'-CH ₃ - riboC	4 μ M β -D-2'-CH ₃ - riboC	8 μ M β -D-2'-CH ₃ - riboC
200 IU/mL Interferon alpha-2b	5.38x106 \pm 3.03x106	5.75x104 \pm 1.32x104	2.38x102 \pm 2.06x102	0.00x100 \pm 0.00x100
1000 IU/mL Interferon alpha-2b	2.60x106 \pm 1.14x106	3.93x104 \pm 1.80x104	1.34x102 \pm 2.35x102	0.00x100 \pm 0.00x100
2000 IU/mL Interferon alpha-2b	3.23x106 \pm 1.77x106	2.44x104 \pm 2.07x104	1.12x102 \pm 1.93x102	0.00x100 \pm 0.00x100

Table 5B. Effect of β -D-2'-CH₃-riboC and interferon alpha-2b on BVDV (strain I-N-dlms) titers in persistently infected MDBK cells. Numbers represent log values of the BVDV titers.

	0 μ M β -D-2'-CH ₃ - riboC	2 μ M β -D-2'-CH ₃ - riboC	4 μ M β -D-2'-CH ₃ - riboC	8 μ M β -D-2'-CH ₃ - riboC
0 IU/mL Interferon alpha-2b	6.61	5.10	4.55	0.40
5 IU/mL Interferon alpha-2b	6.81	5.42	4.00	0.40
50 IU/mL Interferon alpha-2b	6.95	5.33	2.76	0.00
200 IU/mL Interferon alpha-2b	6.73	4.76	2.38	0.00
1000 IU/mL Interferon alpha-2b	6.41	4.59	2.13	0.00
2000 IU/mL Interferon alpha-2b	6.51	4.39	2.05	0.00

In another experimental setting, treatment time was extended to 10 days and the viral titers (strain NY-1) were monitored after each passage (every three to four days). Again, similar synergistic inhibitory effect of β -D-2'-CH₃-riboC and interferon alpha-2b was observed (**Figure 9**). Notably, when cell cultures were treated with 8 μ M of β -D-2'-CH₃-riboC in combination with 200 IU/mL of Intron A, the virus became undetectable after 7 days of treatment and did not reappear after further passaging for at least 27 days. These data are in agreement with our previously described finding, which is that β -D-2'-CH₃-riboC-resistant BVDV variant, arising after treatment of the persistently infected cells, is sensitive to the Intron A. Taken together, these data further suggest that the

resistant virus populations, emerging after treatment of the persistent virus infection with β -D-2'-CH₃-riboC, can be eliminated by subsequent treatment with the Intron A.

5 This invention has been described with reference to specific embodiments. Variations and modifications of the invention, will be obvious to those skilled in the art from the foregoing detailed description of the invention.